

# In vitro anti-bacterial activity of *Tinospora cordifolia* leaf extract and its phytochemical screening

Prajwala B<sup>1</sup>, Priyanka S<sup>2</sup>, Raghu N<sup>3</sup>, Gopenath N<sup>4</sup>, Gnanasekaran A<sup>5</sup>, Karthikeyan M<sup>6</sup>, Indumathi R<sup>7</sup>, Ebrahim NK<sup>8</sup>, Pugazhandhi B<sup>9</sup>, Pradeep P<sup>10</sup>, MS Ranjith<sup>11</sup>, Balasubramanian S<sup>12</sup>, Kanthesh M Basalingappa<sup>13\*</sup>

## \*Corresponding author:

Dr. Kanthesh M Basalingappa, Ph.D., Assistant Professor, Division of Molecular Biology, Faculty of Life Science, JSS Academy of Higher Education & Research, SS, Nagara Mysore -570015

Email: [kantheshmb@jssuni.edu.in](mailto:kantheshmb@jssuni.edu.in) [ORCID](#)

## Information about the article:

**Received:** Nov. 10, 2018

**Accepted:** Jan. 12, 2019

**Published online:** April 15, 2019

## Cite this article:

Prajwala B, Priyanka S, Raghu N, Gopenath N, Gnanasekaran A, Karthikeyan M *et al.* In vitro anti-bacterial activity of *Tinospora cordifolia* leaf extract and its phytochemical screening. Journal of Biomedical Sciences. 2018;5(2):10-17

## Publisher

Nepal Health Research Society, Bahundhara -6, Gokarnesow Municipality, Kathmandu, Nepal  
eISSN 2382-5545

© The Author(s). 2018

Content licensing: [CC BY 4.0](#)

## ABSTRACT

### Background

Antimicrobial resistance is the main concern worldwide to combat infectious. Over the years studies on leaf extracts *Tinospora cordifolia* have demonstrated the potent role its antibacterial property. The current study is an attempt to test its antibacterial property against *Escherichia coli* cell division.

### Material and methods

Phytochemical screening assay of *T. cordifolia* leaf extract was done using standard procedure and the results showed the presence of alkaloid, carbohydrate, terpenoid, steroid, tannin, amino acid, flavonoid and glycoside components.

### Results

HPLC analysis revealed the presence of berberine in *T. cordifolia* leaf extract. Further *E. coli* cells were treated with berberine to study its efficacy in inhibiting cell division. Antibacterial assay was performed by using disc diffusion method.

### Conclusion

Among aqueous, methanolic, ethanolic, chloroform, hexane and acetone extract only methanolic extract showed zone of inhibition.

### Keywords

*T. cordifolia*, Berberine, HPLC, *E. coli*

## Introduction

Microorganisms, like any other life form, tend to evolve over time. Survival is the key for existence and it must adapt to its surrounding. Any interference to their survival must be overcome. The use of antimicrobials, even when used under appropriate conditions, can lead to antimicrobial resistance let alone its indiscriminate usage resulting in medication ineffectiveness and allowing the infections persist in the body for a longer duration. Resistant strains of microbes continue to proliferate and poses major risk factor for severe infections.

Plant and plant based antimicrobial agents were used against the microbial infection for ages all over the countries. Plant-derived medicine is the major antimicrobial agents for 60 to 80% of populations in the developing countries [1]. Traditionally, crude plant extracts are used as herbal medicine for the treatment of human infectious diseases. Natural compounds present in medicinal plants enhance the immune status and maintain disease free state [2]. Because of this medicinal products research gains important role in clinical research.

