Anti-inflammatory, Anxiolytic and Antioxidant Property of *Lactuca sativa* L and Formulation of Microspheres Loaded Sustained Release Anti-inflammatory Gel

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Abstract

Anti-inflammatory and anxiolytic properties of *Lactuca sativa* L extract was evaluated by formalin induced mice paw edema test, and elevated plus maze animal model respectively. Solid microspheres containing dispersed extract were obtained by water-in-oil-in-water multiple emulsion system followed by solvent evaporation method. Optimization was carried out using Minitab 16 and optimized microspheres were loaded into a gel base. Anti-oxidant activity of optimized microspheres loaded gel was evaluated using DPPH (2,2-diphenyl-1-picrylhydrazyl) assay. The gel was also evaluated for its physical properties, drug release kinetics and anti-inflammatory activity using formalin induced mice-paw edema test where the anti-inflammatory potential was compared to marketed diclofenac gel (1.16% w/w). All doses of extract as well as microsphere loaded sustained release anti-inflammatory gel showed a dose and time dependent inhibition of edema (P<0.05), the extract also showed a dose dependent inhibition of level of anxiety (P<0.001). Potential anti-oxidant activity was seen with different concentration of crude extract and microsphere loaded gel. The anti-oxidant potential was compared using IC50 value which was 5.62 µg/mL, 6.34 µg/mL, 8.25 µg/mL for ascorbic acid, methanolic extract of *L. sativa* and microspheres loaded gel respectively.

Keywords: *Lactuca sativa* L, anti-inflammatory and anxiolytic activity, antioxidant, microspheres, sustained release gel

Introduction

Inflammation is a normal biological process in response to tissue injury, microbial pathogen infection, and chemical irritation. Inflammation is initiated by migration of immune cells from blood vessels and release of mediators at the site of damage. This process is followed by recruitment of inflammatory cells, release of ROS, RNS, and pro-inflammatory cytokines to eliminate foreign pathogens, and repairing injured tissues. In general, normal inflammation is rapid and self-limiting, but aberrant resolution and prolonged inflammation cause various chronic disorders [1]. The unique antioxidant combinations could be used therapeutically to reduce the inflammatory response and stimulate the immune system. Mechanisms of antioxidant action can include; suppression of ROS formation either by inhibition of enzymes or by chelating trace elements involved in free radical generation; scavenging ROS; and up-regulation or protection of antioxidant defenses. Anxiety disorders may be caused by problems in the functioning of brain circuits that regulate fear and other emotions. Moreover, certain environmental factors such as a trauma or significant event may trigger an anxiety disorder in people who have an inherited susceptibility to developing the disorder [2-4]. Oxidative stress and inflammation are the basis of many chronic diseases.

*Lactuca sativa* L is a leafy vegetable, an annual plant with basal rosette leaves. It requires relatively low temperature to prevent from flowering quickly. Its extracts are used in the skin cream and lotions to...
to be beneficial in liver issues. It possesses sedative, anticonvulsant, anxiolytic, analgesic, hypoglycemic and antifungal, anticancer properties. It was also demonstrated that it possesses antioxidant, anti-inflammatory and laxative property [5].

Microspheres are monolithic sphere or therapeutic agent distributed throughout the matrix either as a molecular dispersion of particles. Microspheres provide constant and prolonged therapeutic effect, which will reduce the dosing frequency and thereby improve the patient compliance. Better drug utilization will improve the bioavailability and reduce the incidence or intensity of adverse effects. Microsphere morphology allows a controllable variability in degradation and drug release [6].

Plant derived phytochemicals and extracts have recently attracted the great interest towards their applications in health care system. The development of herbal formulations into suitable doses form and evaluating the pharmaceutical parameters are utmost important for scientific assurance of such traditional applications and public acceptance of these herbs. The medicinal plants described in the present investigation have been selected on the basis of its traditional applications in Nepal. To promote the use of Nepalese medicinal plants as potential sources of skin care products, it is important to thoroughly find out their phytochemical profile, bioactive properties, design suitable formulation. Taking these precedents into consideration, the objective of the present study was to formulate and evaluate the pharmacological properties of extract and formulate the anti-inflammatory gel.

