Prevalence of Carbapenem-Resistant Gram-Negative Clinical Isolates and its Antibiotic Susceptibility Patterns

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Abstract

Carbapenem-resistant Gram-negative pathogens are recognized as a major health concern. However little information is available in Nepal regarding carbapenem-resistant Gram-negative bacteria. Therefore, a cross-sectional study was carried out at a tertiary care hospital to examine the prevalence of carbapenemase, metallo-beta-lactamases, and AmpC-beta-lactamases producing organisms in carbapenem-resistant Gram-negative bacteria. A total of 82 Gramnegative bacteria were identified from 825 clinical specimens based on colony characters, Gram staining catalase test, oxidase test, and other biochemical tests. Antibiotic susceptibility testing was performed according to clinical standard institute (CLSI) guidelines 2014. Of 82 isolates, 24 (29.3%) were carbapenemresistant. Of these 33.4% were E. coli, 25% were Pseudomonas, 25% were Klebsiella pneumoniae and 16.7% were Acinetobacter spp. A total of 13 (54.2%) were MBL producers. All MBL producers were multi-drug resistant (MDR). Single isolate of K. pneumoniae was KPC positive. Out of 24 carbapenemresistant isolates, 20.8% isolates were positive for the AmpC beta-lactamases test. All AmpC beta-lactamases and single isolates of KPC producers were MDR. Among the total of 82 Gram-negative bacterial isolates, 48.8% were MDR organisms. Based on in-vitro testing polymyxin B was found effective against all isolates. Although, MBL and other beta-lactamase producers were not higher than that found in other studies from Nepal, beta-lactamases and MDR organisms should be detected in the hospital setting to prevent the spread of these organisms.

Introduction

Antibiotic resistance is an important public health problem. Antibiotic resistance is now well recognized as a major problem in the treatment of infections in hospitals and the community. The World Health Organization identified antimicrobial resistance as one of the three greatest threats to human health (Davies *et al.*, 2010). Drug-resistant bacterial infections and their morbidity and mortality are on the rise all over the world (Bisht *et al.*, 2009).

Beta-lactams are the major antibiotics used in the world today. It comprises more than 65% of the total antibiotics consumed in the world today (Elander, 2003). Major mechanisms causing resistance to the beta-lactam antibiotics are the production of beta-lactamases, reduced outer membrane permeability, and altered affinity of target Penicillin-binding proteins. The infections due to beta-lactamase are difficult to treat because of the emergence of newer beta-lactamases such as Extended Spectrum beta-



lactamases (ESBL), AmpC beta-lactamases, and Carbapenemases. The beta-lactamases inactivate beta-lactam antibiotics by cleaving the structural beta-lactam ring. Failure to detect these enzymes producing strains has contributed to their uncontrolled spread in the Health Care setup and therapeutic failure (Parker *et al.*, 1990, Washington *et al.*, 2006).

Among all of the bacterial resistance problems, Gram-negative pathogens both fermentative and non-fermentative bacteria are the leading cause of human infection. Antibiotic-resistant pathogens are common in Nepal. This is due to the misuse of antibiotics. Many patients use antibiotics without a prescription from a physician and also some physicians prescribed antibiotics without the results of antibiotics susceptibility testing. Also, people fail to finish full course of treatment. circumstances, the improper dosing will fail to eliminate the disease agent and will furthermore: encourage the growth of most resistant strains. The study was mainly conducted to evaluate the burden of MDR with special reference to Gram-negative bacteria and to determine their antibiotic-resistant. MBL. AmpC beta-lactamase, and carbapenemase.

Materials and Methods

This was a cross-sectional study conducted at a tertiary care hospital in Kathmandu. Patients of

all age groups and both sexes were included in the study population after getting consent from the patient. Nonrepeating samples which were sent for routine culture and antibiotic susceptibility testing were processed.

A total of 82 Gram-negative bacteria were identified from 825 clinical specimens based on colony characters, Gram staining, catalase test, oxidase test, and other biochemical tests. Antibiotic susceptibility testing was performed according to clinical standard institute (CLSI) guidelines 2014. Confirmation of MBL, AmpC, and KPC are done according to clinical standard institute (CLSI) guidelines 2014.

Results

A total of 825 different clinical specimens were received for routine culture and susceptibility testing during the study period, of which urine (54.3%), blood (26.1%), and sputum (11.2%) were the common samples submitted for culture. Of the total clinical specimens, 120 (14.5%) specimens showed significant growth from which 124 bacterial isolates were obtained. Gram-negative bacteria were predominant constituting 82 (66.2%). Gram-positive bacteria constituted 42 (33.8%) of total isolates. Among isolates, Escherichia coli (50%) Gram-negative most frequently isolated, followed by *Pseudomonas* spp (15.8%)and Klebsiella pneumoniae (10.9%).