*T. cordifolia* is one of the most widely used herb in Ayurveda, one of the ancient systems of medicine practiced in India. It belongs to Menispermaceae family and commonly called heart leaved moonseed plant in English, Guduchi in Sanskrit, Giloe in Hindi, Teppatige in Telugu [3]. It has large, deciduous shrub with twinning branches, leaves are simple, alternate and long petiole. Flowers are unisexual and small; fruits are drupes, turning red when ripe. Fruits aggregate of *T. cordifolia* belongs to different classes of alkaloids, steroids, glycosides, diterpenoid lactones, sesquiterpenoids, phenolics, aliphatic compound [4-5]. These compounds are present in all parts of *T. cordifolia* but its concentration is more in root, stem and leaves. Berberine and furanolactone is the main compound of the plant [6]. The plant is of great interest to researchers because of its medicinal properties like anti-bacterial, anti-fungal, anti-cancer, anti-periodic, anti-spasmodic, anti-stress, anti-inflammatory, anti-arthritis, anti-neoplastic, anti-diabetic activity [7]. All parts of the plant are reported to be useful in ethnobotanical survey conducted by ethnobotanist [8]. Medicinal properties of *T. cordifolia* is presently applied in modern medicine and used to overcome the adverse effect of chemotherapy [9]. The plant is also used in the treatment of jaundice, arthritis, urinary tract infections (UTI) etc. and is rich in micronutrients such as calcium, phosphorous, copper, manganese, zinc and iron [3]. Roots, stem and leaves of *T. cordifolia* has no toxic effect on human body when orally consumed [2]. In our present study preliminary phytochemical screening and anti-microbial activity of different extract of leaves of *T. cordifolia* were evaluated using standard procedure.

## Material and methods

### Collection of plant material

Fresh plant leaves were collected from Nursery Horticulture Department, Mysore, authenticated by Department of Botany, Maharani's College, and Mysore. Leaves were washed in running tap water, shade dried for 15 days.

### Solvent extraction

Dried leaves were crushed and finely powdered using pestle and mortar. Leaf powder was extracted with aqueous, methanol, ethanol, chloroform, hexane and acetone by Maceration method [10]. 20gms of well grinded leaf powder was dissolved in 200ml of solvent. The obtained solution was filtered using filter paper and the final filtrate was evaporated to dryness at room temperature. Scraped out the content and kept it at 4° C in order to minimize the possibility of contamination and to maintain its bioactivity for longer period.

### I. Phytochemical screening

Various tests were conducted to confirm the presence of phytochemical in *T. cordifolia* leaf extract such as alkaloid, carbohydrate, terpenoid, steroid, saponin, tannin, amino acid, flavonoid, glycoside and protein. Phytochemical analysis was carried out according to following standard procedures.

#### i. Amino acid:

Ninhydrin Test: 2 drops of ninhydrin solution with 2 ml of extract, boil, and appearance of blue color indicate the presence of amino acid [9].

#### ii. Alkaloid:

Wagner's test: For 1 ml sample solution adds few drops of Wagner's reagent; reddish brown precipitate will form.

#### iii. Carbohydrate Test:

Fehling's test: For 1 ml Fehling A and 1 ml Fehling B reagents add 1 ml of extract boiled (10min). Formation of brick red color precipitate indicates the presence of carbohydrate [11].

#### iv. Terpenoid test:

Salkowski's test: 1 ml of crude extract with 1ml of chloroform, evaporated to dryness, adds 2ml of concentrated H<sub>2</sub>SO<sub>4</sub> and heated for about 2 minutes. Grayish color will appear.

#### v. Saponin test:

Froth test: 2 ml of crude extract with 2ml of distilled water in a test tube, shaken vigorously (warm); formation of stable foam indicates the presence of saponin.

#### vi. Glycoside test:

Liebermann's test: For 2 ml of the extract add 2 ml of chloroform and 2 ml of acetic acid, solution cooled well in ice, add Sulphuric acid carefully color changes into green.

**Table 1: Qualitative phytochemical analysis of *Tinospora cordifolia* leaf extract**

Phytochemicals	Alkaloid	carbohydrate	Terpenoid	steroid	Saponin	Tannin	Amino acid	Flavonoid	Glycoside	Protein
Sample										
Aqueous	+	-	-	+	-	-	+	-	-	+
Methanol	+	-	+	+	-	+	-	-	+	+
Ethanol	+	-	-	+	-	+	-	-	+	+
Chloroform	+	+	-	+	-	-	-	-	-	+
Hexane	+	-	+	-	-	+	-	-	+	+
Acetone	+	-	+	-	-	+	-	+	+	+

+ve Present, -ve Absent

#### vii. Protein test:

Millon's Test: 3 ml extract and 5 ml of Millon's reagent, heated. Color changes from white precipitate to brick red on heating [12].

#### viii. Steroid test:

Salkowski's test: 2ml of crude extract dissolved in 2ml of chloroform add 2ml of con. H<sub>2</sub>SO<sub>4</sub> sidewise, red color ring is produced [13].