Materials and Methods

Plant material

_Lactuca sativa_ L (Asteraceae family) leaves were collected from local agricultural farm at Kathmandu valley, Nepal and identified by Botanist Tirtha Maiya Shrestha, visiting faculty of Department of Pharmacy, Kathmandu University, Nepal. The air dried ground (mesh 80) plant material was soaked in 90% ethanol in conical flask for 48 h. The extract was separated from the residues by filtering through Whatman no. 1 filter paper. The extraction procedure was carried out twice with sample. Thus, the extraction was completed in two weeks. The combined extracts were dried at 40 °C using a rotary evaporator. The dried crude extracts were weighted to calculate the yield and stored in refrigerator (-4 °C) until used for analyses.

Animals

The Swiss albino mice were purchased from Okharpaunwa Animal Farm, Tripureshwar, Kathmandu. The mice were kept in animal house of University before the experiment but transferred in polypropylene cages with free access to diet and water.

Preliminary phytochemical screening

Preliminary screening for the presence of alkaloids, tannins, polyphenols, reducing sugar, coumarins, saponins, anthraquinone glycoside and flavones by the reported protocol [7].

Anti-inflammatory activity

For the evaluation of anti-inflammatory activity of gel, total of 20 Swiss albino male mice of average weight 25 g were placed into cages and grouped into four (A-D) consisting five mice in each group. Diclofenac gel (1.16% w/w) was applied to group A mice on the inflamed area, 2% and 4% optimized microsphere loaded gel to group B and C respectively. Group D was used as control and therefore carbopol base was applied to the inflamed area. Paw volume was measured plethysmometrically before standard and extract administration and 30 minutes post administration of doses for 2 hours. Level of inflammation was calculated and data were statistically analyzed [8].

Anxiolytic activity

The anxiolytic activity was examined by using the Elevated Plus Maze apparatus. A total of 20 Swiss albino male mice of average weight 25 g were used. They were placed into cages and grouped into four (A-D) consisting five mice in each group. The animals were deprived of feed for 12 hours prior to experiment and were allowed access to pure drinking water. Individual oral dosage 0.25 mL of diazepam 2mg/kg to group A, test dose 200 mg/kg extract to group B and test dose 400 mg/kg to group C and group D was used as control i.e. distilled water was given orally. After 60 minutes, each of them was placed in elevated
plus maze apparatus consisting open arm and closed arm. The entry in open arm and closed arm were noted and compared. The residence time in respective arm and relative preference to open arm was analyzed to determine anxiolytic potential of doses [9].

Microspheres development and formulation of microsphere loaded gel
Ethyl cellulose was dissolved in 1:1 solution of acetonitrile and chloroform. Aqueous drug solution was added drop wise using high-speed magnetic stirrer operating around 900 rpm for about 10 minutes to prepare w/o primary emulsion. Primary emulsion was then subjected to probe sonication for given time interval and then added to external phase containing surfactant (tween 80) at magnetic stirrer speed around 900 rpm for 10 minutes and then again probe sonication was done for the same time interval. It was finally stirred at 900 rpm for 2 hours at room temperature for evaporation of organic solvent. Obtained microspheres were collected by filtration and then dried in hot air oven for 15 minutes and stored in desicator at ambient temperature until further evaluation [10]. Microspheres were evaluated for physical parameters including particle size and entrapment efficiency. Particle size analysis was done using trinocular stereo zoom microscope and the images were analyzed for spherical surface morphology and size was noted with the help of graduated scale on the slide. The drug content of microsphere was determined spectrophotometrically (λ max = 281 nm). A sample of microsphere (10 mg) was dissolved in 10 mL of phosphate buffer (pH 6.8) and kept overnight. The drug content in microspheres was calculated according to following formula:

\[
\text{Drug release percentage} = \frac{\text{Conc. (mg/mL)} \times 50 \text{ (mL)} \times \text{Dilution factor} \times 100}{\text{Drug content in microspheres (mg)}}
\]