Table 1. Distribution of gram-negative bacteria among different clinical samples.

Clinical	Number of sample	E. coli	Pseudomonas	K. pneumoniae	Others*	Total
sample	(%) (n=825)					
Urine	448 (54.3)	29	8	6	6	49
Blood	215 (26.1)	4	0	1	2	7
Sputum	92 (11.2)	2	1	1	8	12
Tip	27 (3.3)	4	2	0	1	7
Pus	19 (2.3)	1	2	0	2	5
Body fluid	17(2)	1	0	1	0	2
Others	7 (0.8)	0	0	0	0	
Total	825	41	13	9	19	82

*Others bacterial isolates include; Citrobacter spp = 5, Acinetobacter spp=5, Proteus mirabilis= 3, P. vulgaris= 2, Moraxella spp =3, Salmonella spp=1

Multidrug resistant Gram-negative bacteria

Out of 82 Gram-negative bacterial isolates, 40 (48.8%) were MDR organisms. All the bacterial isolates showed a high degree of resistance except *Proteus* spp.. *Moraxella* spp. and *Salmonella* spp.

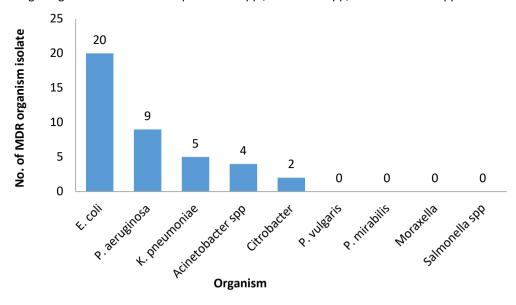


Fig. 1. Multidrug resistant Gram-negative bacteria.

Metallo beta-lactamases producing Gram-negative bacteria

Of 82 total Gram-negative isolates, 24 (29.3%) were carbapenem-resistant isolates. Among 24 carbapenem-resistant isolates, 13 (54.2%) were MBL producers. The Highest MBL producer's isolates were among *P. aeruginosa* (66.7%).

Table 2. Metallo beta-lactamases producing Gram negative bacteria.

Organisms	Total no. of	Carbapenem	% among	MBL-positive	% among
	isolates	resistant isolates (%)	total	(%)	total
E. coli	41	8 (19.5)	33.4	4 (50)	16.7
P. aeruginosa	13	6(46.2)	25	4 (66.7)	16.7
K. pneumoniae	9	6 (66.7)	25	3 (50)	12.5
Acinetobacter spp	5	4 (80)	16.7	2 (50)	8.4
Citrobacter	5	0	0	0	0
P. vulgaris	2	0	0	0	0
P. mirabilis	3	0	0	0	0
Moraxella	3	0	0	0	0
Salmonella spp	1	0	0	0	0
Total	82	24 (29.3)		13 (54.2)	54.2%

Klebsiella pneumoniae carbapenemase (KPC) producing Gram-negative bacteria

Out of 24 carbapenem-resistant isolates tested for KPC, only one isolate of *K. pneumoniae* was positive. All others were negative for KPC production. Of a total 6 carbapenem resistant, *K. pneumoniae* isolates 1 (16.7%) isolate was a KPC producer. This isolate was negative for the MBL test. KPC producers' isolate was MDR and sensitive to polymyxin B.

AmpC beta-lactamases producing bacterial isolates

Out of 24 carbapenem-resistant isolates tested for AmpC beta-lactamases, 5 (20.8%) isolates were positive for the AmpC beta-lactamases test (Figure 1). AmpC beta-lactamase producers were *E. coli* (1 isolate), *K. pneumoniae* (1 isolate), *P. vulgaris* (1 isolate), *Citrobacter* spp (1 isolate), and *Acinetobacter* spp. (1 isolate). These isolates were negative for MBL detection and KPC detection test.

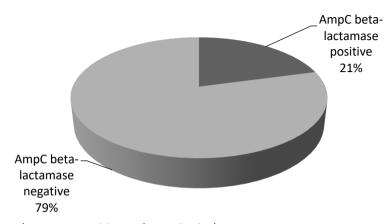


Fig. 2. AmpC beta-lactamase positive and negative isolates.