#### ix. Tannin test:

FeCl<sub>3</sub> test: 2ml of extract dissolved in 2ml of distilled water (stirred), add few drops of FeCl<sub>3</sub>, green precipitate will form [14].

#### x. Flavonoid test:

FeCl<sub>3</sub> test: 1ml of extract dissolved with few drops of ferric chloride, green color will form [15-18].

### II. Qualitative analysis of HPLC

Qualitative analysis of HPLC was carried out to check presence of berberine in aqueous, methanol, ethanol, chloroform, hexane and acetone extract. Analysis was carried out using Shimadzu HPLC, C-18 column with a mobile phase of orthophosphoric acid at flow rate of 1ml/min with UV detector at 345nm.

### III. Quantitative analysis of HPLC

Berberine standard was purchased from Sigma-Aldrich, Bangalore and it was run against aqueous, methanol, ethanol and acetone extract. Qualitative analysis of HPLC was carried out using C-18 column with a mobile phase of acetonitrile at flow rate of 1ml/min with UV detector at 345nm.

### IV. Anti-bacterial activity

Antibacterial activity of *T. cordifolia* leaf extract was carried out using disc diffusion method against *Escherichia coli*. Aqueous, methanolic, ethanolic, chloroform, hexane and acetone extracts were used as test compound and ampicillin was used as reference standard. Initially *E. coli* was cultured in nutrient broth for 24 hours at 37°C. Agar plate was prepared by adding 25ml of nutrient agar media into the sterile petri plate, allowed to solidify. 10µl of bacteria was

inoculated and streaked on agar plate using cotton swabs. Disc containing different concentration of *T. cordifolia* leaf extract were placed on agar plates. Plates were incubated overnight at 37°C. Zone of inhibition was measured after 24 hours using ruler and compared with standard ampicillin [19].

### Results

Phytochemicals are chemicals produced by plants through primary or secondary metabolism. Preliminary phytochemical screening assay of *T. cordifolia* leaf extract was conducted to check the presence of phytochemicals. *T. cordifolia* leaf extract showed positive test result which indicates the presence of alkaloid, carbohydrate, terpenoid, steroid, tannin, amino acid, flavonoid, glycoside and protein. The test for alkaloid and protein has given positive result and saponin shown negative result in all the six extract. Carbohydrates were present only in chloroform extract, amino acid in aqueous extract and flavonoid in acetone extract. Methanol and acetone extract showed highest number of phytochemicals.

**Table 2: Qualitative HPLC analysis of *Tinospora cordifolia* leaf extract**

Sample	Retention time
Aqueous	3.609
Ethanol	3.582
Methanol	3.580
Chloroform	Nil
Hexane	Nil
Acetone	3.577

Qualitative analysis of HPLC was carried out to check the presence of berberine in all the six extract. Results revealed the presence of berberine in aqueous, ethanol, methanol and acetone extract and absence of berberine in chloroform and hexane extract. Based on retention time berberine peak was identified. Retention time 3.5 corresponded to berberine [14].

