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ABSTRACT

The hepatoprotective properties of the ethanolic extracts of *Punica granatum* peels, *Crataeva religiosa* leaves and *Jasminum polyanthrum* leaves were evaluated for the paracetamol induced liver toxicity in mice. Antioxidant, cytotoxicity and phytochemical screening were carried out to assure the therapeutic efficacy of these plants. Oral administration of plant extracts for 7 days in mice significantly reduced the impact of paracetamol toxicity on the serum markers of liver damage, aspartate transaminase (AST), alanine transaminase (ALT) and total protein. The extract showed significant hepatoprotective effects as evidenced by decreased serum enzyme activities like ALT, AST and changed total protein. The phytochemical and antioxidant studies also supported that plant extract markedly reduced the toxicity due to presence of antioxidant phytochemicals. The result suggests that *Jasminum polyanthrum* is potent hepatoprotective agent against paracetamol induced hepatotoxicity in mice.

Key words: Plants, Antioxidant, Hepatoprotective, Enzyme, Protein.

INTRODUCTION

Natural remedies from medicinal plants are considered to be effective and safe alternative treatments for hepatotoxicity. Over the past decades, herbal medicines have garnered growing attention as potential therapeutic agents to prevent and treat liver disease (Yao et al., 2016). These herbal sources have shown the ability to maintain the normal functional statues of liver. In Nepal, 100 more than medicinal plants are ethnomedicinally used in the preparations of liver remedies (Baral & Kurmi, 2006). Liver protective plants contain complex chemical constituents with multi-target pharmacological multi-level and development of plant based activity. The hepatoprotective drugs has been given importance in accordance with the composition principles of herbs (Sharma et al., 1991). The efficacy of such plants has been previously explored by various hepatoprotective researchers for their and immunomodulatory activities (Ilyas et al., 2016).

The liver plays an astonishing array of vital functions in the maintenance, performance and regulating homeostasis of the body. It is involved with almost all the biochemical pathways. Most of the hepatotoxic chemicals damage liver cells mainly by lipid peroxidation and other oxidative damages (Dianzai *et al.*, 1991). High dose of paracetamol is one of the factors for hepatic necrosis that can result in the liver failure. Some researches have been carried out on the hepatoprotective property of *Punica granatu* (Celik *et al.*, 2009) and *Crataeva religiosa* (Jaikanth *et al.*, 2015). But similar studies in plants originated from Nepal have not been done. Therefore, in the present study, efficacy of the plants was studied on biochemical and pharmacological parameters with an objective to ameliorate the liver disease induced by the paracetamol.

MATERIALS AND METHODS

Plants material

The peels of *Punica granatum*, leaves of *Crataeva religiosa* and leaves of *Jasminum polyanthrum* were collected. Plant materials were identified by botanist and herbarium specimen was deposited in the Department of Pharmacy, Kathmandu University. Samples were washed, dried under shade and powdered to coarse. The powdered material was extracted with alcohol (80% ethanol) in Soxhlet apparatus for 48 hrs. For oral administration, extracts were dissolved in distilled water.

Phytochemical Screening

The crude methanolic extracts of the barks of different plants were screened to detect the presence of phytochemicals, as per the standard screening procedure (Trease & Evans1996).

DPPH radical scavenging assay

The DPPH radical scavenging activity of extracts was determined using the method of Brand-Williams et al. (1995) with some modifications. Briefly, test samples were prepared by adding 3ml of 100µmol methanolic DPPH solution in 3ml methanolic plant extract solution of varying concentrations (2, 4, 6, 10µg/ml). Corresponding blank sample was prepared by adding 3ml plant methanolic extract and 3ml of methanol. Mixer of 3ml methanol and 3ml DPPH solution was used as control. All these mixtures were immediately kept in dark to protect from light and were allowed to react. After 30 minutes, absorbance was noted at UV-visible 517nm using spectrophotometer. Standard solution 2, 4, 6, 10µg/ml of ascorbic acid was prepared from stock solution.

