Leishmania Donovani Promastigote: Effect of Adrenergic Agonists and Antagonist on Growth and Lipophosphoglycan Synthesis

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ABSTRACT

Background: Leishmania is the causative agent of a spectrum of diseases in human ranging in severity from self-healing cutaneous form to disfiguring mucocutaneous lesion to deadly visceral form of leishmaniasis. The treatment is still suffering from toxicity of the present drug regimen and the emergence of drug resistant leishmaniasis. Successful immunotherapy is yet to develop. It is beneficial to venture the parasite second messenger systems for the development of successful antileishmanial agents. The role of adrenergic receptor agonists and antagonist in the growth of L. donovani promastigotes and the synthesis of phosphorylated macromolecules were addressed in this report. Materials and Methods: The present work is entirely experimental. The effect of dibutyrly cAMP (db-cAMP), dibutyrly cGMP (db-cGMP), epinephrine, isoproterenol and propranolol on the growth of L. donovani promastigotes using defined medium were studied. Biosynthetically pulse labeling of promastigotes with [32P]-orthophosphate with or without experimental agents following SDS-PAGE and autoradiography was analyzed. Results: The growth of L. donovani promastigotes at 96 h culture was inhibited by 69%, 83, 52% and 95% with db-cAMP, epinephrine, isoproterenol and propranolol at 100 μM each, respectively in compared to control. Multiplication of parasite was stimulated by db-cGMP. The expression of lipophosphoglycan of the parasite was drastically affected in the following order with propranolol>epinephrine>cAMP>isoproterenol at 100 μM of each agent for 6 h exposure to the promastigotes. Conclusion: The growth of the parasite and the synthesis of lipophosphoglycan were significantly more inhibited by epinephrine or propranolol in a dose dependent manner than cAMP or isoproterenol.

Keywords: cAMP; cGMP; epinephrine; Leishmania donovani; LPG; propranolol.

INTRODUCTION

Leishmania donovani is a digenetic hemoflagellate, alternating between a flagellated promastigote form in the hostile midgut of the vector phlebotomine insect and a rudimentary flagellated obligatory intracellular amastigote form in macrophages causing visceral leishmaniasis (kala-azar) in human being. The disease is an often fatal and endemic in certain region of India and some other tropical and subtropical countries around the world.1

The expression of a major phosphorylated glycoconjugates on its surface membrane i.e., lipopolysaccharide (LPPS),2 which is most commonly known as lipophosphoglycan (LPG)3-5 anchored by glycosylphosphatidylinositol (GPI) and glycosylinositolphospholipid (GIPL) and Gp63, a metaloprotease are the ligand for macrophage receptor for the attachment of the parasite, which is the initial step for infection regulated by cAMP both in parasite and macrophage.6 The parasite also expresses highly glycosylated secreted acid phosphatase, which is considered beneficial by dephosphorylating a wide spectrum of bioactive molecules7,8 in both the environment encountered in its life cycle. Regulation of parasite physiology essential for the survival in hostile environments of host or vector in two distinct morphological forms is largely depend on signal transduction systems. Cyclic AMP inhibited the growth of Leishmania promastigotes9,10 but the nature of its action remains to be elucidated. Intracellular cAMP possibly involves in signal transduction in Leishmania promastigotes. The mRNAs of a family of putative receptor adenylyl cyclase are expressed in insect stage of L. donovani possibly involved in the increase of intracellular cAMP,11 the necessity of which is yet to be determined. The parasite exposes to the presence of intracellular cAMP modifying agents e.g. epinephrine, norepinephrine in host as
well as in vectors blood meal. However, the role of adrenergic receptor agonists and antagonists in the growth of *L. donovani* promastigotes is yet to be elicited.

In this report, we studied the effects of membrane permeable cAMP and some adrenergic receptor agonists and antagonist of intracellular cAMP formation on the growth of *L. donovani* promastigotes.