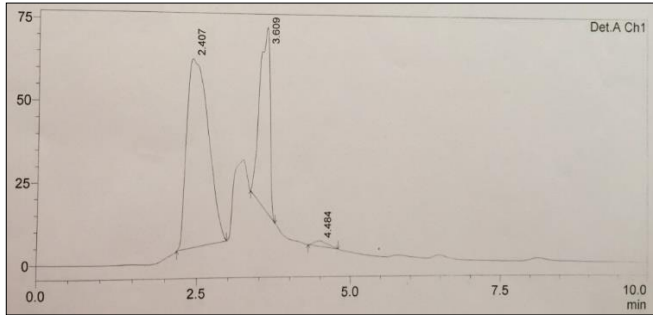


Figure 1: HPLC analysis of aqueous extract of *T. cordifolia*

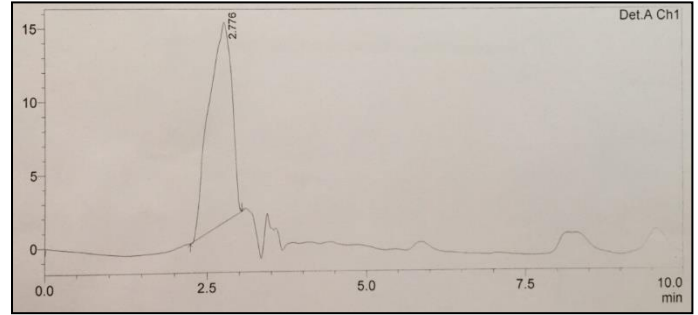


Figure 5: HPLC analysis of hexane extract of *T. cordifolia*

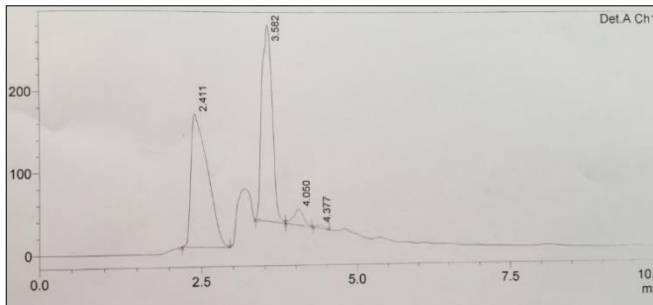


Figure 2: HPLC analysis of methanolic extract of *T. cordifolia*

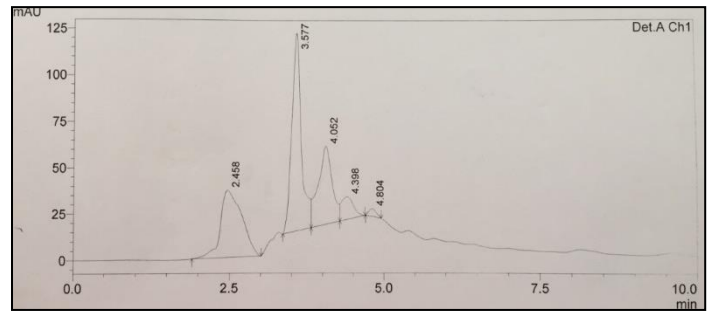


Figure 6: HPLC analysis of acetone extract of *T. cordifolia*

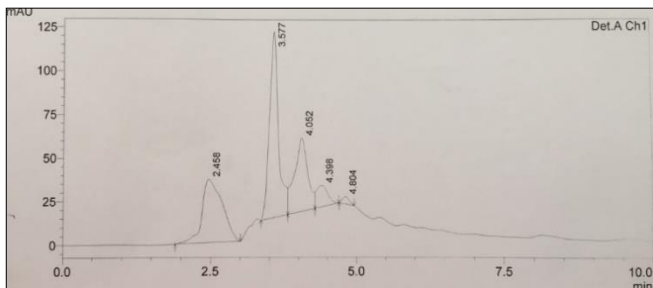


Figure 3: HPLC analysis of ethanolic extract of *T. cordifolia*

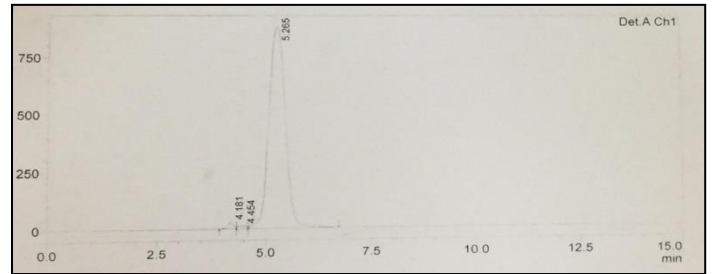


Figure 7: HPLC analysis of Berberine standard (The peak at retention time 5.26 corresponding to Berberine)

