Synthesis of Some 3d Metal Complexes of Quinine and Their Toxicological Studies

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Abstract

Four metal complexes of antimalarials cinchona alkaloids; $[Co(Quin)Cl_3]$ -1, $[Fe(Quin)Cl_2H_2O]SO_4.3H_2O$ -2, $[Cu(Quin)ClSO_4]\infty$ -3 and $[Cd(Quin)ClSO_4]\infty$ -4 have been synthesized and characterized by elemental analysis, conductivity, magnetic moment, IR, UV-Vis and mass spectroscopic techniques. The tetrahedral geometry is assigned for all the four complexes and the quinine ligand is protonated at quinuclidine N atom with the coordination of the metal at quinoline N atom for all the complexes. Complexes 1 and 2 have zwitterionic structures while complexes 3 and 4 are polymeric in nature with zig-zag extended chains (MCl- O -SO₂- O -MCl- O -SO₂---).

Stability constant of the complexes were evaluated for the metal salts. The value of the stability constant (β) was found to be log 10.48, 9.92, 7.98 and 7.81 for Cu(II), Fe(III), Co(II) and Cd(II), respectively and the order of β was found to be Cu(II)>Fe(III)>Co(II)>Cd(II). The stability constant data revealed that concomitantly administration of quinine with preparation containing these metals may reduce their efficacy. Toxicological study was carried out by investigating the effect of administration of the complexes on alkaline phosphatase activity of kidney, liver and serum of Albino rats. All the examined four metal complexes were found to be non-toxic.

Keywords: Quinine, stability constant, FTIR, MS, phosphates activity.

Introduction

The first attempt at the treatment of malarial was in the 17th century with the use of alkaloids obtained from the bark of the cinchona tree. The French chemists Pierre Pelletier and Joseph Coventou in 1820 isolated and identified the basic alkaloids from the bark of cinchona tree to be quinine. It still remains an active member of the anti-malarial drugs¹. Beside its pharmacological activity, quinine is used as a chiral resolving agent and the activity in this area did not seem to slow down². Meanwhile, many synthetic antimalarials

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drugs have been discovered over the years for the treatment of malarial disease like chloroquine, sulphadoxine and pyrimethamine being among the most effective³. However, malarial parasites resistant to these drugs are now widespread in America, Asia and Africa.

Resistance to antimalarial drugs first to chloroquine and then to others was first noticed in the 1950s and since then, it has spread all over the world. Resistance of *Plasmodium falciparum* to chloroquine has become a major health concern of developing world. Therefore, it becomes highly necessary to come up with alternative antimalarial drugs with different structures and mode of action to deal with the development of resistance to the drugs in current use⁴. It is in line with this goal that this study based on the synthesis of metal complexes of selected known antimalarial drug has been undertaken. The synthetic strategy involves enhancing the activity of antimalarial drugs through the incorporation of transition metals into their molecular structures⁵.

In continuation of our efforts⁶⁻¹⁰ to search for novel chemotherapeutic drugs against the resistant strains of *Plasmodium falciparum*, we reported the synthesis, characterization, toxicological study and stability constant of Co(II), Fe(III),Cu(II) and Cd(II) complexes of quinine in this work.

Experimental Methods

Quinine hemi-sulfate was purchased from Sigma Chemical Co, (USA). All solvents and other reagents were of high purity (Aldrich and Sigma) and were used without further purification. CuCl₂.2H₂O, CdBr₂.4H₂O, CoCl₂.6H₂O and FeCl₃.6H₂O were used as metal ion sources.

IR spectra of the samples in KBr pellets were obtained in the ranges of 4000-400 cm⁻¹ using FTIR spectrometer. Metal Analyses were determined by atomic absorption spectroscopy with Perkin-Elmer Spectrometer, model 3110. The analyses of carbon, hydrogen and nitrogen were carried out on a Perkin-Elmer 204C microanalyzer. Mass spectra recorded on a micromass platform spectrometer and kratos concept ARE instrument using DMSO as solvent. UV-Vis spectra were obtained on Aquamate v4.60 spectrophotometer. Magnet susceptibility measurement of the complexes was determined at room temperature by the Gouy method¹¹. Mercury tetrathiocynatocobaltate(II) $\{Hg[Co(NCS)_4]\}$ was used as calibrant. The following four compounds were synthesized.

