Increase in MMP-9 Expressing CD11b+ cells in a CD44 Dependent Way Reduces Severity of Experimental Autoimmune Encephalomyelitis

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ABSTRACT

Background: Multiple sclerosis (MS) is a dangerous neurodegenerative disorder. Various aspects of the disease have been studied in experimental animal model. Migration of immune cells to the central nervous system (CNS) is a predominant feature of MS. CD44 molecule has been reported to be involved in many important biological processes including contribution in severing inflammation in experimental autoimmune encephalomyelitis (EAE). Matrix metalloprotease-9 (MMP-9) interaction with CD44 has been well known to be involved in cellular adhesion, transmigration and inflammation. In this study, we were interested to examine the role of phagocytic cells expressing MMP-9 in resolving EAE.

Materials and Methods: C57BL/6 WT and CD44 KO mice were used as EAE animal model. The level of phagocytic cells expressing MMP-9 in the secondary lymphoid organs were assessed in EAE induced WT as well as CD44 KO animals. Results: EAE severity was found in CD44 KO group compared to WT. Level of CD11b cells (marker of phagocytic cell) in the peritoneal cells expressing MMP-9 was higher in WT compared to CD44 KO. CD11b stained area found to be greater in WT lymph node compared to CD44 KO.

Conclusions: This observation suggests the role of CD44 molecule in modulating the immune scenario which is related to disease severity. This study also opens avenues for the specific inflammatory roles of different immune cells in MS.

Keywords: multiple sclerosis; EAE; CD44; MMP-9; CD11b; CNS.

INTRODUCTION

Experimental Autoimmune Encephalomyelitis (EAE) is the primary animal model for multiple sclerosis (MS) research. 1 The blood brain barrier (BBB) plays a pivotal role in protecting the CNS by preventing entry of unwanted cells and other materials. Proliferation and migration of auto reactive lymphocytes to the CNS is known to be a predominant criteria of EAE. Easy migration of the immune cells is certainly related to the permeability through blood BBB. 2 CD44 is a novel transmembrane glycoprotein which is able to alter the membrane permeability and its absence severe EAE. 3 The protective role of CD44 in EAE has been suggested by many researchers in relation to matrix metalloproteases (MMPs), cytokines, chemokines and specific T lymphocyte populations. 3

CD44 is known to mediate faster rolling of neutrophils along with macrophages towards the site of infection and inflammation. 4 CD11b is a well-known molecule and expressed mainly on monocytes, macrophages, dendritic cells, neutrophils. 5 This molecule (CD11b) is frequently used to identify macrophages and microglia. This study suggests that CD44 exerts a protective effect in EAE which is linked to MMP-9+ phagocytic cell increase in the secondary lymphoid organs and probable migration of some specific immune cells to the central nervous system (CNS).

MATERIALS AND METHODS

Mice and reagents
C57BL/6 wild-type (WT) and CD44-KO mice (The Jackson Laboratory, Bar Harbor) were housed in the animal facilities at Yale University School of Medicine according to Yale University and NIH guidelines. All mice were used between 8 and 10 weeks of age. Murine MOG peptide was synthesized by the W.M. Keck Biotechnology Resource Laboratory at Yale University (New Haven, CT). Fluorescent conjugated primary antibodies anti-CD11b and anti-MMP-9 were purchased from BD Bioscience (San Jose, CA).

