

# Role of Peroxisome Proliferator Activated Receptor-gamma and its Ligands in Inflammatory Bowel Disease

Umid Kumar Shrestha,<sup>\*a</sup> Bing Xia,<sup>b</sup>

<sup>a</sup>Department of Internal Medicine, Manipal College of Medical Sciences & Manipal Teaching Hospital, Pokhara, Nepal

<sup>b</sup>Department of Internal Medicine and Gastroenterology, Zhongnan Hospital, Wuhan University School of Medicine, Hubei, PR China

## Date of submission

November 1st, 2011

## Date of acceptance

December 5th, 2011

## Available online

January 25th, 2012

## Keywords

peroxisome proliferator-activated receptor-gamma, inflammatory bowel disease, ulcerative colitis, rosiglitazone

## Citation

Shrestha UK, Xia B. Role of Peroxisome Proliferator Activated Receptor-gamma and its Ligands in Inflammatory Bowel Disease. *Journal of Advances in Internal Medicine*. 2012;01(1)33-38.

## ABSTRACT

Peroxisome proliferator-activated receptor-gamma (PPAR- $\gamma$ ), a nuclear receptor, is highly expressed in the colonic epithelium in contrast to its impaired expression in the patients with ulcerative colitis (UC). Several natural and synthetic ligands of PPAR- $\gamma$  with some effects in the colon have been identified. The aim of this review is to provide an overview of the role of PPAR- $\gamma$  and its ligands in inflammatory bowel disease (IBD). Review of article was done using a PubMed search. Animal model studies have revealed that PPAR- $\gamma$  is the key receptor for 5-aminosalicylic acid that mediates its main effects in the colon. Moreover, the clinical trials have shown that the PPAR- $\gamma$  agonist rosiglitazone is effective in the treatment of mild to moderately active UC. PPAR- $\gamma$  gene therapy, used as an adjunct intervention, may be effective in suppressing inflammation in colitis. Some commensal bacteria and natural ligands present in food may induce PPAR- $\gamma$  expression and activation in the colon which suggest the possibility of associating a natural regulator and a synthetic ligand of PPAR- $\gamma$  as drug therapy for IBD patients. Further studies are required for the development of unique and effective therapies with PPAR- $\gamma$  agonists in IBD patients.

## INTRODUCTION

Current advances suggest that an inappropriate response of a defective mucosal immune system to the indigenous intestinal flora and other luminal antigens in a genetically susceptible host is at the core of the pathophysiology of inflammatory bowel disease (IBD), which includes ulcerative colitis (UC) and Crohn's disease (CD).<sup>1,2</sup> Because of the regulatory action in the colon and involvement in immune response, peroxisome proliferator-activated receptor-gamma (PPAR- $\gamma$ ) has become a hot research topic in gastroenterology. After a brief description of PPAR- $\gamma$ , this review aims to provide an overview of the role of PPAR- $\gamma$  with the latest findings about the use of its ligands in IBD. A computerised medical literature search of all English language articles was done from the "PubMed" online database with the keywords "peroxisome proliferator activated receptor-gamma", "inflammatory bowel disease", "Crohn's disease", "ulcerative colitis", "colitis", "PPAR gamma ligands" and "rosiglitazone".

### PPAR- $\gamma$ structure and its expression in different tissues including colon

The peroxisome proliferator-activated receptors (PPAR) are nuclear receptors, which are intracellular transcription factors that regulate the activity of complex gene networks.<sup>3</sup> The PPAR subfamily of nuclear hormone receptors include distinct genes that code for several PPAR isoforms denoted: PPAR $\alpha$ ,  $\beta/\delta$  and  $\gamma$ .<sup>4</sup> The human PPAR- $\gamma$  gene is composed of nine exons spanning more than 100 kb of genomic DNA5 on chromosome 3p25 in the proximity of the locus for the retinoic acid receptor RAR- $\beta$  (3p24) and the thyroid

receptor TR- $\beta$  (3p21).<sup>6,7</sup> The PPAR- $\gamma$  was initially identified for its role in adipocyte differentiation and regulation of genes involved in lipid and glucose metabolism. However, activation of PPAR- $\gamma$  also can antagonize nuclear factor  $\kappa$ B (NF $\kappa$ B) action in macrophages resulting in downregulation of proinflammatory cytokines.<sup>8-15</sup>