In-vitro release profile of gel was studied by dissolution apparatus USP XXIII (paddle type). Cellophane membrane was used and weighed quantity of gel containing 100 mg of drug was introduced in the membrane and clamped on both sides. This was then dipped in basket containing 6.8 pH buffer as dissolution medium. The speed of the rotation was 50 rpm and temperature was maintained at 37±0.5 °C. Sample aliquots were withdrawn from dissolution medium at predetermined time intervals and were analyzed by UV spectrophotometer at 281 nm. The drug release percentage was calculated as:

\[
\text{Drug release percentage} = \frac{\text{Conc. (mg/mL)} \times 900 \text{ (mL)} \times \text{Dilution factor} \times 100}{\text{Drug content in microspheres (mg)}}
\]

Antioxidant activity
The scavenging activity of sample for DPPH was monitored according to method of Yen [11]. Briefly a 1 mL of aliquot test sample was added to 1 mL of 0.1 mM DPPH methanolic solution. The mixture was vortexed for 1 minutes and left to stand at room temperature and absorbance was read at 517 nm. The ability to scavenge DPPH radical was calculated using formula:

\[
\text{Percentage scavenging} = \frac{A_0 - A_1}{A_0} \times 100
\]

Where, \( A_0 = \text{Absorbance of DPPH solution} \)
\( A_1 = \text{Absorbance of DPPH along with different concentration of microsphere loaded gel and crude extracts (1 µg/mL, 2 µg/mL, 3 µg/mL, 4 µg/mL, 5 µg/L, 10 µg/mL)} \)

IC\(_{50}\) (Inhibitory concentration) was calculated from equation of line obtained by plotting a graph of concentration versus % scavenging of DPPH.
Statistical analysis
All experiment results were analyzed and expressed statistically by Dunnet’s multiple comparison test and analysis of variance (ANOVA).

Results and Discussion

Anti-inflammatory activity
The effect of extract of L. sativa L (400 mg/kg) on paw edema showed that it has maximum inhibitory effect during second phase and showed its effect after half an hour after the injection of formalin with good anti-inflammatory property (P<0.05) in dose dependent pattern (Table 1). Indomethacin showed its significant inhibitory action towards early phase also. The extract (200 mg/kg) showed its inhibitory action towards the later phase and maintained up to 5 hours, which might be due to inhibition of prostaglandin-mediated inflammation.

<table>
<thead>
<tr>
<th>Group</th>
<th>0 min</th>
<th>30 min</th>
<th>60 min</th>
<th>120 min</th>
<th>180 min</th>
<th>240 min</th>
<th>300 min</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>0.226±0.02</td>
<td>0.274±0.019</td>
<td>0.336±0.011</td>
<td>0.374±0.011</td>
<td>0.394±0.011</td>
<td>0.428±0.013</td>
<td>0.464±0.017</td>
</tr>
<tr>
<td>Standard (indomethacin)</td>
<td>0.256±0.026</td>
<td>0.274±0.011</td>
<td>0.294±0.019</td>
<td>0.332±0.013</td>
<td>0.354±0.024</td>
<td>0.378±0.008</td>
<td>0.394±0.011</td>
</tr>
<tr>
<td>Test 1 (200 mg/kg)</td>
<td>0.24±0.016</td>
<td>0.276±0.013</td>
<td>0.314±0.011</td>
<td>0.33±0.007</td>
<td>0.358±0.008</td>
<td>0.37±0.007</td>
<td>0.384±0.013</td>
</tr>
<tr>
<td>Test 2 (400 mg/kg)</td>
<td>0.234±0.0009</td>
<td>0.25±0.07</td>
<td>0.278±0.008</td>
<td>0.292±0.008</td>
<td>0.316±0.0089</td>
<td>0.332±0.008</td>
<td>0.356±0.013</td>
</tr>
</tbody>
</table>

Evaluation and optimization of microspheres
Box-Beihken of three factors in three level namely amount of polymer, probe sonication time and surfactant concentration was performed using Minitab 16 in response to encapsulation efficiency and particle size. The independent parameters selected for optimization were concentration of surfactant, amount of polymer and sonication time. The dependent parameters were entrapment efficiency and particle size. Thus formed microspheres prepared from solvent evaporation method were solid, spherical and greenish in color. The results obtained during evaluation of particle size and entrapment efficiency showed that it is higher when the particle size is also big.