EAE Induction
Mice received 300mg of rodent MOG peptide emulsified in complete Freund’s adjuvant containing heat-killed Mycobacterium butyricum (Difco, Detroit, MI) via subcutaneous injection on day 0 and...
7; on day 0 and 2, mice received 500ng of pertussis toxin via intra-peritoneal injection. The mice were monitored daily and graded for clinical symptoms of EAE on the following basis: 0, no disease; 1, flaccid tail; 2, hind-limb weakness/unsteady gait; 3, hind limb paralysis; 4, hind-and fore-limb weakness/paralysis; 5, moribund.9

**Immunofluorescence and Flow Cytometry**
For immunofluorescence the mice were anesthetized and processed as described.3 Six micrometer thick sections of lymph nodes were used for CD11b staining. Before staining, sections were washed with PBS and blocked for 1 hour at room temperature with 1%bovine serum albumin, 3% normal goat serum, and 0.3% Triton X-100 in PBS. Those sections were incubated with ratanti-CD11b (BD Bioscience) for 2 hours at room temperature, followed by goat anti-rat IgG conjugated to Alexafluor-594 (Invitrogen). For flow cytometry, peritoneal cells were collected by peritoneal lavage from WT and CD44 KO mice and prepared for flow cytometry as described.5 Staining for CD11b and MMP-9 were performed using fluoresceinconjugated CD11b (BD Bioscience) and MMP-9 (Santa Cruz) antibodies.

**Statistical analysis**
Differences between WT and KO datasets were analyzed using P values determined by Student’s t-test. Error bars represent SEM (standard error of mean).

**RESULTS**

**Absence of CD44 resulted in severity of EAE**
To reconfirm the effect of deletion of CD44 on EAE severity, we assessed the disease scores in WT and CD44 KO mice in each of the set of experiments. We found increased EAE severity in the CD44 KO mice compared to WT mice in each set of experiment (Figure 1).

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**Decrease in the number of CD11b+ cells in the lymph node of CD44 KO mice**
To evaluate quantities of phagocytic cells with in the secondary lymphoid organ, lymph nodes were isolated from WT and CD44-KO animals with EAE at the same time point. Immunofluorescent analysis revealed significantly decreased numbers of CD11b+ (phagocytic cells) cells in the lymph node of the CD44-KO animals compared to WT (Figure 2).

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**Decrease in the number of MMP-9+ phagocytic cells in the peritoneal lavage fluid of CD44 KO mice**
To assess the scenario of MMP-9+ phagocytic cells in the secondary lymphoid organs, we stained the peritoneal cells for CD11b. Those cells were double stained also with MMP-9 antibody. FACS analysis of the peritoneal cells revealed that MMP-9+ phagocytic cells were obviously higher in WT mice compared to CD44 KO (~95% vs ~84%) (Figure 3).
DISCUSSION
EAE, the animal model of MS is involved with disturbance in the body compartments those provide immunity. The pathogenesis of the disease is complex and depends on multiple cell types and processes. The present finding suggests that CD44 provides protective role to some extent in EAE, which is involved with MMP-9 expressing phagocytic cells.

CD44 has already been reported as a negative regulator of inflammation related to T-cell differentiation, adhesion, transendothelial migration and BBB permeability. CD44 KO junctions are weaker barrier and obviously permeable to the immune cells in an easier way. The weaker barrier in the CD44 KO case is reported to restrict larger molecules. Increased expression of MMPs in CD44 KO endothelial cells links to weak junctional adhesion, MMP mediated degradation of extracellular matrix and loss of vascular integrity. This could allow the T lymphocytes to migrate into the CNS and increase disease severity. On the other hand, decrease in the number of phagocytic cells in CD44 KO mice and co-relation of disease severity is meaningful, because phagocytic cells are so important for causing as well as resolving inflammation.

CONCLUSIONS
Higher number of CD11b+ cells in secondary lymphoid organs (which is a popular marker of phagocytic cells) is obvious in WT mice compared to CD44 KO mice and suggesting the role of phagocytic cells in regulation of the disease severity. These CD11b+ cells were also MMP-9+. Presence of CD11b+ cells was higher in the lymph node of WT compared to CD44 KO mice. So MMP-9 expressing phagocytic cells probably taking part in the healing of inflammation to some extent which is not happening in CD44 KO mice. It would be of great interest to study and unwind the MMP-9 expression in specific subsets of immune cells in CNS co-related with EAE severity.

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