The expression of PPAR- $\gamma$  is found not only in adipocytes but also in a number of other cells types, such as macrophages, lymphocytes, hepatocytes, and skeletal muscle.<sup>16</sup> Very high expression levels of PPAR- $\gamma$  are also found in the colonic epithelium.<sup>17</sup> In both rodents and humans, the level of receptor in colon tissue is equal to, or greater than, that in adipose tissue. In addition, studies have reported higher expression levels of PPAR- $\gamma$  in the distal colon than in the proximal colon and small intestine.<sup>18</sup> Moreover, PPAR- $\gamma$  expression is primarily localized in the more differentiated epithelial cells of the colon.<sup>19,20</sup> The localization in differentiated cells is consistent with the numerous reports of PPAR- $\gamma$  induction upon differentiation of cultured colon cells.<sup>19, 21-23</sup> Thus, the expression and activation of PPAR- $\gamma$  are associated with a differentiated phenotype in intestinal cells.

### \*Corresponding author

Department of Internal Medicine, Manipal College of Medical Sciences and Manipal Teaching Hospital, Phulbari-11, Pokhara, Kaski, Nepal  
Email address - umidshrestha@gmail.com

The expression of PPAR- $\gamma$  by epithelial cells could be regulated by bacteria, which might explain the characteristic and important PPAR- $\gamma$  pattern expression in the colon compared with other parts of the digestive tract.<sup>24</sup> It is noteworthy to mention that lipopolysaccharide (LPS) of Gram negative bacteria seems to be critical in colonic steady state PPAR- $\gamma$  expression through Toll-like receptor (TLR).<sup>4,24</sup>

### IMPACT OF MUTATIONS IN PPAR- $\gamma$ GENE

The polymorphism in the PPAR- $\gamma$ 2 gene (Pro12Ala), which is a CCA-to-GCA missense mutation in codon 12 of exon B of the PPAR $\gamma$  gene, was recently identified.<sup>25</sup> This substitution possibly results in a conformational change in protein structure and reduced function of the PPAR $\gamma$  gene. At the cellular level, reduced binding of the Ala variant to the PPAR $\gamma$ -responsive DNA elements and reduced transcription of specific genes in cells overexpressing the Ala variant have been reported.<sup>26,27</sup> The Ala allele of the common Pro12Ala polymorphism is associated with a reduced risk of type 2 diabetes.<sup>28</sup> This polymorphism also appears to have a protective effect against diabetic nephropathy.<sup>29</sup> Individuals with the Ala allele are also found to have a reduced risk of colorectal cancer.<sup>30-32</sup> Another single nucleotide polymorphism (SNP) in the PPAR- $\gamma$  gene (C161T) is a silent C to T substitution in nucleotide 161 of exon 6 and does not cause an amino acid change.<sup>33</sup> The C161T polymorphism has been correlated with the colorectal cancer,<sup>34</sup> colorectal adenoma<sup>35</sup> and with other conditions such as psoriatic arthritis,<sup>36</sup> diabetic nephropathy,<sup>37</sup> plasma leptin levels in obese subjects,<sup>33</sup> extent of coronary artery disease by angiography,<sup>38</sup> carotid intima media thickness,<sup>60</sup> and incidence of myocardial infarction among individuals younger than age.<sup>29</sup>

### Association of PPAR gamma polymorphism with IBD

Different studies have shown the variable results about the association of polymorphism of PPAR- $\gamma$  with IBD. The significant association between PPAR- $\gamma$  polymorphisms and the development of CD and UC at single loci level and also in haplotype combinations, was shown in a Hungarian study, suggesting a potential protective effect of the Ala allele in IBD.<sup>39</sup> In the study done in Chinese population, the potential association was found between the PPAR- $\gamma$  C161T polymorphism and UC patients, but the finding was not replicated in the Dutch population.<sup>40</sup> In one recent Danish study, a statistically significant (although modest) association was determined between the homozygous PPAR- $\gamma$  Pro12Ala variant genotype and an increased risk of IBD.<sup>41</sup> However, in one study from Japan, Plo12Ala polymorphism of PPAR- $\gamma$  was not found to be associated with the risk of developing UC.<sup>42</sup> Similarly, another study from Turkey also showed that Pro12Ala polymorphism in the PPAR- $\gamma$  gene was not related to the risk of the development of inflammatory bowel disease.<sup>43</sup> The inconsistent results of different studies demand for the more studies with larger representative samples of IBD to find the concrete data about the polymorphism of PPAR- $\gamma$  in IBD patients.