Brine shrimp Lethality Test

Ethanolic extract of the samples were evaluated for lethality to Brine shrimp larvae (A. salina Leach) according to the published procedure (Meyer et al., 1982). Brine shrimp eggs were suspended for 48 hours in a conical flask containing 300 ml of artificial seawater for hatching (the eggs). The flasks were well aerated with the aid of an air pump, and kept in a water bath at 29-30°C. The extracts were dissolved in 1% aqueous DMSO, and then in sea water to obtain a concentration of 1000 ppm, 500 ppm, 250 ppm, 100 ppm, 50 ppm, 25 ppm and 10 ppm. An aliquot of each concentration (1 ml) was transferred, in triplicate, into clean sterile vials with pipette, and aerated seawater (9 ml) was added. Ten Brine shrimp nauplii were transferred to each vial. 1% DMSO in seawater was used as negative control. After 24 hours, the numbers of survivors were counted, and the mortality percentage was calculated. The lethal concentration for 50% mortality after 24 hours of exposure, the chronic LC50 and 95% confidence intervals were determined by using the Probit method as the measure of toxicity of the extract or fractions. The extracts were considered as toxic at LC_{50} less than 10 µg/ml (Setzer *et al.*, 2001).

Animals

Inbred Swiss albino mice weighing 20- 30 g of either sex, bred in Central Animal House, Kathmandu University, Department of Pharmacy were procured and used for the study. The animals were fed food pellets and water ad libitum. Animals were maintained in standard lab conditions (12 h: 12 h dark and light cycle at 25±2 °C temperature). The study was approved by the Institute Animal Ethics Committee, Kathmandu University, Dhulikhel.

Hepatoprotective activity

The treatments of mice were categorized under 6 groups, each comprising six animals.

- Group 1: Control group with aqueous vehicle for 7 days.
- Group 2: Paracetamol (250mg/kg) on 7th day only.
- Group 3: Silymarin (extracted from the *Silybum marianum*) (500mg/kg) for 7 days + Paracetamol (250mg/kg) on 7th day
- Group 4: Pomegranate extract (200 mg/kg) for 7 days + Paracetamol (250 mg/kg) on 7th day.
- Group 5: Crataeva extract (200 mg/kg) for 7 days + Paracetamol (250 mg/kg) on 7th day.
- Group 6: Jasmine extract (200 mg/kg) for 7 days + Paracetamol (250 mg/kg) on 7th day.

Collection of blood and biochemical analysis

On the 9th day, all the animals were sacrificed by mild ether anesthesia. About 0.5 ml of blood was drawn from each mouse and transferred to separate vials. All the blood samples were kept in a container containing icepacks and then taken to the laboratory for the biochemical estimation of alanine aminotransferase (ALT), Aspartate aminotransferase (AST) and total protein (Babson *et al.*, 1966).

Toxicity evaluation

The acute oral toxicity study of the extracts was carried as per the Organisation for Economic Cooperation and Development (OECD 420) guidelines. Administration of the extracts stepwise up to the dose 200mg/kg in aqueous vehicle 20 ml/kg of body weight caused no considerable signs of toxicity in the tested animals.

RESULTS AND DISCUSSION

Phytochemical screening of the plants

The randomly selected plants subject to phytochemical screenings were found to contain different metabolites *viz.* alkaloids, tannin, flavonoids, terpenoids, resin and glycosides.

Antioxidant activity

The effect of plant extracts on antioxidant property showed that, Jasmine extract showed strongest antioxidant activity with IC_{50} 0.73. Pomegranate extract was also found equally strong in comparison to standard i.e. ascorbic acid. Liver damage causes and releases the enzymes ALT, AST and total protein in the serum due to the membrane and cellular damages of the liver. Paracetamol is a hepatotoxin which induces the liver damage by affecting the metabolic functions. The liver enzyme level

concentration was increased in serum due to the toxic effect of paracetamol. The decrease in liver enzyme levels in serum is due to the protective effect of given plant extracts on liver cells followed by restoration of membrane permeability (Sethuraman, 2003). Phenol compounds are responsible for hepatoprotective and antioxidant activity of plant extracts (Singh et al., 1993). They exhibit antioxidant activity by inactivating lipid free radicals or preventing decomposition of hydroperoxides into free radicals.