**MATERIALS AND METHODS**

Adenine, hemin, D-biotin, L-glutamine, folic acid, epinephrine, isoproterenol, propranolol, dibutyryl cyclic GMP (db-cGMP) and dibutyryl cyclic AMP (db-cAMP) from Sigma Co., USA; medium 199, HEPES and beef heart infusion (BHI) from Gibco Co., USA and agar from Difco Co., USA were purchased for the experiments. Radioactive \[^{32}P\]-orthophosphate was obtained from BARC, India. Other reagent grade chemicals were used in the study.

Parasite maintenance and effects of experimental agents on promastigote stage: Promastigotes of *L. donovani* (MHOM/IN/78/ UR6), an Indian strain subpassaged in solid blood agar medium consisting of BHI (3.7%, w/v), D-glucose (1.8%, w/v), agar (1.5%, w/v) and rabbit blood (2%, v/v) grown at 24±1°C for 144 hour. The growth profiles were recorded by counting promastigotes present in aseptically withdrawn 20µl sample from each experimental set diluting in formol-saline (4% formol) in a ratio of 1:1 for early period of culture or 1:10 as depending on the parasite concentration using a hemocytometer and the results were expressed as the mean of four separate experiments as described.\(^1^3\)

**Biosynthetically pulse labeling of the promastigotes with \[^{32}P\]-orthophosphate and its analysis:**

Promastigotes of log phase culture in S1-199 medium were washed twice with sterile saline (0.85%, w/v) by centrifugation at 1000 x g for 10 min at 4 °C. These washed parasites were allowed to grow in the fresh S1-199 medium to which \[^{32}P\] (50 µCi/ml of culture) was added in presence of experimental agents. Initial parasite concentration was adjusted to 2.0x10\(^7\)/ml. After 6 h of incubation at 24 ± 1 °C the parasites were harvested by centrifugation in eppendorf centrifuge and washed thrice with phosphate buffered saline, pH 7.2, at 4 °C. Immediately, the [\(^{32}P\)]-labeled parasites were suspended in denaturing sample buffer (1X, final) and boiled for 5 min at 100 °C and the macromolecules were subjected to separation by SDS-PAGE, Coomassie Brilliant Blue stained, dried and followed by autoradiography using Kodak X-Omat AR film with intensifying screen at -70°C. Statistical analysis: The results of the experiments were tested for statistical significance using the Student t test using Microsoft excel, Microsoft Corporation. Comparisons were made between the control (no drug) and each experimental set containing different drugs at different concentrations and further in between combination group and individual agent of that group. P value of <0.05 was considered as statistically significant.

**RESULTS**

The effect of db-cAMP, db-cGMP, epinephrine, isoproterenol and propranolol on the growth of *L. donovani* promastigotes:

The growth curve of promastigotes in control was presented in Fig.1 as inset based on mean with SD value of four identical experiments for each time point. The results obtained with experimental agents were also treated in the same way to determine the means, SD values and P values. The effects of experimental agents were expressed as percent of stimulation or inhibition in compared to growth in control at 72 h and 96 h incubation were shown in figure 1. Although the trends of effect of these compounds on cell division were very similar at 72 h and 96 h culture, it was more prominent in 96 h as shown in figure 1. Multiplication of *L. donovani* promastigotes was inhibited by 16% (P<0.01), 28% (P=0.001), 43% (P<0.0005) and 69% (P<0.0001) at 96 h culture by 10, 20, 50 and 100 µM db-cAMP compared to control, respectively (Fig-1A). On the contrary the activity of db-cGMP on the growth of the parasite was found significantly stimulatory by 15% (P<0.005), 41% (P<0.0005), 49% (P=0.0001) and 50% (P<0.0001) with 10, 20, 50 and 100 µM of the agent, respectively, at 96 h culture.