### ROLE OF PPAR- $\gamma$ IN IBD

The strongest evidence for an anti-inflammatory role of PPAR- $\gamma$  comes from the studies which indicated that heterozygous PPAR- $\gamma$  deficient mice were more susceptible to dextran sodium sulphate (DSS) and 2,4,6-trinitrobenzene sulfonic acid (TNBS) induced

colitis.<sup>44</sup> DSS induced colitis, in particular, is an acute inflammation model primarily driven by epithelial disruption and macrophage infiltration. The data indicates that PPAR- $\gamma$  expression in certain cell types of the colon plays an anti-inflammatory role. Recent studies elaborated on these findings by showing that mice deficient in PPAR- $\gamma$  expression in epithelial cells and macrophages displayed increased pro-inflammatory gene expression and susceptibility to DSS colitis.<sup>45,46</sup> These findings suggest that PPAR- $\gamma$  expression in at least two cell types, epithelial and macrophage, can protect against at least one model of acute colitis (DSS). Experimental models of colitis can also be initiated by distinct mechanisms and driven by infiltration of different cell types, both epithelial and immune cells.<sup>47-49</sup> However, the importance of tissue specific PPAR- $\gamma$  expression may depend largely on the model of colitis examined, emphasizing the need to utilize multiple models to accurately represent manifestations of human colitis. Despite evidence for anti-inflammatory actions of the RXR/ PPAR- $\gamma$  heterodimer in the colon in animal models, the role of PPAR- $\gamma$  in IBD in humans is little explored.

The study done in the UC patients showed an impaired expression of PPAR- $\gamma$  at the mRNA and protein levels.<sup>50</sup> This study also revealed the comparable levels of PPAR- $\gamma$  in peripheral mononuclear cells of IBD patients and controls and absence of specific mutations of the PPAR- $\gamma$  gene or its promoter in UC patients which suggest that epigenetic events may account for impaired PPAR- $\gamma$  expression in UC patients.<sup>50</sup>

The study was done to test the relationship between PPAR- $\gamma$  alleles and CD in humans, which was based on SAMP1/YitFc animal findings developing spontaneous ileitis due to a defect in expression of PPAR- $\gamma$  in ileal crypts, secondary to inheritance of AKR alleles in the region of PPAR- $\gamma$ .<sup>51</sup> The study showed that two intronic polymorphisms SNP1 ( $p < 10^{-5}$ ) and SNP2 ( $p \leq 10^{-3}$ ) exhibited lower allele frequencies in 134 CD patients compared with 125 controls.<sup>51</sup>

### PPAR- $\gamma$ LIGANDS IN IBD

#### Natural PPAR- $\gamma$ ligands

Naturally occurring substances such as polyunsaturated fatty acids, certain eicosanoids, and 15deoxy- $\Delta$ 12,14-PGJ2 (15d-PGJ2) have been found to be the weak activators of PPAR- $\gamma$ .<sup>52</sup> They have intrinsically low binding affinities and weak in vivo concentrations in intestinal cells; hence, many of these compounds do not support physiological functions. The minimal concentrations of 15d-PGJ2 required to activate PPAR- $\gamma$  are approximately 10–150-fold higher than those found in human intestinal epithelial cells.<sup>53</sup> Studies have shown that conjugated linoleic acids protected mice from experimental colitis by the activation of PPAR- $\gamma$ .<sup>54</sup> This effect was not seen in mice with colonic knockout of PPAR- $\gamma$ . Since the food derived bacterial metabolites are the main source of linoleic acids in the gut, the food supplements might have positive effect on intestinal inflammation mediated via PPAR- $\gamma$ .<sup>55</sup> This PPAR- $\gamma$  could play an important role in the homeostasis of intestinal microflora and the epithelial barrier. In normal mucosa, PPAR- $\gamma$  in intestinal epithelial cells could recognize luminal bacterial metabolites and then set the threshold of NF $\kappa$ B activity as one of the most important proinflammatory transcription factors. The unsaturated fatty acid derivative nitrolinoleic acid (LNO2), generated via nitric oxide dependent oxidative inflammatory reactions, has been identified as a

new PPAR- $\gamma$  agonist.<sup>56</sup> Present in the vascular cell wall as the most abundant bioactive oxide of nitrogen and in the blood of healthy individuals at concentrations of approximately 500 nM, LNO2 is considered at present to be one of the most potent physiological endogenous natural ligand of PPAR- $\gamma$ .<sup>57</sup> Further studies are needed to determine intestinal effects of LNO2 in the maintenance of gut homeostasis and during inflammatory disorders.