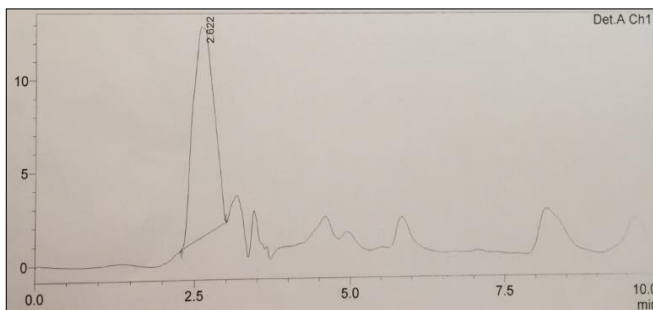


Figure 4: HPLC analysis of chloroform extract of *T. cordifolia*

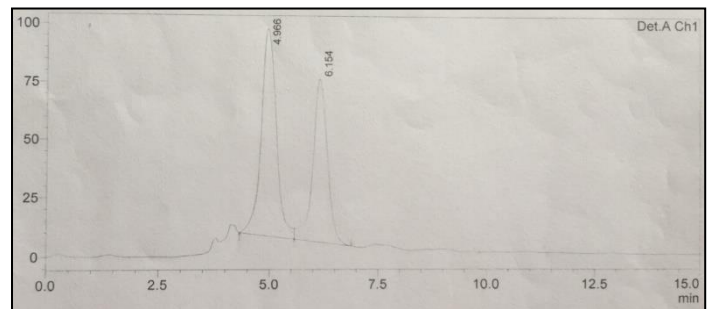


Figure 8: HPLC analysis of methanolic extract of *T. cordifolia* (Analysis showed well separated berberine peak at retention time 4.906)

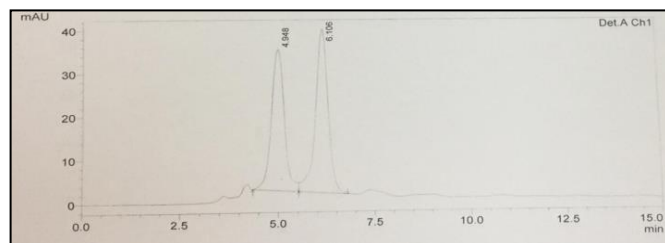


Figure 9: HPLC analysis of ethanolic extract of *T. cordifolia* (HPLC analysis of ethanolic extract showed well separated berberine peak at retention time 4.948)

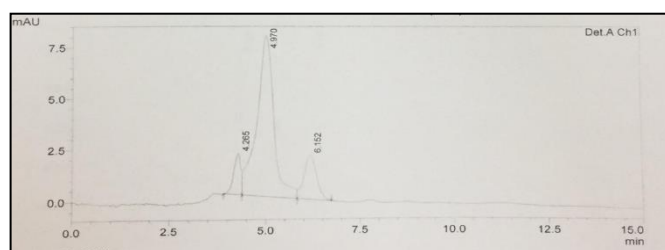


Figure 10: HPLC analysis of aqueous extract of *T. cordifolia* (HPLC analysis of aqueous showed sharp and well resolved berberine peak at retention time of 4.970)

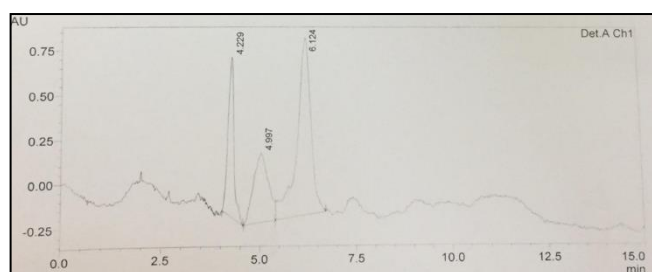


Figure 11: HPLC analysis of acetone extract of *T. cordifolia* (HPLC analysis of acetone extract showed sharp and well separated berberine peak at retention time 6.124)

**Table 3: Quantitative analysis of HPLC**

Sample	Concentration (%)
Aqueous	0.2
Ethanol	0.5
Methanol	1.1
Acetone	0.1

Quantitative analysis of HPLC was carried out to know the concentration of berberine in aqueous, ethanol, methanol and acetone extract along with berberine standard. In qualitative analysis chloroform and hexane extract showed the absence of berberine, hence chloroform and hexane extract was not taken in quantitative analysis. Based on the retention time of berberine standard, berberine peak was identified in remaining chromatogram. In quantitative analysis methanolic extract showed 1.1% of berberine.

**Table 4: Anti-bacterial activity**

Test organism	Test compound	Concentration(µl/disc)	Zone of inhibition	
<i>Escherichia coli</i>	Methanol	10µl	0.3mm	
		20µl	0.5mm	
		30µl	0.8mm	
	Aqueous	10µl, 20µl, 30µl	Nil	
		Ethanol	10µl, 20µl, 30µl	Nil
			Chloroform	10µl, 20µl, 30µl
	Hexane	10µl, 20µl, 30µl		Nil
		Acetone	10µl, 20µl, 30µl	Nil
	Ampicillin		10µl	1.6mm
		20µl	1.6mm	
30µl		1.6mm		

*T. cordifolia* leaf extract was used as test compound to inhibit the growth of *E. coli* with Ampicillin as reference. Methanolic extract showed zone of inhibition or inhibitory effect on growth of *E. coli*. Compared to the remaining extracts which may be attributed to chemical constituents responsible for antibacterial activity are more soluble in methanolic extract.