Compound-1 [Co(QUIN)Cl₃]: A solution of CoCl₂.6H₂0 (0.236 g, 1 mmol) in ethanol (15 ml) was added to a solution of quinine hemi-sulfate (0.373 g, 1 mmol) in a 15 ml mixture of methanol/ 0.01MHCl (2:1). The mixture was stirred and heated at 50[°]C for three hours, during which a blue precipitate was rapidly formed. Upon cooling to room temperature the mixture was filtered and the isolated solid was recrystallized by slow concentration of a solution in methanol/butanol to give blue crystals. The complex was formulated as [Co(Quin)Cl₃]. Yield: 72 %, mol.wt. = 490.43, M.P. = 250[°]C; anal.calcd. for $C_{20}H_{25}Cl_2N_2O_2CoCl_3$): C, 49.00, H. 5.10; N, 5.70, Co, 12.08; found: C, 50.16; H, 5.19; N, 5.61; Co, 12.11; IR (KBr,cm⁻¹) 3390(br), 2930, 2600, 1621, 1592, 1516, 1228,1100, 629, 462; UV-Vis (CH₃OH) λ (nm): 490, 595, 668.

Compound-2 [Fe(Quin)Cl₂H₂O]SO₄.3H₂O: Quinine hemi-sulfate (0.373 g, 1 mmol) was dissolved in 10 ml of ethanol followed by slow addition of FeCl₃.6H₂O (0.270 g, 1 mmol) in 10 ml of ethanol. The resulting brown solution was refluxed for 1hour and left for 2 days. The brown precipitate formed was filtered under vacuum and washed twice with ethanol and dried in a desiccator. A brownish crystalline powder obtained was formulated as [Fe(Quin)Cl₂ H₂O]SO₄.3H₂O; Yield: 64 %, mol.wt.= 618, M.P.= 250°C; anal. calcd. for C₂₀H₃₁N₂O₁₀FeCl₃.SO₄]: C,38.8; H,5.34; N,4.53; Fe, 9.14. found: C, 39.26; H, 5.06; N, 4.61; Fe 9.18.; IR (KBr, cm⁻¹); 3421, 2934, 2645, 1620, 1602, 1542, 1142, 1076, 994, 853, 603; UV-Vis (CH₃OH) λ (nm): 353,635; MS (positive Cl-methane): m/z, (Relative intensity): 97(88), 136(50), 188(23), 217(12), 251(13), 279(10), 309(28), 325(26), 367(4), 403(2), 445(1), 474(0.5).

Compound-3 [*Cu* (*Quin*)*Cl* SO₄]_{∞}: CuCl₂. 2H₂O (0.170 g, 1 mmol) was added to 30 ml of ethanolic solution of Quinine hemi-sulfate (0.740 g, 2 mmol). The mixture was stirred and heated at 50°C for two hours and cooled to room temperature. The resulting solution was filtered. Diethyl ether was added to the filtrate until the solution became turbid, green colored crystals deposited after 2 days which were filtered, washed with ethanol and diethyl ether and dried under vacuum. The product was formulated as [Cu(Quin)Br SO₄] ∞ . Yield: 57%, mol.wt.=520,M.P.=280°C; anal. calcd. for C₂₀H₂₅N₂O₆SCuCl: C, 45.91; H, 4.85; N,5.40;Cu,12.60:found:C,46.10;H,4.81;N,5.38;Cu,12.21.; IR(KBr, cm⁻¹): 3390, 3117, 2978, 2658, 1642, 1621, 1607, 1247, 1130, 1047, 852, 619, 463, 432 .UV-Vis (DMSO) λ (nm): 912.