The counter plot indicates that with the increase in amount of polymer the particle size also increased and with increase in sonication time particle size

<table>
<thead>
<tr>
<th>Batch no.</th>
<th>Amount of polymer (mg)</th>
<th>Sonication time (sec)</th>
<th>Surfactant Conc. (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>ML1</td>
<td>175</td>
<td>20</td>
<td>1.5</td>
</tr>
<tr>
<td>ML2</td>
<td>250</td>
<td>10</td>
<td>1.5</td>
</tr>
<tr>
<td>ML3</td>
<td>175</td>
<td>20</td>
<td>1.5</td>
</tr>
<tr>
<td>ML4</td>
<td>175</td>
<td>20</td>
<td>1.5</td>
</tr>
<tr>
<td>ML5</td>
<td>250</td>
<td>20</td>
<td>0.5</td>
</tr>
<tr>
<td>ML6</td>
<td>100</td>
<td>30</td>
<td>1.5</td>
</tr>
<tr>
<td>ML7</td>
<td>175</td>
<td>10</td>
<td>2.5</td>
</tr>
<tr>
<td>ML8</td>
<td>175</td>
<td>30</td>
<td>2.5</td>
</tr>
<tr>
<td>ML9</td>
<td>250</td>
<td>30</td>
<td>1.5</td>
</tr>
<tr>
<td>ML10</td>
<td>100</td>
<td>10</td>
<td>1.5</td>
</tr>
<tr>
<td>ML11</td>
<td>250</td>
<td>20</td>
<td>2.5</td>
</tr>
<tr>
<td>ML12</td>
<td>100</td>
<td>20</td>
<td>0.5</td>
</tr>
<tr>
<td>ML13</td>
<td>100</td>
<td>20</td>
<td>2.5</td>
</tr>
<tr>
<td>ML14</td>
<td>175</td>
<td>10</td>
<td>0.5</td>
</tr>
<tr>
<td>ML15</td>
<td>175</td>
<td>30</td>
<td>0.5</td>
</tr>
</tbody>
</table>

Anti-anxiety activity
Extract at doses 200 and 400 mg/kg (P<0.001) very significantly increased the number of entry and time in the open arm with decrease in closed arm when compared to the control treated group (Figure 1). In elevated plus maze mice normally prefers closed arm but this preference appears to reflect an aversion towards open arm generated by fear of open spaces.

<table>
<thead>
<tr>
<th>Group</th>
<th>Time spent in open arm (min)</th>
<th>Inhibition (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>3.216±1.2687</td>
<td>0.0</td>
</tr>
<tr>
<td>Standard</td>
<td>6.916±0.9386</td>
<td>109</td>
</tr>
<tr>
<td>Test A</td>
<td>7.3±0.97</td>
<td>127</td>
</tr>
<tr>
<td>Test B</td>
<td>7.78±0.8578</td>
<td>141</td>
</tr>
</tbody>
</table>

Table 1: Various sizes of edema expressed ad Mean ± SD vs. doses of standard and extract

Table 2: Time spent in open arm by control vs. standard and test samples

Table 3: Box Behnken design for formulation of microspheres

https://www.nepjol.info/index.php/JNCS
decreased (Fig 1). The P value < 0.05 was obtained for polymer, surfactant polymer interaction and surfactant concentration which indicates that these parameters significantly affect the particle size. Counter plot showing that entrapment efficiency is higher i.e. >70% when the polymer concentration is high and it also increases with increase in surfactant concentration but this is possible only at definite amount of drug (Fig 2). The p value <0.05 is obtained for polymer polymer interaction, surfactant surfactant interaction, surfactant polymer interaction and surfactant sonication time interaction. This suggests that above mentioned interactions significantly affect the entrapment efficiencies. Higher entrapment efficiency was observed with high surfactant concentration and high polymer. It was also seen in case of high surfactant and low sonication time.