### Synthetic PPAR- $\gamma$ ligands

Thiazolidinediones (TZDs), such as troglitazone, rosiglitazone and pioglitazone, are high affinity synthetic ligands of PPAR- $\gamma$ , frequently referred to as "PPAR- $\gamma$  agonists".<sup>58</sup> TZDs are currently used as insulin sensitizing agents in the treatment of type 2 diabetes mellitus.<sup>57</sup> Glitazar is a novel family of dual acting PPAR- $\alpha/\gamma$  agonist developed as an oral treatment for insulin resistance related glucose and lipid abnormalities associated with type 2 diabetes and the metabolic syndrome.<sup>59</sup> Non-steroidal anti-inflammatory drugs are also reported *in vitro* as PPAR- $\gamma$  ligands but *in vivo* their binding affinities of 0.1 mM are 1000-fold higher than the mean concentrations found in patients conventionally treated with these drugs.<sup>60</sup>

5-Aminosalicylic acid (5-ASA) is an anti-inflammatory drug widely used in the treatment of IBD. It is known to inhibit the production of cytokines and inflammatory mediators, but the mechanism underlying the intestinal effects of 5-ASA remained unknown. The study showed that PPAR- $\gamma$  is the key receptor for 5-ASA that mediates its main effects in the colon.<sup>61</sup> A small-sample open-label study showed that the patients with mild to moderately active UC refractory to the standard therapies may benefit from therapy with PPAR ligands.<sup>62</sup> Another study showed that the combined treatment with rosiglitazone and 5-ASA had better therapeutic effect than 5-ASA alone in mild to moderately active UC.<sup>63</sup> Encouraging results were reported in yet another multi-center, randomized, double-blind, placebo-controlled clinical trial, which showed that rosiglitazone was effective in the treatment of mild to moderately active UC patients.<sup>64</sup> However, there is considerable concern regarding whether the adverse effects of thiazolidinediones would outweigh the potential benefit for patients with UC.

The reports were published referring about the greater risk of myocardial infarction estimated with the last updated myocardial event rates as an odds ratio of 1.29 (95% CI, 1.01–1.66,  $P = 0.05$ ).<sup>65-67</sup> The cardiovascular adverse events occurred more frequently in subgroups of susceptible patients treated with rosiglitazone for at least 24 weeks,<sup>68</sup> having type 2 diabetes, long-term nitrate use, and/or concurrent insulin therapy.<sup>69</sup> The mechanism for the

apparent increase in the rosiglitazone-induced myocardial infarction rate remains unknown but is not regarded as a PPAR- $\gamma$  ligand effect because the other thiazolidinedione pioglitazone widely used to treat type 2 diabetes has significant protective effects on coronary and peripheral vascular events<sup>70</sup>; pioglitazone does not increase the risk for myocardial infarction and may decrease the risk for stroke and revascularization.<sup>71</sup> However, while treating the UC patients with thiazolidinediones, it is better not to use such therapy in patients with concomitant diseases such as liver disease, congestive heart failure, or in those at particularly high risk for myocardial infarction.

### PPAR- $\gamma$ GENE THERAPY IN IBD

In order to enhance the limited therapeutic efficacy of synthetic PPAR- $\gamma$  ligands in established colitis, the study was done with the PPAR- $\gamma$  gene therapy as an adjunct intervention and has shown that the gene therapy alone also was effective in suppressing inflammation and attributed this finding to the action of endogenous agonists.<sup>72</sup>

### CONCLUSION

Although the current data supports a role for PPAR- $\gamma$  expression and activation in epithelial and immune cell types in the control of colonic inflammation, given that only few studies are available so far about the PPAR- $\gamma$  in IBD, more studies are still necessary to confirm the understanding of the mechanism of the anti-inflammatory actions of PPAR- $\gamma$ , understanding of factors affecting thiazolidinedione efficacy and understanding of the adverse effects of short and long term use of thiazolidinediones in IBD. The recent discovery that some commensal bacteria and natural ligands present in food may induce PPAR- $\gamma$  expression and activation in the colon suggest about the potential of associating a natural regulator and a synthetic ligand of PPAR- $\gamma$  as drug therapy for IBD patients. The report of 5-ASA as a new synthetic ligand of PPAR- $\gamma$  has encouraged the researchers to develop new drugs, similar to 5-ASA but with a more topical effect in the gut, a stronger affinity to PPAR- $\gamma$  and minimal adverse effects, so that they could be used more effectively in the induction and maintenance treatment of UC in the future. PPAR- $\gamma$  gene therapy may be a promising adjunct therapy in suppressing the inflammation in the patients of UC. However, more animal and clinical studies of PPAR- $\gamma$  in IBD are needed for the development of unique and effective therapies for IBD patients.