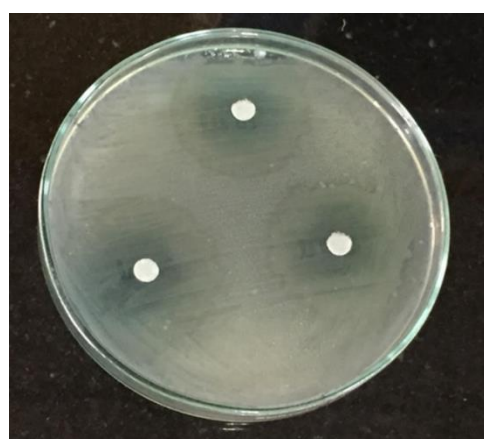


Figure 12: Inhibition of growth of *E. coli* by Ampicillin

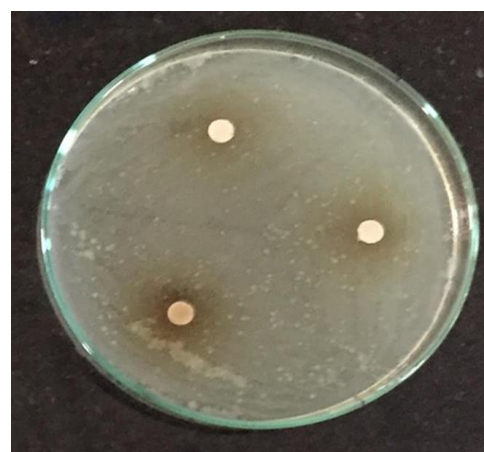


Figure13: Inhibition of growth of *E. coli* by methanolic extract

## Discussion

Phytochemicals are biologically active compound found in plants. Naturally they help plants to defend against microbes and these important properties very useful for future drug discovery [20-22]. In this study, preliminary phytochemical screening assay was conducted by using aqueous, methanolic, ethanolic, chloroform, hexane and acetone extract using standard procedure. Result revealed the presence of alkaloid, carbohydrate, terpenoid, steroid, saponin, tannin, amino acid, flavonoid, glycoside and protein. Methanol and acetone extract showed more number of phytochemical. Preliminary phytochemical screening assay proved that *T. cordifolia* leaf extract is rich in primary and secondary constituents. Qualitative analysis of HPLC was carried out using aqueous, methanol, ethanol, chloroform, hexane and acetone extract. Aqueous, methanol, ethanol and acetone extract showed presence of berberine [23]. Chloroform and hexane extract did not show berberine peak. Quantitative analysis of HPLC was carried out with aqueous, methanol, ethanol and acetone extract. Aqueous extract contains 0.2%, methanol extract contains 1.1%, ethanol extract contains 0.5% and acetone extract contains 0.1% of berberine [24]. Methanol extract contains more concentration of berberine compare to other extracts. Anti-bacterial assay was performed with aqueous, methanolic, ethanolic, chloroform, hexane and acetone extract [25-29]. Among that methanol extract showed anti-bacterial activity. Remaining extracts did not show much inhibitory activity. The result obtained from this experiment is just a rough estimate in finding the medicinal property in terms of antibacterial activity of the plant.

## Conclusion

The present study indicates *T. cordifolia* leaf extract was rich in primary and secondary constituents. Most of the biologically active phytochemicals were present in methanolic extract. In HPLC analysis berberine was present in aqueous, methanolic, ethanolic and acetone extract its concentration was more in methanolic extract. In anti-bacterial activity methanolic extract was effective against *E. coli*. Since the methanolic extract showed positive result in every assay, the present results can be considered beneficial for further investigation.

## Abbreviations

urinary tract infections (UTI)

## Acknowledgments

The authors would like to appreciate the JSS AHER, Mysuru, for their cooperation throughout the study.

## Authors' contribution

Conceptualization, data curation was done by PB, PS, RN and BMK., formal analysis, manuscript writing, and editing

was done by all authors. Final manuscript was approved by all authors.

## Competing interests

The authors declare no conflicts of interest.