Compound-4 [*Cd*(*Quin*)*Cl SO*₄]_{∞}: To a solution of quinine hemi-sulfate (0.782 g, 2 mmol) in methanol (10 ml), a solution of CdBr₂.4H₂O (0.688 g, 2 mmol) in methanol (10 ml) was added. The mixture was stirred and heated for 60 minutes at 60°C and cooled to room temperature. The resulting turbid solution was filtered. By slow evaporation of the filtrate at room temperature, colorless crystals were obtained after 3 weeks. The product was formulated as [Cd(Quin)ClSO₄]_{∞}. Yield: 57 %, mol.wt. = 613, M.P.= 245°C, anal. calcd. for C₂₀H₂₅N₂O₆SCdBr: C, 39.15; H, 4.08; N. 4.57; Cd, 18.28. found: C, 38.28; H, 4.23; N,4.45; Cd, 18.17; IR (KBr, cm⁻¹): 3411, 3002, 2664, 1622, 1592, 1518, 1365, 1207, 1097, 620, 464, 432. UV-Vis (DMSO) λ (nm): 221,277,320.

Complexes of quinine with metal ions were studied in solution using ethanol as a solvent, in order to determine [M/L] ratio. A series of solutions were prepared having a constant concentration (10^{-3} M) of metal ions and quinine. The results of complexes in ethanol suggested that the metal to ligand ratio was 1:1 for all complexes which were similar to that obtained from solid state study.

The stability constant of metal ions (Cu^{2+} , Fe^{3+} , Cd^{2+} and Co^{2+}) with quinine ligand were determined spectrophotometrically using procedure of Hilderbrand and Benesi¹² as modified by previous workers^{13,14}. A series of seven solutions were prepared with constant concentration of metal ion [Mo] and variable ligand [Lo] concentration at PH 7.4 with ionic strength (0.1 M KN0₃). The reaction mixture was stirred continuously and allowed to stand for 15 minutes. The absorbance of each of the mixture was taken at λ_{max} previously determined using UV-Vis spectrophotometer. The stability constant β was determined using Hilderbrand and Benesi equation¹² Mo/A= $1/\beta\epsilon c (1/Lo) + 1/\epsilon c$. Plot of Mo/A versus 1/Lo gave slope $1/\beta\epsilon c$ from which β (Stability constant) can be evaluated.

Investigation of the hazardous effects of the complexes on albino rats were carried out as described here. A total of 36 male *albino* rats of average body weight of 150 g were obtained from Biochemistry Department of University of Ilorin. The rats were fed for 2 weeks prior to their usage. The rats were divided into 6 groups (6 rats each). Group-1 {control}; Group-2 {Quinine}; Group-3 $\{Co(Quin)Cl_3\};\$ Group- $4\{[Fe(Quin)Cl_2H_2O)]SO_4.3H_2O\};$ Group-5 $\{ [Cd(Quin)ClSO_4] \infty \}$ Group-6 and $\{ [Cu(Quin)ClSO_4] \infty \}$. The rats were injected two times daily for five days at dosage level of 0.640 mg/150 g Quinine. The drugs were dissolved in DMSO administered orally to the rats in proportion to their body weight. The control groups were given only DMSO.

The rats were sacrificed at 24 hours after the last day of administration of drugs by anaesthetizing them in a glass jar containing cotton wool soaked in chloroform. The blood was collected into clean labeled sample bottles and allowed to coagulate. They were then centrifuged and the serum pipetted out for analysis. The liver, heart, and kidney were quickly removed from the sacrificed rats into ice-cold 0.25 M sucrose solutions. The tissues were then cleaned and dried. The weights of tissue were taken and a known weight of the liver and small intestines were cut out with a clean scissors. The weighed tissues were placed in a mortar and homogenized using the pestle with ice-cold sucrose solution to give a final volume of five times the original tissue weight (1.5w/v). The homogenates were kept in well labeled container and stored in the freezer for further use.