 JAIM

## REFERENCES

1. Baumgart DC, Carding SR. Inflammatory bowel disease: cause and immunobiology. *Lancet*. 2007;369:1627-40.
2. Shanahan F. Inflammatory bowel disease: immunodiagnos- tics, immunotherapeutics, and eotherapeutics. *Gastroenterol- ogy*. 2001;120:622-35.
3. Bain DL, Heneghan AF, Connaghan-Jones KD, et al. Nuclear receptor structure: implications for function. *Annu Rev Physiol*. 2007;69:201-20.
4. Schoonjans K, Staels B, Auwerx J. The peroxisome prolif- erator-activated receptors (PPARs), and their effects on lipid metabolism, and adipocyte differentiation. *Biochem Biophys Acta*. 1996;1302:93-109.
5. Fajas L, Auboeuf D, Raspe E, et al. The organization, pro- moter analysis and expression of the human PPARgamma gene. *J Biol Chem*. 1997;272:18779-89.
6. Greene ME, Blumberg B, McBride OW, et al. Isolation of the human peroxisome proliferator activated receptor gamma cDNA: expression in hematopoietic cells and chromosomal mapping. *Gene Expression*. 1995;4:281-99.
7. Beamer BA, Negri C, Yen CJ, et al. Chromosomal localiza- tion and partial genomic structure of the human peroxisome proliferator activated receptorgamma (hPPAR gamma) gene. *Biochem Biophys Res Commun*. 1997;233:756-9.
8. Chinetti G, Fruchart JC, Staels B. Peroxisome proliferator-ac- tivated receptors (PPARs): nuclear receptors at the crossroads between lipid metabolism and inflammation. *Inflamm Res*. 2000;49:497-505.
9. Jiang C, Ting AT, Seed B. PPAR-gamma agonists inhibit production of monocyte inflammatory cytokines. *Nature*. 1998;391:82-6.
10. Wang P, Anderson PO, Chen S, Paulsson KM, Sjögren HO, Li S. Inhibition of the transcription factors AP-1 and NF-kappaB in CD4 T cells by peroxisome proliferator-activated receptor gamma ligands. *Int Immunopharmacol*. 2001;1:803-12.
11. Panzer U, Schneider A, Guan Y, et al. Effects of different PPAR gamma-agonists on MCP-1 expression and monocyte recruitment in experimental glomerulonephritis. *Kidney Int*. 2002;62:455-64.
12. Delerive P, Fruchart JC, Staels B. Peroxisome proliferator- activated receptors in inflammation control. *J Endocrinol*. 2001;169:453-9.
13. Ricote M, Huang JT, Welch JS, Glass CK. The peroxisome proliferator-activated receptor (PPARgamma) as a regulator of monocyte/ macrophage function. *J Leukoc Biol*. 1999;66:733-9.
14. Welch JS, Ricote M, Akiyama TE, Gonzalez FJ, Glass CK. PPARgamma and PPARdelta negatively regulate specific subsets of lipopolysaccharide and IFNgamma target genes in macrophages. *Proc Natl Acad Sci U S A*. 2003;100:6712-7.
15. Cunard R, Ricote M, DiCampli D, et al. Regulation of cytokine expression by ligands of peroxisome proliferator activated receptors. *J Immunol*. 2002;168:2795-802.
16. Ricote M, Li AC, Willson TM, Kelly CJ, Glass CK. The peroxi- some proliferatoractivated receptor-gamma is a negative regu- lator of macrophage activation. *Nature*. 1998;391:79-82.
17. Su CG, Wen X, Bailey ST, et al. A novel therapy for colitis utilizing PPARgamma ligands to inhibit the epithelial inflam- matory response. *J Clin Invest*. 1999;104:383-9.
18. Letebvre AM, Paulweber B, Fajas L, et al. Peroxisome prolif- erator-activated receptor gamma is induced during differentia- tion of colon epithelium cells. *J Endocrinol*. 1999;162:331-40.
19. Mansen A, Guardiola-Diaz H, Rafter J, Branting C, Gustaf- son JA. Expression of the peroxisome proliferator-activated receptor (PPAR) in the mouse colonic mucosa. *Biochem Biophys Res Commun*. 1996;222:844-51.
20. Brockman JA, Gupta RA, Dubois RN. Activation of PPARgam- ma leads to inhibition of anchorage-independent growth of hu- man colorectal cancer cells. *Gastroenterology*. 1998;115:1049- 55.
21. Kitamura S, Miyazaki Y, Shinomura Y, Kondo S, Kanayama S, Matsuzawa Y. Peroxisome proliferator-activated receptor gamma induces growth arrest and differentiation markers of human colon cancer cells. *Jpn J Cancer Res*. 1999;90:75-80.
22. Wachtershauser A, Loitsch SM, Stein J. PPAR-gamma is selectively upregulated in Caco-2 cells by butyrate. *Biochem Biophys Res Commun*. 2000;272:380-5.
23. Huin C, Schohn H, Hatier R, et al. Expression of peroxi- some proliferatoractivated receptors alpha and gamma in differentiating human colon carcinoma Caco-2 cells. *Biol Cell*. 2002;94:15-27.
24. Dubuquoy L, Rousseaux C, Thuru X, et al. PPARgamma as a new therapeutic target in inflammatory bowel diseases. *Gut*. 2006;55:1341-9.
25. Yen CJ, Beamer BA, Negri C, et al. Molecular scanning of the human peroxisome proliferator activated receptor gamma (hPPAR gamma) gene in diabetic Caucasians: identification of a Pro12Ala PPAR gamma 2 missense mutation. *Biochem Biophys Res Commun*. 1997;241:270-4.
26. Deeb SS, Fajas L, Nemoto M, et al. A Pro12Ala substitution in PPARgamma2 associated with decreased receptor activity, lower body mass index and improved insulin sensitivity. *Nat Genet*. 1998;20:284-7.
27. Masugi J, Tamori Y, Mori H, Koike T, Kasuga M. Inhibi- tory effect of a proline-to-alanine substitution at codon 12 of peroxisome proliferator-activated receptorgamma 2 on thiazolidinedione-induced adipogenesis. *Biochem Biophys Res Commun*. 2000;268:178-82.
28. Altshuler D, Hirschhorn JN, Klannemark M, et al. The com- mon PPARgamma Pro12Ala polymorphism is associated with decreased risk of type 2 diabetes. *Nat Genet*. 2000;26:76-80.
29. Herrmann SM, Ringel J, Wang JG, Staessen JA, Brand E. Peroxisomeproliferator-activated receptor-gamma2 polymor- phism Pro12Ala is associated with nephropathy in type 2 dia- betes: The Berlin Diabetes Mellitus (BeDiaM) Study. *Diabetes*. 2002;51:2653-7.
30. Landi S, Moreno V, Gioia-Patricola L, et al. Association of common polymorphisms in inflammatory genes interleukin (IL)6, IL8, tumor necrosis factor alpha, NFkB1, and peroxisome proliferator-activated receptor gamma with colorectal cancer. *Cancer Res*. 2003;63:3560-6.
31. Murtaugh MA, Ma KN, Caan BJ, et al. Interactions of peroxi-