## Publisher's Note

NHRS remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

The publisher shall not be legally responsible for any types of loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

## Author information

<sup>1</sup>Miss Prajwala, Research Scholar

<sup>2</sup>Priyanka S, Research Scholar

<sup>3</sup>Raghu Nataraj, Assistant Professor

<sup>4</sup>Dr. Gopenath N, Associate Professor, Ph.D.

<sup>5</sup>Dr. Ashok Gnanasekaran, Ph.D. Associate professor, Microbiology

<sup>6</sup>Dr. Murugesan Karthikeyan, Ph.D., Senior lecturer, Microbiology

<sup>7</sup>Indumathi Rajaraman, M.Sc., M.Phil, PhD scholar,

<sup>8</sup>Dr. Ebrahim Nangarath Kottakal Cheriya, Ph.D., Senior lecturer, Physiology

<sup>9</sup>Mr. Pugazhandhi Bakthavatchalam, M.Sc., Senior lecturer, Anatomy

<sup>10</sup>Dr. Pradeep Palanisamy, MD, Senior lecturer, Anatomy

<sup>11</sup>Prof. Dr. Ranjith Mehenderkar, Ph.D., Professor, Microbiology

<sup>12</sup>Balasubramanian S, Director Research, JSS AHER, Mysuru.

<sup>13</sup>Dr. Kanthesh M Basalingappa, Ph.D. Assistant Professor, Molecular Biology

<sup>1-3,13</sup>Division of Molecular Biology, Faculty of Life Science, JSS Academy of Higher Education & Research, Mysore.

<sup>4</sup>Division of Biotechnology, Faculty of Life Science, JSS Academy of Higher Education & Research, Mysore.

<sup>5-11</sup>Faculty of Medicine, Quest International University Perak, No. 227, Plaza Teh Teng Seng (level 2), Jalan Raja Permaisuri Bainun, 30250 Ipoh, Perak Darul Ridzuan, Malaysia

<sup>12</sup>Director of Research, JSS AHER, SS Nagara, Mysuru-570015

## References

- Ojiako C. Herbal medicine: Yesterday, today and tomorrow. *Alternative & Integrative medicine*, 2015; 4(3):1-5. DOI: <https://doi.org/10.4172/2327-5162.1000195>