The amount of phosphate ester that is split within a given period of time is a measure of the phosphatase enzyme activity. The method of Bassey et al.¹⁵ as modified by Wright and Plummer¹⁶ and Gornall et al.¹⁷ was employed in this assay. Tissue homogenates were incubated with p-nitrophenyl phosphate (PNPP) buffered at pH 10.1 for 10 minutes at 37°C. The hydrolysis product, p-nitrophenyl reacted with NaOH to give a yellow color, determined spectrophotometrically at 440 nm which is a measured of the enzyme activity. Two test tubes labeled blank and test were placed in the test tube rack. 2.2 ml of 0.1 M MgSO₄.7H₂O was added into the test tube. 0.2 ml distilled water and homogenate was added into blank and test tubes respectively. The mixture was shaken and incubated in water bath at 37°C for 10 minutes. 0.5 ml, 19 mM (PNPP) was added into each of the test tubes and incubated in water bath at 37°C for another 10 minutes. Finally, 2 ml of 1 M NaOH was added into the test-tubes and absorbance of test against blank was taken at 0, 1, 2 and 3 minutes in 400 nm.

The activity of ALP in the sample was calculated using the following equation (1).

Activity
$$(mM/min/ml) = \frac{\Delta E/min \times 1000 \times V \times F}{18.8 \times V \times 1}$$
 (1)

where, $\Delta E/min =$ change in optical density per minute; V is total reaction volume; F is dilution factor; V is volume of the enzyme source; 1 is light path (1cm); 1000 is factor introduced so that enzyme activity can be measured in mM/min/ml and 18.8 is extinction coefficient of 1 µmol of p– nitro-phenol in an alkaline solution of 1 ml volume and 1cm light path at 400 nm.

Specific activity (mM/mg protein/min) =
$$\frac{\operatorname{activity(mM/min/ml)}}{\operatorname{protein conc.(mg/ml)}}$$
 (2)

Statistical Analysis was determined using Duncan multiple Range Test and values were considered.

Results and Discussion

The Fe(III), Co(II), Cu(II) and Cd(II) complexes of quinine were synthesized by reaction of metal salts with quinine. The complexes were characterized by elemental analysis, Conductivity, Magnetic moment, infrared, UV-Vis spectroscopy. [Fe(Quin)Cl₂ H₂O]SO₄. 3H₂O was further characterized by mass spectroscopy. The elemental analysis results of the complexes are in good agreement with the formula: [Co(Quin)Cl₃], [Cu(Quin)Cl SO₄] ∞ , [Fe(Quin)Cl₂ H₂O]SO₄.3H₂O and [Cd(Quin)ClSO₄] ∞ . The complexes are generally soluble in methanol, DMSO but insoluble in non-polar organic solvent.

The molar conductance values measured in DMSO solution (10⁻³ M) for these complexes are in 96-116 λ^{-1} cm² mol⁻¹ range (Table 1). According to these results, the complexes are electrolytes. Determination of stoichiometric ratio using job's method suggested mole ratio 1:1 metal to ligand stoichiometry for the complexes. The proposed structures are presented in figs 1-3. Fe(III) and Co(II) complexes formed Zwitterionic structures. The structures of these complexes may be compared with a molecular Lhistidinum trichlorozincate (L-histidine \hat{H}^+) ZnCl₃¹⁸. The quinine ligand is protonated at quinuclidine N atom to form a Zwitterionic quinine ligand¹⁹, with coordination of the metal at quinoline N atom and tetrahedral structures were proposed for the complexes (Figs 1 and 3). Cd(II) and Cu(II) quinine complexes are iso-structural with polymeric zinc reported previously. They formed polymeric complexes with quinine (Fig. 2). The complexes exhibit zig-zag extended chains-MCl-O-SO₂-O-MCl-O-SO₂-O-. The structures of these complexes are iso-structural with $[Zn(Quin)ClSO_4] \infty$ as reported earlier in our previous study⁶. They formed tetrahedral geometries with each metals ion centre coordinated to an oxygen atom from each of two sulfates bridge, Cl atom and quinoline atom N(4) of the quinine moiety. The second quinuclidine Nitrogen is protonated.