- some proliferator activated receptor  $\{\gamma\}$  and diet in etiology of colorectal cancer. *Cancer Epidemiol Biomarkers Prev*. 2005;14:1224-9.
32. Koh WP, Yuan JM, Van Den Berg D, Ingles SA, Yu MC. Peroxisome proliferator-activated receptor (PPAR) gamma gene polymorphisms and colorectal cancer risk among Chinese in Singapore. *Carcinogenesis*. 2006;27:1797-802.
33. Meirhaeghe A, Fajas L, Helbecque N, et al. A genetic polymorphism of the peroxisome proliferator-activated receptor gamma gene influences plasma leptin levels in obese humans. *Hum Mol Genet*. 1998;7:435-40.
34. Jiang J, Gajalakshmi V, Wang J, et al. Influence of the C161T but not Pro12Ala polymorphism in the peroxisome proliferator-activated receptor-gamma on colorectal cancer in an Indian population. *Cancer Sci*. 2005;96:507-12.
35. Siezen CL, van Leeuwen AI, Kram NR, Luken ME, van Kranen HJ, Kampman E. Colorectal adenoma risk is modified by the interplay between polymorphisms in arachidonic acid pathway genes and fish consumption. *Carcinogenesis*. 2005;26:449-57.
36. Butt C, Gladman D, Rahman P. PPAR-gamma gene polymorphisms and psoriatic arthritis. *J Rheumatol*. 2006;33:1631-3.
37. Szeto CC, Chow KM, Poon PY, Kwan BC, Li PK. Peroxisome proliferator activated receptor-gamma gene polymorphism and risk of cardiovascular disease in patients with diabetic nephropathy. *Am J Nephrol*. 2008;28:715-22.
38. Wang XL, Oosterhof J, Duarte N. Peroxisome proliferator-activated receptor gamma C161-T polymorphism and coronary artery disease. *Cardiovasc Res*. 1999;44:588-94.
39. Poliska S, Penyige A, Lakatos PL et al. Association of Peroxisome Proliferator-activated Receptor gamma Polymorphisms with Inflammatory Bowel Disease in a Hungarian Cohort. *Inflamm Bowel Dis* 2011 (early view; published online on 24 June 2011; available at <http://onlinelibrary.wiley.com/doi/10.1002/ibd.21798/full>)
40. Shrestha UK, Karimi O, Crusius J, Bart A, et al. Distribution of Peroxisome Proliferator-Activated Receptor-Gamma Polymorphisms in Chinese and Dutch Patients with Inflammatory Bowel Disease. *Inflamm Bowel Dis*. 2010;16:312-319.
41. Andersen V, Christensen J, Ernst A, et al. Polymorphisms in NF- $\kappa$ B, PXR, LXR, PPAR $\gamma$  and risk of inflammatory bowel disease. *World J Gastroenterol*. 2011;17:197-206
42. Wang F, Tahara T, Arisawa T, et al. Polymorphism of peroxisome proliferator-activated receptor gamma is not associated to Japanese ulcerative colitis. *Hepatology*. 2008;55:73-5.
43. Atug O, Tahan V, Eren F et al. Pro12Ala Polymorphism in the Peroxisome Proliferator-Activated Receptor-gamma (PPAR $\gamma$ ) Gene in Inflammatory Bowel Disease. *J Gastrointest Liver Dis*. 2008;17:433-7.
44. Desreumaux P, Dubuquoy L, Nutten S, et al. Attenuation of colon inflammation through activators of the retinoid X receptor (RXR)/peroxisome proliferator activated receptor gamma (PPAR $\gamma$ ) heterodimer: A basis for new therapeutic strategies. *J Exp Med*. 2001;193:827-38.