The growth of the parasite was drastically inhibited by epinephrine in a dose dependent manner. The inhibition was posted by 20% (P=0.001), 38% (P=0.0005), 59% (P<0.0001), and 83% (P=0.0001) with 10, 20, 50 and 100 µM epinephrine,
respectively at 96 h culture and the similar pattern of inhibition was recorded at 72 h culture (Fig. 1). We tested the effect different doses of isoproterenol, an agonist of \( \beta \)-adrenergic receptor on the growth of promastigote and found 11% (P<0.05), 24% (P<0.005), 32% (P<0.001) and 52% (P<0.0001) inhibition with 10, 20, 50 and 100 \( \mu \)M of the drug, respectively.

The effect of dibutyryl cAMP, epinephrine, isoproterenol and propranolol in the phosphorylated macromolecules LPG profile of \( L. \) donovani promastigotes: Regulation of LPG in \( L. \) donovani promastigotes by cAMP, epinephrine, isoproterenol and propranolol were visualized by autoradiography after SDS-PAGE analysis of \(^{32}\)P-labeled parasite in presence or absence of these agents individually for 6 h. A typical experimental result of three replicate experiments was shown in figure 2. There was no prominent change in protein profiles of promastigotes treated with or without experimental agents (Figure 2B. a’-e’). In control a heterogeneous macromolecule spanned in approx. 28-40 kDa region of the gel (Figure 2A. Lane a) that was leishmanial lipophosphopolysaccharide (LPPS), which is known as LPG confirmed by phenol extraction of the metabolically phosphate-labeled whole cell and analyzed by SDS-PAGE and autoradiography as described by Kar et al. 1991\(^2\) (Fig. not Shown). The reduction in the intensity of

Figure 1. Histograms show the percent of stimulation or inhibition of growth of \( L. \) donovani promastigotes treated with experimental agents for 72 h (blue bar) or 96 h (red bar), where, the growth of the parasite in control (no drug) was considered as zero at respective time points of culture based on the mean of four separate identical experiments. The growth curve with error bar (±SD, N=4) for control was presented as inset where each point stands for mean of four separate identical experiments. The experimental agents and their different concentrations were mentioned below for each pair of histogram for 72 h and 96 h culture, respectively.

Figure 2. Autoradiograph after SDS-PAGE (12.5% acrylamide) analysis of the effect of agonists and antagonists of \( \beta \)-blocker on the \(^{32}\)P-phosphorylation of \( L. \) donovani UR6 promastigotes was shown in figure 2A. Equal number of \(^{32}\)P-labelled UR6 promastigotes(1x10\(^{7}\)) for each sample were loaded per lane. Lane a, control (without any drug); lane b, db-cAMP (100 \( \mu \)M); lane c, adrenaline (100 \( \mu \)M); lane d, propranolol (100 \( \mu \)M) and lane e, isoproterenol (100 \( \mu \)M). The corresponding protein profiles were shown in Fig 2B: lane a’, b’, c’, d’ & e’ for control, db-cAMP, adrenaline, propranolol and isoproterenol, respectively.
The band observed in the autoradiograph from higher to lower was registered with the agents in the following manner: propranolol > epinephrine > cAMP > isoproterenol at 100 μM each (Figure 2A. Lane d, c, b & e, respectively).

**DISCUSSIONS**

Intracellular levels of cAMP have been demonstrated to modulate metabolism and replication of prokaryotic and eukaryotic cells. It was reported that the growth of *L. tropica* and *L. donovani* were inhibited by 70% with dibutyryl cAMP at 1 μM. We have found the same degree of inhibition with one tenth of that concentration of the agent. This difference obtained may be due to the strain variation and/or culture condition. The variation may be attributed to the difference in the level of cAMP phosphodiesterase activity which is essential for the down regulation of cAMP.

It is prominent that the growth stimulatory effect was observed with the gradual increase of db-cGMP upto 50 μM, and beyond this concentration there was no more beneficial to the parasite may be due to the limitation of the culture medium as a whole. Availability of cGMP in a large amount from external source was found beneficial to *Leishmania* promastigotes may be attributed to modulate some intracellular signaling, which is different than the effect of cAMP and/or serve as added purine source as the purine salvage pathway is active in *Leishmania* and the parasite essentially expresses enzymes including phosphatases and nucleases for scavenging purine.