Discussion

In the present study, a total of 124 bacterial species were isolated from 825 various clinical specimens. Of 124 total isolates, Gram-negative bacteria were predominant constituting 82 (66.2%). Gram-positive bacteria constituted 42 (33.2%) of total isolates. The study was in consonant with the study of Ali et al. (2016). Among Gram-negative isolates, E. coli (50%) was most prevalent followed by *Pseudomonas* (15.8%), K. pneumoniae (11%), and single isolates of Salmonella spp were obtained. Out of total Gram-negative bacteria, a majority (59.7%) of isolates were obtained from urine samples; E. coli (70.7%), Pseudomonas (61.5%), K.pneumonia e (66.7%), and others (31.6%). This may be due

to the larger number of urine samples included in our study as the hospital has got dialysis facility from which a large number of urine samples were obtained E. coli and K. pneumoniae were the most commonly isolated organisms from a urine sample. This finding was supported by other results of other studies from Nepal showing the urine as the major source of E. coli (Dhakal et al., 2012; Raut et al., 2015) pneumoniae (Nepal et al., and *K*. 2017). Further, E. coli and K. pneumoniae are common causes of urinary tract infections.

MDR Gram-negative bacteria have been frequently reported from different parts of the world as the emergence of treatment problems.

In this study, 48.8% of Gram-negative isolates were MDR. Yadav et al. (2015) have observed 96.84 % MDR Enterobacteriaceae. A higher rate of MDR was also reported by Sharma et al. (2013) (90.8%) whereas Khanal et al. (2013) reported that 50% of Gram-negative isolates were MDR, similar to our findings. In the present study. 69.3% of Pseudomonas isolates and 80% of *Acinetobacter* isolates were MDR. Pseudomonas and Acinetobacter are intrinsically resistant to many commonly used antibiotics and a high rate of MDR among these isolates was observed worldwide. Diverse resistance mechanisms are implicated in MDR isolates. Gram-negative bacteria are involved in various infection types and the increasing prevalence of MDR Gram-negative bacteria makes therapeutic options limited.

In this study, among 24 carbapenem-resistant isolates, 54.2% were MBL producers. The Highest producer's isolates were aeruainosa: 66.7% of carbapenem-resistant isolates were MBL producers. In accordance with our findings, Acharya et al. (2017) reported that 68.6% of imipenem-resistant isolates were MBL producers. In a tertiary level hospital in Egypt, 68.7% of carbapenem-resistant *P. aeruginosa* were MBL producers which is comparable to the result of the present study (Zafer et al., 2015). Figueiredo-Mendes et al. (2005) described higher rates of MBLs production (77.8%) among P. aeruginosa strains previously resistant carbapenems in hospitals in São Paulo, Brazil. However, another study from Nepal reported MBL-producing P. *aeruginosa* in 33.3% imipenem-resistant isolates (Khanal et al., 2013), according to which MBL may not play a major role in imipenem resistance. These great differences in MBL prevalence between different

countries are probably due to different antibiotic therapy policies.

In this study, 50% of each isolate of *E. coli* and *K. pneumoniae* resistant to carbapenem were MBL producers, indicating carbapenem resistance in MBL negative isolates is probably due to other mechanisms. However, in another study from Nepal, Bora *et al.* (2014) reported all carbapenem-resistant *E. coli* and *K. pneumoniae* isolates were MBL producers. In the present study, the prevalence of MBL in carbapenem-resistant *Acinetobacter* was relatively low (50%) than that reported in India, where 80.3% of imipenem-resistant *Acinetobacter* isolates were MBL producers (Kaur *et al.*, 2014).

It has been observed that MBL encoding genes are usually associated with many other non-Beta-lactam resistance determinants, which give rise to "MDR and pan-drug resistant" isolates. We also observed 100% MBL producers as "MDR".

Although the prevalence of KPC-producing Enterobacteriaceae is increasing worldwide, in this study, single isolates of carbapenemresistant isolates of *K. pneumoniae* were KPC producers. In some parts of the world, up to 60% of carbapenem-resistant isolates were KPC producers (Mathers *et al.*, 2015).

AmpC enzymes possess a low potential for carbapenem hydrolysis and their overproduction may contribute to carbapenem resistance combined with diminished outer-membrane permeability and/or efflux pump overexpression (Meletis *et al.*, 2016). In this study out of 24 carbapenem-resistant isolates tested for AmpC beta-lactamases, 5 (20.8%) isolates were positive for AmpC beta-lactamases.

Conclusion

Despite the increasing prevalence of carbapenem-resistant Gram-negative bacteria globally, pooled data suggest the prevalence of such pathogens within the reported range in Nepalese hospitals. The full in vitro susceptibility of isolates to polymyxin B shows the effectiveness of such a drug in the curing of infections caused by them. Data also suggest that the dissemination of carbapenem-resistant bacteria may worsen the therapeutic options in Nepalese hospitals.

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