45. Adachi M, Kurotani R, Morimura K, et al. Peroxisome proliferator-activated receptor gamma in colonic epithelial cells protects against experimental inflammatory bowel disease. *Gut*. 2006;55:1104-13.
46. Shah YM, Morimura K, and Gonzalez FJ. Expression of peroxisome proliferator activated receptor-gamma in macrophage suppresses experimentally induced colitis. *Am J Physiol Gastrointest Liver Physiol*. 2007;292:G657-66.
47. Byrne FR and Viney JL. Mouse models of inflammatory bowel disease. *Curr Opin Drug Discov Devel*. 2006;9:207-17.
48. Wirtz S, Neurath MF. Mouse models of inflammatory bowel disease. *Adv Drug Deliv Rev*. 2007;59:1073-83.
49. Strober W, Fuss IJ, Blumberg RS. The immunology of mucosal models of inflammation. *Annu Rev Immunol*. 2002;20:495-549.
50. Dubuquoy L, Jansson EA, Deeb S, et al. Impaired expression of peroxisome proliferator-activated receptor gamma in ulcerative colitis. *Gastroenterology*. 2003;124:1265-76.
51. Sugawara K, Olson TS, Moskaluk CA, et al. Linkage to peroxisome proliferator activated receptor-gamma in SAMP1/YitFc mice and in human Crohn's disease. *Gastroenterology*. 2005;128:351-60.
52. Schoonjans K, Martin G, Staels B, Auwerx J. Peroxisome proliferator-activated receptors, orphans with ligands and functions. *Curr Opin Lipidol*. 1997;8:159-66.
53. FitzGerald GA, Loll P. COX in a crystal ball: current status and future promise of prostaglandin research. *J Clin Invest*. 2001;107:1335-7.
54. Bassaganya-Riera J, Reynolds K, Martino-Catt S, et al. Activation of PPAR gamma and delta by conjugated linoleic acid mediates protection from experimental inflammatory bowel disease. *Gastroenterology*. 2004;127:777-91.
55. Greicius G, Arulampalam V, Pettersson S. A CLA's Act: feeding away inflammation. *Gastroenterology*. 2004;127:994-6.
56. Schopfer FJ, Lin Y, Baker PR, et al. Nitrolinoleic acid: an endogenous peroxisome proliferator-activated receptor gamma ligand. *Proc Natl Acad Sci U S A*. 2005;102:2340-5.
57. Martens FM, Visseren FL, Lemay J, de Koning EJ, Rabelink TJ. Metabolic and additional vascular effects of thiazolidinediones. *Drugs*. 2002;62:1463-80.
58. Berger J, Wagner JA. Physiological and therapeutic roles of peroxisome proliferator-activated receptors. *Diabetes Technol Ther*. 2002;4:163-74.
59. Lohray BB, Lohray VB, Bajji AC, et al. (-)-3-[4-[2-(Phenoxy-10-yl)ethoxy]phenyl]-2-ethoxypropanoic acid [(-)-DRF 2725]: a dual PPAR agonist with potent antihyperglycemic and lipid modulating activity. *J Med Chem*. 2001;44:2675-8.
60. Lehmann JM, Lenhard JM, Oliver BB, Ringold GM, Kliewer SA. Peroxisome proliferator-activated receptors alpha and gamma are activated by indomethacin and other nonsteroidal anti-inflammatory drugs. *J Biol Chem*. 1997;272:3406-10.
61. Rousseaux C, Lefebvre B, Dubuquoy L, et al. Intestinal anti-inflammatory effect of 5-aminosalicylic acid is dependent on peroxisome proliferator-activated receptor-gamma. *J Exp Med*. 2005;201:1205-15.
62. Lewis JD, Lichtenstein GR, Stein RB, et al. An open-label trial of the PPAR $\gamma$  ligand rosiglitazone for active ulcerative