Complexes	Magnetic moment µ (B.M)	Conductivity (Ohm ⁻¹ cm ² mol ⁻¹)
Cd(QUIN)ClSO ₄)] _∞	Diamagnetic	116
[Fe(QUIN)Cl ₂ .H ₂ O]SO ₄ .3H ₂ O	5.84	102
[Co (QUIN) Cl ₃]	4.50	87
$[Cu(QUIN)Cl(SO_4)]_{\infty}$	1.35	108

Table 1: Magnetic susceptibility & conductivity of quinine complexes in DMSO

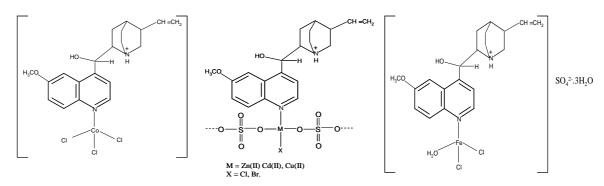


Figure 1: Proposed structureFigure 2: Proposed structureFigure 3: Proposed structure of $of Co(QUIN)Cl_3$. $of M(QUIN)ClSO_4]_{\infty}$. $Fe(QUIN)Cl_2H_2O]SO_4.3H_2O$.

The UV-Vis spectra of the quinine ligand and its complexes are shown in Table 2. The UV-Vis spectrum of the quinine ligand shows two intense bands at 277 and 340 nm, the bands are assigned to $\pi \to \pi^*, n \to \pi^*$ transition over the whole conjugated system of the ligand¹⁹. The same peaks are obtained in the complex Cd(II)Quin with spectroscopic term. The cadmium ion has its 5d orbital completely vacant and hence ligand metal $(L \rightarrow M)$ binding can take place by the acceptance of lone pair of electrons from the donor nitrogen atom of the ligand²⁰. Extra bands observed for Co(II), Fe(III) and Cu(II) complexes are attributed to d-d transition. The intense blue color of the Co(II) complex suggests a tetrahedral geometry²⁰. The d-d spectrum of the Co(II)Quin Complex shows band around (20408 cm⁻¹) 490nm, (16807cm⁻¹) 595 nm and (14970 cm⁻¹) 668 nm, suggesting existence of ${}^{4}A_{2} \rightarrow {}^{4}T_{2}$, ${}^{4}A_{2} \rightarrow {}^{4}T_{1}$ and ${}^{4}A_{2} \rightarrow {}^{4}T_{1}$ (P) transitions with tetrahedral configuration²¹. Fe(III) is a moderately oxidizing ion and many of its complexes exhibit $n \rightarrow n^*$ and ligand to metal charge transfer transition $(L \rightarrow MCT^{19})$. For the Fe(III)Quin, these bands are observed at (44248 cm⁻¹) 226 nm, (26455 cm⁻¹) 378 nm. The electronic spectra of Cu(II)Quin exhibited only one broad d-d transition at (10965 cm⁻¹) 912 nm assigned to the ${}^{2}T_{2} \rightarrow {}^{2}E$ transition which is in conformity with tetrahedral configuration²². Though three transitions are expected in this case they are very close in energy and often appear in the form of one broad band envelop 23 .

The IR spectra of the complexes in the 4000-400 cm⁻¹ were compared with that of the ligand. The latter presents a broad band 3373-3100 cm⁻¹ due to vOH/H₂O stretching vibration this is attributed to vOH with OH bending vibration located at 1620 and 1640 cm⁻¹. The band became strong and intensified at 3390 cm⁻¹ in Co(II), 3390 cm⁻¹ in Fe(III)Quinine complexes. The coordinated bond between water molecules and cation results in the appearance of a vibrational band around 650-850 cm⁻¹ (vM-OH₂) in Fe(III)Quinine complex^{18, 24, 25}. The absorption band around 2580 cm⁻¹ due to vNH₂⁺ observed in free ligand is also present in all the complexes, with little shift due to hydrogen bonding indicating that the protonated state of quinuclidine nitrogen remains upon complexation. This shift is probably due to involvement of vC=N quinoline group in complex formation. Shift of the frequency to higher and lower frequency side as well as no change; have also been reported in the literature²⁶. Disappearance of the bands at 1345 cm⁻¹ and the observation of the bands of the SO₄²⁻ group due to S-O stretching at 1100-977 cm⁻¹ in the complexes indicate the

coordination of SO_4^{2-} to the metal²⁷. These frequencies agree with values obtained for C2v, bridging sulfate group complex. The bands in the range 600-618 cm⁻¹ are assigned to v(M-N) vibration modes²⁷.