**Contour plots of particle size**

**Figure 1: Effects of surfactant concentration, amount of polymer and sonication time on particle size**

**Contour plots of entrapment efficiency**

**Figure 2: Effects of surfactant, polymer and sonication time on entrapment efficiency**

Depending upon the optimization plot the microspheres were developed with particle size 140 µm. Since the desired particle size was not obtained therefore microspheres were developed by slightly increasing the sonication time and keeping all other parameters constant. Finally the microsphere with nearly optimum size was obtained i.e. 120 µm. Entrapment efficiency was calculated as follow:

\[
\text{Entrapment efficiency} = \frac{\text{Observed concentration}}{\text{Theoretical concentration}} \times 100\%
\]

\[
= \frac{0.2585}{0.33} \times 100\%
\]

\[
= 78.35\%
\]

**Drug release kinetics**

**Figure 3: Optimization plot of microsphere**

**Figure 4: Zero-order release profile of optimized microsphere loaded gel**

https://www.nepjol.info/index.php/JNCS
From the $R^2$ values obtained in different mathematical model, we conclude that optimized microsphere loaded formulation correlates with Higuchi model. Therefore the formulation follows Higuchi model release.

**Drug release behavior of crude extract and optimized microsphere-loaded gel**

a) The release profile of the crude extract was tested by loading the weighed amount of extract into cellophane membrane and introducing it to USP dissolution apparatus (Paddle type) at 37± 0.5 °C and 50 rpm using 900 mL phosphate buffer pH 6.8 (Fig 7).

b) The release profile of the formulation was tested by loading the formulation into cellophane membrane and introducing it to USP dissolution apparatus (Paddle type) at 37± 0.5 °C and 50 rpm using 900 mL phosphate buffer pH 6.8 (Fig 8 and 9).

**Antioxidant activity**

Anti-oxidant activity of crude extract and microsphere-loaded gel was calculated using IC$_{50}$ value. The IC$_{50}$ value of reference standard ascorbic acid in methanol was found to be 5.62 µg/mL. Methanolic extract of *L. sativa* L and optimized microsphere (developed from *L. sativa* L extract) loaded gel showed effective antioxidant activity with IC$_{50}$ value of 6.34 µg/mL and 8.25 µg/mL. This study revealed that *L. sativa* L crude extract as well as formulation possess significant anti-oxidant activity and this may be due to presence of various bioactive component present in the plant, which was evaluated during phytochemical screening.
The significant inhibitory activity shown by the extract of *L. sativa* L (200 and 400 mg/kg) over a period of 6 h in formalin-induced inflammation was quite similar to that exhibited by the group treated with diclofenac diethylamine (Table 4). This shows the formulation has potential anti-inflammatory action (P< 0.05). These results indicate that the formulation acts in later phases in dose dependent manner, probably involving arachidonic acid metabolites, which produce an edema dependent on neutrophils mobilization. The early phase (1-2 h) of the model is mainly mediated by histamine, serotonin and increased synthesis of prostaglandins in the damaged tissue surroundings. The late phase is sustained by prostaglandin release and mediated by bradykinin, leukotrienes, polymorphonuclear cells and prostaglandins produced by tissue macrophages.

**Figure 10:** Antioxidant activity of crude extract and optimized microsphere loaded gel

### Anti-inflammatory activity of optimized microsphere loaded gel

The anti-inflammatory effect of the extract observed might be due to the presence of flavonoids in the plant [14].

