A comparative study of fluorescent microscopy with Ziehl-Neelsen staining and culture for the diagnosis of pulmonary tuberculosis

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Abstract

Background: For developing countries with a large number of cases and financial constraints, evaluation of rapid and inexpensive diagnostic methods has great importance. The bacilli in the sputum can be detected microscopically by ZN stain and fluorochrome stain.

Objectives: To study the efficacy of fluorescence microscopy in the diagnosis of pulmonary tuberculosis in comparison to Ziehl-Neelsen staining and culture of sputum samples from patients suspected of pulmonary tuberculosis.

Materials and methods: 306 sputum samples collected from 102 patients suspected of pulmonary tuberculosis were processed by the Petroff’s method, and subjected to Ziehl-Neelsen staining (ZN), fluorescent Auramine-O staining (AO) and culture on modified Lowenstein-Jensen media (gold standard) for detection of Mycobacterium tuberculosis. Positive smears were graded according to Forbes BA et al, and culture isolates were biochemically tested for confirmation of species.

Results: Out of 102 patients, 44.1%, 71.6% and 70% were found positive by ZN, AO and culture respectively. AO was found to be superior to ZN on several aspects. The difference in their case detection rates was statistically significant ($\chi^2 = 24.93, p < 0.001$). AO was also able to detect more pauci-bacillary cases than ZN. There was more agreement between culture and fluorescence microscopy (95.1%) than with ZN microscopy (69.6%). The percentage of false negative by AO staining was only 2.78% which was in sharp contrast to that of ZN (40.27%).

Conclusion: The better case detection rates of AO over ZN were comparable to those found by several studies. Since screening was done under lower power of magnification (400x), fluorescence microscopy has been found to be less time consuming as compared to ZN method (1000x) in the diagnosis of tuberculosis. The tubercle bacilli stood out as bright objects against a dark background in fluorescence microscopy which makes them easily identifiable hence causing less eye-strain. The efficacy of fluorescence microscopy proved to be much higher than conventional light microscopy and comparable to that of culture.

Key words: Ziehl-Neelsen staining, Mycobacterium tuberculosis, Auramine-O

Tuberculosis is one of the oldest known diseases of mankind. Yet, the disease is still at large. For developing countries with a large number of cases and financial constraints, evaluation of rapid and inexpensive diagnostic methods like demonstration of acid-fast bacilli (AFB) in smears has great importance.

The bacilli in the sputum can be detected microscopically by ZN stain and fluorochrome stain. In fluorochroming, a direct chemical interaction occurs between the fluorescent dye and a component of the bacterial cell; this interaction is the same as occurs with the stains used in light microscopy1. But, a significant difference in the sensitivity of the two methods has been reported2.

Confirmed diagnosis cannot be made by ESR, Mycodot test and even by ELISA technique. Newer tests, based on advanced technology, such as, PCR, gas liquid chromatography and luciferase activity, are costly and only available in few laboratories, and further, specificity is not 100%. The results are quite variable from centre to centre. The diagnosis of pulmonary tuberculosis by chest X-ray is unreliable.

In this study, the staining methods are compared and evaluated taking culture on modified Lowenstein-Jensen (LJ) medium, as “gold standard”3.
Materials and methods
This comparative study was conducted in the Department of Microbiology, Regional Institute of Medical Sciences, Imphal, Manipur, India, from January 2004 to June 2005, on the sputum specimens of 102 patients suspected of pulmonary tuberculosis.

Patient inclusion and exclusion criteria: Patients attending the Respiratory Diseases Department OPD and the DOTS Centre, RIMS, and having fever, night sweats, cough for more than 3 weeks with sputum, loss of appetite, loss of weight, chest pain, haemoptysis and/or radiological evidence of tuberculosis were included. Those cases who have not taken a course of antibiotics, known cases of carcinoma lung and paediatric cases were excluded. Those unable to produce at least 5 ml of mucopurulent sputum were also excluded.

Sample collection: 3 sputum samples were collected on 2 consecutive days from each patient - spot specimen on the first day, one early morning and one spot specimen on the second day. Samples were collected in clean, sterile, leak-proof, wide-mouth containers.

At the time of sample collection, a Proforma was used to collect data about the patients. On obtaining the study results, the data was completed and analysed statistically using $\chi^2$ and p-value.

The processings of the samples were carried out in a biosafety cabinet. Each sample was processed by the Petroff’s method and subjected to Ziehl-Neelsen (ZN) staining, Fluorescent Auramine-O (AO) staining and culture on modified Lowenstein-Jensen medium.