Demonster	Groups				
Parameter	Ascorbic acid	Pomegranate	Jasmine	Crataeva	
IC50 value (mcg/ml)	6.29	5.86	0.73	10.74	

Biochemical characterization

The present study was performed to assess the hepatoprotective activity of various extracts of *Punica granatum*, *Crataeva religiosa* and *Jasminum polyanthrum* in mice against paracetamol as hepatotoxin which cause liver damage. Altering liver microsomal membranes of hepatocytes and cell damage results in release of enzymes AST, and ALT. Levels of AST, ALT and total protein in serum used to assess hepatic function in mice were

changed (Table 2). The hepatic enzymes were significantly controlled by plant extracts. Among the 6 groups, ethanolic extract of Jasmine and Creativa have effectively controlled the liver damage by restoration of the levels of liver function. The level of ALT, AST and total protein was found to be significantly improved in comparison to paracetamol treated mice. Similar result on Jasmine was found in a recent study (Shekhar & Prasad, 2015).

Table 2. The levels of ALT, AST, and total protein in different groups of mice.

Parameters	Groups						
	Control	Paracetamol	Silymarin	Pomegranate	Crataeva	Jasmine	
ALT (IU/l)	48.6 ± 26.8	250.2 ± 37.65	170.8 ± 14.25	204.4±14.51	155±33	151.4±46.74	
AST (IU/l)	130±59.83	299.60±73.73	112.2±12.37	220.2±20.21	196.6±37.65	204.8±24.91	
Total protein (gm%)	5.24±1.24	6.28±1.06	8.44±1.59	4.74±0.69	5.28±0.75	7.52±1.08	

Brine Shrimp Lethality Test

Based on the Brine shrimp bioassay analysis, the LD_{50} value of the Crataeva plant was found to be strong while Pomegranate and Jasmine were similar (Table 3). Though these plants are edible, they were found to be toxic during Brine shrimp assay. But after the administration of extracts at the dose of 200mg/kg in mice, they did not show any toxicity *in vivo*.

Table 3. The cytotoxicity of plant extracts inBrine shrimp larvae.

Parameters	Groups			
rarameters	Pomegranate	Crataeva	Jasmine	
LD50 (ug/m)	3.58	2.52	3.025	

The result of DPPH assay indicates that these plants possessed good anti-oxidant properties. Administration of 200 mg/kg of each extract has shown no toxicity in mice. Statistical analysis of the enzyme levels in the mice was significant in producing hepato-protective action. The levels of ALT and AST were reduced significantly in extract treated groups and the effect is comparable to that of standard drug treatment with extracts decreased the serum levels of ALT and AST towards the respective values. This is an indication of stabilization of plasma membrane as well as repair of hepatic tissue damage caused by paracetamol. The above changes can be considered as an expression of the functional improvement of hepatocytes, which may be caused by an accelerated regeneration of parenchymal cells. Protein metabolism is a major role of liver and healthy functioning of liver is required for the synthesis of the serum proteins except the gammaglobulins. Hypoproteinemia is a feature of liver damage due to significant fall in protein synthesis (Kanchana & Sadi, 2011). Hypoproteinemia was observed after paracetamol ingestion but the trend turned towards normal in pomegranate and crataeva extract treated mice. Some researchers have carried out the hepatoprotective property of Punica granatum and revealed that constituents present in plant impart protection against carcinogenic chemical induced oxidative injury that may result in development of cancer during the period of a 52day protective exposure (Celik et al., 2009). In another study, Crataeva religiosa was found to significantly restore hepatoprotective enzymes (Jaikanth et al., 2015). This restoring effect of the extracts indicate that the phytochemicals present in extracts such as flavonoids, tannins and terpenoids might have acted in a coherent manner to neutralize the toxicity of paracetamol and also they may have exhibited a stabilizing effect on the plasma membranes of hepatocytes to restore the levels of serum enzymes. On the basis of ALT level, which is more specific to the liver, Jasmine plant was found to possess significant hepatoprotective property.