2. Mittal J, Sharma MM. *Tinospora cordifolia*: a multipurpose medicinal plant-A review. Journal of Medicinal Plant Studies, 2014; 2(2):32-47.
3. Gupta BM, Ahmed KKM, Gupta R. Global research on *T. cordifolia* (Medicinal plant) with special reference to India. A scientometric assessment publication output during 2001-2016. International Journal of Pharmacognacy and Chinese Medicine, 2018;2(4):000141.
4. Upadhyay AK. *T. cordifolia* (Wild.) Hook.f. and Thoms (Guduchi)-Validation of the Ayurvedic pharmacology through experimental and clinical studies. Int J Ayurveda Res. 2010;1(2):112-21. DOI: <https://doi.org/10.4103/0974-7788.64405>.
5. Joshi BC, Uniyal S. Pharmaco-gnostical review of *T. cordifolia*. Inventi. Rapid: Planta Activa. 2017(1):1-10.
6. Kumar VD, Geethanjali B, Avinash KO, Chandrashekrappa GK, Kanthesh M Basalingappa. *Tinospora cordifolia*: the antimicrobial property of the leaves of amruthaballi. Journal of Bacteriology & Mycology, 2017;5(5):363-71. DOI: <https://doi.org/10.15406/jbmoa.2017.05.00147>
7. Sohamsaha. *T. cordifolia*: One plant many roles. Anc Sci Life. 2012; 31(4): 151–159. DOI: <https://doi.org/10.4103/0257-7941.107344>
8. Choudhary N, Siddiqui MB, Khatoon S. Pharmacognostic evaluation of *T. cordifolia* (Wild.) Miers and identification of biomarkers. Indian Journal of Traditional Knowledge. 2014;13(3):543-50.
9. Tiwari P, Kumar B, Kaur M, Kaur G, Kaur H. Phytochemical screening and extraction: A review. Internationale Pharmaceutica Scientia. 2011;1(1):98-106.
10. Kamble N, Puranik DB, Salooja MK. Preliminary phyto-chemical analysis of aqueous extracts of leaves and stem of *T. cordifolia*. International Journal of Engineering Technology Science and Research. 2017; 4(12):592-6.
11. Joseph BS, Kumbhare PH, Kale MC. Preliminary phytochemical screening of selected medicinal plants. Int. Res. J. of Science & Engineering, 2013;1(2):55-62.
12. RNS Yadav. Phytochemical analysis of some medicinal plants. Journal of Phytology 2011, 3(12):10-14.
13. Singh KL, Bag G. Phytochemical analysis and determination of total phenolics contents in water extracts of three species of *Hedychium*. International Journal of Pharm Tech Research, 2013; 5(4):1516-21.
14. Kumar ABS, Kumar JR, Karthikeyan M, Gnanasekaran A, Akshay V, Reddy V *et al.* Preliminary phytochemical analysis of methanolic extract of *T. cordifolia* and its antibacterial action on *E. coli* cell division. Hygeia.J.D.Med, 2017;9(1): 52-60. DOI: <https://doi.org/10.15254/H.J.D.Med.9.2017.16>
15. Bhandary SK, Kumari NS, Bhat VS, Sharmila KP, Bekal MP. Preliminary phytochemical screening of various extracts of *Punicagranatum* peels whole fruit and seed. Nitte University Journal of Health and Science, 2012; 2(4):34-38.
16. Abebe H, Gebre T, Haile A. Phytochemical investigation on the roots of *Solanum Incanum*, Hadiya zone, Ethiopia. Journal of medicinal plants studies, 2014; 2(2):83-93.
17. Yadav R, Khare RK, Singhal A. Qualitative phytochemical screening of some selected medicinal plants of Shivpur District. Int. J. Life. Sci. Scientifi. Res, 2017;3(1):844-7. DOI: <https://doi.org/10.21276/ijlssr.2017.3.1.16>
18. Shanthi K, Sengottuvel R. Qualitative and quantitative phytochemical analysis of *Moringa concanensis* Nimbo. Int. J. Curr. Microbiol. App. Sci, 2016;5(1):633-40. DOI: <http://dx.doi.org/10.20546/ijemas.2016.501.064>
19. Bargah RK. Preliminary test of phytochemical screening of crude ethanolic and aqueous extract of *Moringapterygosperra Gaertn.* Journal of pharmacognacy and phytochemistry 2015;4(1):7-9.
20. Banu KS, Cathrine L. General techniques involved in phytochemical analysis. International Journal of Advanced Research in Chemical Sciences, 2015;2(4):25-32.
21. Aziz S, Saha K, Sultana N, Ahmed S, Al-Mansur A. Phytochemical and elemental screening on leaves and flowers of *Catharanthus roseus*: An important medicinal plant of Bangladesh. Int.J.Chem.Sci., 2014;12(4):1328-36.
22. Bansal D, Bhasin P, Punia A, Sehrawat AR. Evaluation of antimicrobial activity and phytochemical screening of extracts of *T. cordifolia* against some pathogenic microbes. Journal of pharmacy research 2012;5(1):127-9.
23. Valgas C. Screening methods to determine antibacterial activity of natural products. Braz.J.Microbiol. 2007; 38(2):369-80. DOI: <https://dx.doi.org/10.1590/S1517-83822007000200034>
24. Balouri M, Sadiki M, Ibsouda KS. Methods for In vitro evaluating antimicrobial activity: A

- review. Journal of Pharmaceutical Analysis, 2016;6(2):71-9.
25. Bhalodia NR, Shukla VJ. Antibacterial and antifungal activities from leaf extracts of *Cassia fistula*. An ethnomedicinal plant. J Adv Pharm Technol Res. 2011 Apr-Jun; 2(2): 104-9.  
DOI: <https://doi.org/10.4103/2231-4040.82956>
26. Sen A, Batra A. Evaluation of antimicrobial activity of different solvent extracts of medicinal plant: *Melia Azedarach*. International Journal of Current Pharmaceutical Research, 2012; 4(2):67-73.
27. Boligon AA, Athayde ML. Importance of HPLC in analysis of plants extracts. Austin Chromatogram. 2014;1(3):2.
28. Shah MD, Hossain MA. Total flavonoids content and biochemical screening of the leaves of tropical endemic medicinal plant *Merremiaborneesis*. Arabian Journal of Chemistry, 2014;7(6):1034-8.
29. Karimi A, Majlesi M, Kopaei MR. Herbal versus synthetic drugs; beliefs and facts. J Nephropharmacol. 2015;4(1):27-30.