Smear reporting is done according to Forbes BA et al.1

Acid-fast smear reporting
Smears are examined using a light microscope scanning at least 300 oil immersion fields before reporting a smear as negative.1,5

AFB stain bright pink to red, beaded or barred forms are seen in Mycobacterium tuberculosis while the tissues cells and other organisms are stained blue.

Auramine-o smear reporting
The fluorescence microscope used is the Axioskop 2 plus Transmitted-Light Microscope with an attached Epifluorescence illuminator manufactured by Zeiss and is fitted in a dark room.

The film is examined with a 40x objective and a 10x eye-piece. The tubercle bacilli are seen as yellow luminous organisms in a dark field (Figure 1).

When fluorescent bacilli have been detected the smear was re-stained with ZN stain for confirmation under oil-immersion examination.3

For the present study, 2+, 3+, and 4+ were classified as multibacillary and 1+ as paucibacillary. Doubtful is considered negative.

<table>
<thead>
<tr>
<th>Number of AFB seen Fuchsin stain (1000x magnification)</th>
<th>Number of AFB seen Fluorochrome stain (400x magnification)</th>
<th>Report</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>0</td>
<td>No AFB seen</td>
</tr>
<tr>
<td>1-2/300 fields</td>
<td>1-2/70 fields</td>
<td>Doubtful</td>
</tr>
<tr>
<td>1-9/100 fields</td>
<td>2-18/50 fields</td>
<td>1+</td>
</tr>
<tr>
<td>1-9/10 fields</td>
<td>4-36/10 fields</td>
<td>2+</td>
</tr>
<tr>
<td>1-9/ field</td>
<td>4-36/ field</td>
<td>3+</td>
</tr>
<tr>
<td>&gt;9/ fields</td>
<td>&gt;36/ field</td>
<td>4+</td>
</tr>
</tbody>
</table>

Smear reporting according to Forbes BA et al.

Culture
Modified Lowenstein-Jensen media were prepared in-house. After processing, 0.25ml of the sediment from the decontaminated sputum specimen was inoculated on to the surface of two modified LJ media. After spreading the inoculum over the surface of the slant, the tubes were incubated at 37 °C and left in the slanted position for 7 days to permit even distribution of the inoculum over the entire surface of the medium. The tubes are then placed upright and incubation continued for 8 weeks. The caps of the bottles were loosened once weekly at the time of inspection for growth. A negative report was given if no growth appeared after 8 weeks7.

Observation of colony morphology:
The colonies of M. tuberculosis were rough, raised, thick, with a nodular or wrinkled surface and an irregular thin margin (Fig 2). They were non-pigmented (off-white to faint buff), tenacious and not easily emulsified.
Identification of Mycobacterium tuberculosis:
- Rate of growth.
- Culture growth characteristics.
- Positive Z-N staining.
- Niacin production test - Positive.
- Nitrate reduction test - Positive.

Rapid growers are excluded in this study and no further steps are undertaken to identify them.

Results
Out of 102 clinically diagnosed pulmonary tuberculosis patients, 45, 73 and 72 cases were found to be positive for AFB by ZN staining, AO staining and culture techniques respectively. The ZN smear positivity rate and the AO smear positivity rate in this study was 44.1% (45/102) and 71.6% (73/102) respectively. The combined smear positivity using both the staining techniques was 72.5% (74/102).

Table 1 shows that scores are definitely higher by fluorescence microscopy: 73 (20+28+16+9) positive as against 45 (3+19+16+7) positive by the ZN method. The difference in the case-yields was found to be highly significant ($\chi^2 = 24.93$, $p < 0.001$).

Disregarding the scores, 72 (28+44) of 102 smears gave identical results. In other words, there was 70.6% agreement or 29.4% disagreement between ZN and AO.

Results that were identical are plotted on the diagonal. Scores higher by fluorescence microscopy are shown above the diagonal and those higher by ZN microscopy below it.

In ZN stained smears, 42 (19+16+7) multibacillary and 3 paucibacillary cases were detected whereas in fluorochrome stained smears 53 (28+16+9) multibacillary and 20 paucibacillary cases were detected.

43 out of 72 culture positive cases were diagnosed by ZN stained smear microscopy. 29 cases missed by ZN were detected by culture. There was agreement in 71/102 cases (69.6%) and disagreement in 31/102 (30.4%).