There has been an increasing amount of drug discovery and drug formulations development from natural compounds in the past days, which can be attributed to development of modern technologies [15]. The herbal formulations are now available into various novel drug delivery system like polymeric nanoparticles, nanocapsules, liposomes, nanoemulsions etc [16]. The use of novel formulations like microsphere can overcome the limitation of conventional formulations like solubility, bioavailability, toxicity and confer advantages like increased stability, pharmacological activity, tissue macrophages distribution and patient compliance. It can further help for the targeted delivery of drugs to desired site of action [16,17]. Anxiety disorders are the most common psychiatric illnesses in the world. Hydro-alcohol extract of *L. sativa* rich in polyphenols possess potent anxiolytic property in pervious investigations. The present result showing that, *L. sativa* includes anti-inflammatory action, anxiolytic activity and anti-oxidant activity development of novel herbal formulation which aims to develop *L. sativa* microsphere and dispersion of optimized microsphere into gel base to produce sustained release anti-inflammatory action. The study also summarizes the method of preparation, characterization of formulation, type of various ingredients, particle size, entrapment efficiency, route of administration, drug-release kinetics, In-vitro release studies, ex-vivo permeation studies and evaluation of biological activity in animal model. Thus, *L. sativa* is found to have potent anti-inflammatory, anxiolytic and antioxidant action.

### Table 4: Summary of sizes of oedema at different time vs. doses of *L. sativa* L & diclofenac gel

<table>
<thead>
<tr>
<th>Group</th>
<th>0 min</th>
<th>30 min</th>
<th>60 min</th>
<th>120 min</th>
<th>180 min</th>
<th>240 min</th>
<th>300 min</th>
<th>360 min</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>0.22±</td>
<td>0.25±</td>
<td>0.326±</td>
<td>0.352±</td>
<td>0.396±</td>
<td>0.42±</td>
<td>0.434±</td>
<td>0.444±</td>
</tr>
<tr>
<td></td>
<td>0.0114</td>
<td>0.013</td>
<td>0.0114</td>
<td>0.0148</td>
<td>0.0114</td>
<td>0.007</td>
<td>0.011</td>
<td>0.0152</td>
</tr>
<tr>
<td>Standard</td>
<td>0.236±</td>
<td>0.26±</td>
<td>0.276±</td>
<td>0.288±</td>
<td>0.268±</td>
<td>0.256±</td>
<td>0.25±</td>
<td>0.248±</td>
</tr>
<tr>
<td></td>
<td>0.021</td>
<td>0.021</td>
<td>0.024</td>
<td>0.013</td>
<td>0.021</td>
<td>0.082</td>
<td>0.012</td>
<td>0.014</td>
</tr>
<tr>
<td>Test 1</td>
<td>0.224±</td>
<td>0.248±</td>
<td>0.27±</td>
<td>0.28±</td>
<td>0.272±</td>
<td>0.26±</td>
<td>0.256±</td>
<td>0.246±</td>
</tr>
<tr>
<td>(2% gel)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>0.033</td>
<td>0.0319</td>
<td>0.038</td>
<td>0.0418</td>
<td>0.042</td>
<td>0.043</td>
<td>0.042</td>
<td>0.0364</td>
</tr>
<tr>
<td>Test 2</td>
<td>0.232±</td>
<td>0.266±</td>
<td>0.282±</td>
<td>0.286±</td>
<td>0.284±</td>
<td>0.258±</td>
<td>0.248±</td>
<td>0.238±</td>
</tr>
<tr>
<td>(4% gel)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>0.039</td>
<td>0.033</td>
<td>0.117</td>
<td>0.037</td>
<td>0.040</td>
<td>0.044</td>
<td>0.038</td>
<td>0.039</td>
</tr>
</tbody>
</table>
optimized formulation showed positive result for sustained releases anti-inflammatory action when the study was conducted in animal model [5, 18]. In the present study we put in correlation of L. sativa grown in Nepal which was rich in phytochemicals such as cardiac glycoside, reducing sugar, saponin, coumarin and flavone, greatly contribute to enhance the inflammatory based diseases. This hypothesis is also enforced for a great therapeutic potential.

**Conclusion**

This study conclusively shows that, glycoside and phenolics enriched extract of *Lactuca sativa* and its microsphere loaded sustained release gel possesses anti-inflammatory, anxiolytic and anti-oxidant activity.

**Acknowledgements**

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**References**