It is evident from our results that the parasite physiology was significantly more modulated by epinephrine than isoproterenol and this may be vested on both α and β adrenergic receptor specificity of epinephrine whereas isoproterenol is an only β1 & β2 adrenergic receptors agonist.

*Leishmania* LPG, is a highly complex macromolecule composed of four distinct domains: a GPI anchor consists of an alkyl phosphatidylinositol having a single saturated C24~26 aliphatic chain; a glycan core consists of a heptasaccharide comprising two galactopyranosides, a galactofuranoside (Galf), two mannosides and a glucosamine residue attached to inositol; a linear phosphoglycan chain (PG) consists of 15–40 phosphodiaccharide (Galβ1,4Manβ1-PO4) units; and finally, LPG is terminated by a oligosaccharide cap having di-, tri- or tetrasaccharide consisting of galactose and mannose assembled as Manα1,2Manα1 or as Gal β1, 4 (Man α1,2) Man α1 depending on the *Leishmania* species. It is clear from our results that at least some of the steps in the synthesis of LPG are affected by cAMP. The reduced infectivity of promastigotes to macrophage by cAMP was documented and this is possibly due to the reduced LPG expression and affecting other cellular physiology of the parasite by cAMP and its modifiers.

This study has clearly shown that the synthesis of LPG, the major glycoconjugates of *L. donovani* was controlled by cAMP and significantly modulated by adrenergic receptor agonists epinephrine, which is effective on both α and β-receptors than isoproterenol, a nonselective β1-adrenoreceptor agonist which is well known for contraction of cardiac muscle by opening of calcium channel and dilatation of smooth muscle by closing of calcium channel via β2-receptor. Propranolol is a β1 and β2 adrenoreceptors antagonist and has membrane stabilizing activity. Propranolol significantly affects the replication of *L. donovani* promastigote and synthesis of LPG. However, the precise nature of activity of propranolol on *Leishmania* promastigotes is yet to be determined.

**CONCLUSION**

This report describes the role of cAMP, epinephrine, isoproterenol and propranolol in *Leishmania donovani* promastigotes. The growth of *L. donovani* promastigotes at 96 h culture was inhibited by 69%, 83, 52% and 95% with db-cAMP, epinephrine, isoproterenol and propranolol at 100 μM each, respectively in compared to control. The similar patterns of growth inhibition of the parasite for all the agents were also observed at 72 h culture. Multiplication of parasite was stimulated by db-cGMP. The maximum growth stimulatory effect was achieved with 50 μM db-cGMP. The expression of LPG of the parasite was drastically affected with propranolol and epinephrine in compare to cAMP or isoproterenol at 100 μM of each agent for 6 h exposer to the promastigotes in defined culture medium, whereas the protein profiles apparently remain identical with the control.

**ACKNOWLEDGEMENT**

The entire experimental work was done in the Indian Institute of Chemical Biology, Kolkata, India, during 1989 to 1997. Data interpretation and manuscript writing were done during our stay in
College of Medical Science-TH, Bharatpur, Chitwan, Nepal. Authors are grateful to Late Prof. A.N. Bhaduri and Late Dr. J. Das, Ex-Directors, and Dr. D.K. Ghosh, Ex-Asst. Director, Indian Institute of Chemical Biology, Kolkata 700032, India, for providing facilities for part of this work. We are thankful to Dr. V. Nattraj Prasad and Dr. M Mathur for their support. Our special thanks go to Dr. Manohar Pradhan for his constant encouragement for preparing this manuscript.

REFERENCES

12. Kar K. Folic acid the essential supplement to brain heart infusion broth for cultivation and cloning of Leishmania donovani promastigotes. Parasitology 1997;115: 231