There was agreement in 97/102 cases (95.1%) and disagreement in 5/102 (4.9%) between fluorescence microscopy and culture.

On comparison against culture, the gold standard in the detection of the tubercle bacilli, the sensitivity and the predictive value of negative test of the ZN stain was much lower than those of AO stain (Table 2). The false positive results of the AO stain were slightly higher than that of ZN. The false negative results of the ZN stain were much higher than that of AO. Combining the results of ZN and AO, the efficiency was significantly increased than that of the individual stains.

Table 1: Correlation of conventional bright-field microscopy (Ziehl-Neelsen) and fluorescence microscopy (Auramine-O) gradewise.
Different smear microscopy results were achieved by Jain A et al ZN 32.7%, AO 41.6%, Githui et al ZN 65%, AO 80%, Ulukanligil et al ZN 67.6%, AO 85.7%, Prasanthi K and Kumari AR ZN 50%, AO 69%.3,8,9,10 It was evident that AO method results scored higher than that of ZN method in all these studies as was the case in this study (ZN 44.1%, AO 71.6%). In this study, AO was found to be 27.46% more effective than ZN staining. This shows that fluorochrome staining of sputum smears in comparison to that of ZN staining is a better method of microscopy ($\chi^2=24.93$, $p < 0.001$).

Detection of smear positive cases is the highest priority in any TB control programme, as these cases are infectious and contribute to transmission of disease. Though smear positivity correlates well with infectivity, much of the transmission occurs before the level of bacilli reach $10^5$/ml in the sputum10. ZN stain can detect bacilli when they are in the order of $10^5$/ml of the sputum whereas a more sensitive AO stain can detect in the order of $10^4$/ml of sputum1. That fluorochrome stain is more efficient over ZN stain in detecting paucibacillary cases has been proved in this study. It could detect 20 paucibacillary cases whereas ZN detected only 3 of them. Failure to detect, and hence, to treat paucibacillary cases can be effectively prevented by the use of fluorescence microscopy.

Kumar N et al had observed that 85% of the culture positive cases could be diagnosed by microscopy alone2. In this study, 59.7% (43/72) of the culture positive cases were diagnosed by ZN stained smear light microscopy. The figure increased significantly to 97.2% (70/72) of culture positive cases by AO stained smear fluorescent microscopy.

On evaluation of the microscopic techniques by comparing them with the gold standard culture technique, it was found that in case of ZN stain there was agreement in 69.6% cases and disagreement in 30.4% whereas for AO stain there was agreement in 95.1% cases and disagreement in 4.9% cases. This proves that AO stain is a better method for its close comparability to the gold standard technique.
that of Jain A et al with sensitivity of ZN at 67.3% and AO 86.6%\(^3\). The specificity of AO is quite high at 90% though it is lower than that of ZN at 93.3%. The high specificity of fluorescent microscopy in the diagnosis of tuberculosis has been confirmed by Jain A et al\(^3\).

Jain A et al noted that around 40-50% of active pulmonary TB (culture positive) cannot be detected by ZN microscopy\(^3\). This was in concordance with the false negative results of ZN staining in this study (40.27%). It is in sharp contrast to only 2.78% of false negative in case of AO stain. Hence, it is evident that a large number of cases missed out by ZN stain are easily detected by fluorochrome staining.

**Conclusion**

Sputum examination for the TB bacilli is usually conducted for patients clinically and/or radiologically suspected of pulmonary tuberculosis. However, the standard method of sputum examination, that is, ZN staining is not sensitive enough and a large number of these suspected cases miss diagnosis. Moreover, many cases remain unsuspected and do not seek treatment.

Fluorochrome stain is more efficient over ZN stain in detecting TB bacilli in sputum, especially the paucibacillary cases. Since screening is done under lower power of magnification (400x), fluorescence microscopy has been found to be less time consuming as compared to ZN method (1000x) in the diagnosis of tuberculosis. Hence, it has been advocated to be a method of choice where a large number of sputum smears are to be examined. The fluorescing bacilli are easily identifiable and cause less eye-strain.

Culture examination, no doubt, is more reliable but is time consuming, expensive and requires trained technical hands. The efficacy of fluorescence microscopy proved to be much higher than conventional light microscopy and comparable to that of culture. In this study, combining ZN and AO has been found to significantly improve sensitivity, predictive value of negative test, percentage of false negative and efficiency. Both staining methods are easy and cheap and may be used effectively instead of doing difficult, expensive and time consuming cultures.

**Acknowledgement**

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**References**