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REFERENCES

Babson, A. L.; Greeley, S. J.; Coleman, C. M. and Philips, G. D. (1966). Phenolphthalein monophosphate as a substrate for serum alkaline phosphatase. *Clinical Chemistry*, **12**: 482-490.

- Baral, S. R. and Kurmi, P. P. (2006). A compendium of medicinal plants in Nepal .Publisher: Mrs. Rachana Sharma, 281 Maiju Bhal, Chabahil, Kathmandu, Nepal.
- Brand-Williams, W.; Cuvelier, M. E. and Berset, C. (1995). Use of a free radical method to evaluate antioxidant activity. *Lebenson Wiss Technology*, **28**: 25-30.
- Celik, K.; Temur, A. and Isik, I. (2009). Hepatoprotective role and antioxidant capacity of pomegranate (Punica granatum) flowers infusion against trichloroacetic acidexposed in rats. *Food Chemistry and Toxicology*, **47** (1): 145-149.
- Dianzai, M. U.; Muzia, G.; Biocca, M. E. and Canuto, R. A. (1991). Lipid perioxidation in fatty liver induced by caffine in rats. *International Journal of Tissue Reactions*, 13: 79-85.
- Ilyas, U.; Katare, D. P.; Aeri, V. and Naseef, P. P. (2016). A review on hepatoprotective and immunomodulatory herbal plants, **10**: 19, 66-70.
- Jaikanth, C. M.; Venkateswaran, K. V.; Selvasubramanian, S. and Sesh, P. S. L. (2015). Effect of Extracts of Crataeva religiosa on Hepatic Markers in Liver Intoxication. *Indian Veterinary Journal*, **92** (9): 86-88.
- Kanchana, N. and Sadi, A. M. (2011).
 Hepatoprotective effect of Plumbago zeylanica on paracetamol induced liver toxicity in rats. *International Journal of Pharmacy and Pharmaceutical Sciences*, **3** (1): 151-154.
- Meyer, B. N.; Ferrigni, N. R.; Putnam, J. E.; Jacobsen, L. B.; Nichols, D. E. and McLaughlin, J. L. (1982). Brine Shrimp: A convenient general bioassay for active plant constituents. *Planta Medica*, **45**: 31-34.
- Sethuraman, M. G.; Lelitha, K. G. and Kapur, B. R. (2003). Hepatoprotective activity of *Sarcostemma brevistigma* against CCl4 induced hepatic damage in rats. *Current Science*, **84**: 1186-1197.
- Setzer, M. C.; Setzer, W. N.; Jackes, B. R.; Gentry, G. A. and Moriarity, D. M. (2001). The medicinal value of tropical rainforest plants from Paluma, North Queensland, Australia. *Pharmaceutical Biology*, **39**: 67–78.

- Sharma, A.; Chakraborti, K. K. and Handa, S. S. (1991). Antihepatotoxic activity of some Indian herbal formulations as compared to silymarin. *Fitoterapia*, **62**: 229-235.
- Shekhar, S. and Prasad, M. P. (2015). Comparative analysis of antioxidant properties of jasmine species by hydrogen peroxide assay. *European Journal of Biotechnology and Bioscience*, **3**(2): 26-29.
- Singh, B.; Saxena, A. K.; Chandan, B. K.; Agarwal, S. G.; Bhatia, M. S. and Anand, K. K.

(1993). Hepatoprotective effect of ethanolic extract of Eclipta alba on experimental liver damage in rats and mice. *Phytotherapy Research*, **7** (2):154-158.

- Trease, G. E. and Evans, W. C. (1996). Pharmacognosy. Alden Press, Oxford, UK.
- Yao, H.; Qiao, Y. J.; Zhao, Y. L.; Tao, X. F.; Xu, L. N.; Yin, L. H.; Qi, Y. and Peng, J. Y. (2016). Herbal medicines and nonalcoholic fatty liver disease. *World Journal of Gastroenterology*, 22 (30): 6890-6905.