The mass spectra, relevant m/z ratios and tentative assignment of $[Fe(Quin)Cl_2H_2O]SO_4.3H_2O$ are shown in Table 3 and Fig. 4. The peak at m/z 325 is assigned to quinine. Loss of CH₃ from m/z 325 gave fragment ion at m/z 309. There was loss of quinoline from m/z 325 resulting in the formation of fragment ions m/z 136, whereas loss of CH₃O produced the peak m/z 279 from m/z 324. The peak observed at m/z 97 was due to loss of (CH=CH₂) from m/z 136. The peak at m/z 367 corresponds to Fe C₁₉H₂₄N₂O₂ and m/z 403 (FeC₁₉H₂₄N₂O₂Cl) confirms the formation of complex as supported by other analyses.

		$Fe(QUIN)Cl_2 H_2 O_1 SO_4 . SH_2 O_1 SO_4 . $				
Complexes/ligand	λmax	Energies	Assignments	M/z	Relative Intensity (%)	Assignment
	(<i>nm</i>)	(cm^{-1})	-	325	25	M.H*
Quinine hemi-sulfate	277	36101	$\pi ightarrow \pi^*$	309	28	M-CH.
	340	29412	$n ightarrow \pi^*$			
$[Cd(QUIN)ClSO_4)]_{\infty}$	221	45249	$\pi ightarrow \pi^*$	136	49	
	277	36101	$n \rightarrow \pi^*$			+ N
	320	31250	MLCT			II CH ₂
[Fe(QUIN)Cl ₂ .H ₂ O]	226	44248	$\pi \rightarrow \pi^*$	97	88	<u> </u>
SO ₄ .3H ₂ O	378	26455	MLCT	21	00	
[Co (QUIN) Cl ₃]	490	20408	${}^{4}A_2 \rightarrow {}^{4}T_1(P)$			Т сн,
	595	16807	${}^{4}A_{2} \rightarrow {}^{4}T_{1}$	367	5	[Fe(H ₂ O) M-CH ₃ O]
	668	14970	${}^{4}A_{2} \rightarrow {}^{4}T_{2}$			
$[Cu(QUIN) Cl(SO_4)]_{\infty}$	912	10965	$^{2}T_{2} \rightarrow ^{2}E_{2}$	403	1	[Fe(H ₂ O)[Cl] (M-CH ₃ O)
				L I		

Table 2: UV-Vis. Spectra of quinine and its complexesTable 3: Mass spectra of
Fe(OUIN)Cl2 H2O1SO4.3H2O

Magnetic moments of the complexes are given in Table 1. Examination of these data reveals that magnet moment of 0.2 B.M for Cd(II) complex confirms that the complex is essentially diamagnetic. The magnetic moment found for Co(II), Cu(II) complexes are 4.50 and 1.35 B.M, respectively, and these values suggest tetrahedral structure which is in good agreement with data of electronic transition as discussed above.

The results of stability constants and hence reactivity of the quinine ligand metal ions follow the trend Cu(II) > Fe(III) > Co(II) > Cd(II). The stability constant was log 10.48, 9.92, 7.98 and 7.81. The stability constants were found to be inversely proportional to the ionic radii of metals. The order indicates that β values increase with decreasing ionic radius of the metals. The gradual decrease clearly showed that Irving and William²⁸ rule is strictly obeyed for all the complexes in solution. The most stable are the Cu(II) complexes and this is due to Jahn-Teller distortion. The Cd(II) complex is least stable and the reason for this is consistent with the fact that their CFSE =0. This finding is in agreement with the literature²⁹. The data also reveals that Fe³⁺ formed more stable complex with quinine compared to other metal ions except Cu(II). The higher stability of Fe³⁺ complex of the drugs may be explained on the basis of difference in charge of the metal ions. The high positive charge on the iron

permits a closer approach of the ligand and better electrostatic attraction. This results in the formation of more stable complex with the ligand. However, a general rule is that trivalent metals form strong bonds than divalent and monovalent ions³⁰. The stability constant data revealed that concomitantly administration of quinine with preparation containing these metals may reduce drug efficacy.