tive colitis. *Am J Gastroenterol.* 2001;96:3323–8.

63. Liang HL, Ouyang Q. A clinical trial of combined use of rosiglitazone and 5-aminosalicylate for ulcerative colitis. *World J Gastroenterol.* 2008;14:114–9.

64. Lewis JD, Lichtenstein GR, Deren JJ, et al. Rosiglitazone for active ulcerative colitis; a randomized placebo-controlled trial. *Gastroenterology.* 2008;134:688–95.

65. Home PD, Pocock SJ, Beck-Nielsen H, et al. RECORD Study Group. Rosiglitazone evaluated for cardiovascular outcomes—an interim analysis. *N Engl J Med.* 2007;357:28–38.

66. Krall RL. Cardiovascular safety of rosiglitazone. *Lancet.* 2007;369:1995–6.

67. Dahabreh IJ, Economopoulos K. Meta-analysis of rare events: an update and sensitivity analysis of cardiovascular events in randomized trials of rosiglitazone. *Clin Trials.* 2008;5:116–20.

68. Nissen SE, Wolski K. Effect of rosiglitazone on the risk of myocardial infarction and death from cardiovascular causes. *N Engl J Med.* 2007; 356:2457–71.

69. Rosen CJ. The rosiglitazone story—lessons from an FDA

Advisory Committee meeting. *N Engl J Med.* 2007;357:844–6.

70. Dormandy JA, Charbonnel B, Eckland DJ, et al; PROactive investigators. Secondary prevention of macrovascular events in patients with type 2 diabetes in the PROactive Study (PROspective pioglitazone Clinical Trial In macroVascular Events): A randomised controlled trial. *Lancet.* 2005;366:1279–89.

71. Nagajothi N, Adigopula S, Balamuthusamy S, et al. Pioglitazone and the risk of myocardial infarction and other major adverse cardiac events: a meta-analysis of randomized, controlled trials. *Am J Ther.* 2008;15:506–11.

72. Katayama K, Wada K, Nakajima A, et al. A novel PPAR gamma gene therapy to control inflammation associated with inflammatory bowel disease in a murine model. *Gastroenterology.* 2003;124:1315–24.