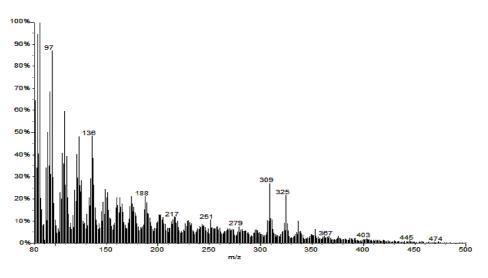


Figure 4: Mass spectra of *Fe*(*QUIN*)*Cl*₂*H*₂*O*]*SO*₄.3*H*₂*O*

The values of alkaline phosphatase activities (ALP) in the kidney, liver and serum following the administration of quinine and its metal complexes are shown in Table 4. The serum ALP activity showed no significant change (P>0.05) on comparison with one another and control after repeated administration of quinine and its metal complexes. However, the kidney ALP activity was significantly increased (P < 0.05) in all the groups when compared with control. The trend presented by this result may be attributed to the fact that these drugs and their metal complexes imposed certain cellular stress on the cells of these tissues and in order to offset this stress (induced by the drugs and complexes) tissues cells switched to increased synthesis or production of this enzyme protein, hence the increased cellular ALP activity. Akanji and Nlumanze³¹ observed increased activities of various enzymes under varying condition of stress. This may account for the repeated administration of these drugs and their complexes. Alkaline phosphatase is a membrane bound protein and marker enzyme for the plasma membrane and endoplasmic reticulum as reported by Wright and Plummer¹⁶. Damage to tissue cell plasma membrane is often characterized by leakage of membrane components into the extracellular fluid³². The non-significant change in ALP activity of the therefore is suggestive of a non – damaging effect on the plasma membrane of liver and kidney cells but a possible induction of cellular stress as corroborated by the increased activity in all the groups under the period of investigation and doses administered. The complexes are generally less toxic than parent quinine ligand.

Ligand/Complexes	Kidney(IU/L)	Liver(IU/L)	Serum(IU/L)
Control	64.19+5.11	20.03+1.90	10.6+0.96
Quinine(Quin)	72.40+6.09	27.14+2.15	10.00 + 0.88
[Fe(QUIN)Cl ₂ .H ₂ O]SO ₄ .3H ₂ O	80.40 +7.83	30.42+2.73	8.76+0.57
[Co (QUIN) Cl ₃]	76.34+5.67	25.46+2.54	9.22 +0.59
$Cd(QUIN)CISO_4)]_{\infty}$	82.27+6.23	23.15+2.16	8.56+0.66
$[Cu(QUIN)Cl(SO_4)]_{\infty}$	73.44 +5.78	26.26 +2.88	9.82 +0.53

Table 4: Effects of administration of quinine and its complexes on alkaline phosphatase activities of rat liver, kidney and serum.

Values are mean of 5 determinations \pm SD. Enzyme activities are expressed in U/I/min/mg protein. Values carrying superscripts different from their parent ligand for each organ are significantly different (P< 0.05) Note Student's t-test and Duncan's multiple range test (Duncan, 1955) were carried out on the results using the computer software package SPSS version 12.

Conclusions

Co(II), Fe(III), Cu(II) and Cd(II) complexes of quinine drug are reported. The complexes were characterized by elemental analyses, magnet susceptibliity, electronic and IR spectra. The (Fe[Quin)Cl₂H₂O]SO₄.3H₂O was further elucidated by mass spectroscopy. All the complexes are found to have tetrahedral geometry. The stability constant data revealed that concomitantly administration of quinine with preparation containing these metals may reduce their efficacy. The complexes are generally less toxic than parent quinine ligand.

Acknowledgements

We are grateful for financial support from the Science and Technology Education Post Basic Project (Step B) and University of Ilorin. We also thank Dr. R. O. Arise, Department of Biochemistry, University of Ilorin, Nigeria for his assistance in interpretation of toxicology data.

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