

Sugarcane Derived Polyphenol Feed Additive Supplemented to the Wheat Diet Improved Broiler Performance

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ABSTRACT

This study examined effects of incremental levels of sugarcane derived polyphenol on most commonly used cereals in the poultry feed industry; wheat and maize. Both cereals rapidly digestible starch, which is usually digested completely than protein itself whereas polyphenol slows down glucose absorption in rat without compromising feed intake. Polyphenol may enhance depositing of muscle in broilers due to synchronous availability of the plasma glucose and amino acid. In this experiment, 240 Ross 308 male broilers (5 birds/cage; n=6) were given 8 different diets, wheat and maize; without or with polyphenol inclusion level at 0.5%, 2% and 4% respectively. Feeds were given ad-libitum in 2 phases as grower (day 11-24) and finisher (day 24-38). Parameters assessed were body weight (BW), average daily feed intake (ADFI), feed conversion ratio (FCR), drip loss%, shear force and meat color. Birds were euthanized at day 38 for meat quality inspection using Pectoralis muscle. The research revealed that 2% polyphenol in wheat diets significantly reduced ADFI (day 24-38) as compared to unsupplemented wheat diets ($p < 0.0001$). At the same level, polyphenol numerically improved FCR (day 24-38) in broilers fed wheat diets (FCR 1.93) and maize diets (FCR 1.63) as compared to unsupplemented wheat diets (FCR 1.98) and maize diets (FCR 1.70) respectively. However, 4% polyphenol inclusion level had adverse effect on BW (day 17 and 38) and ADG (day 10-38, day 24-38) in broilers fed wheat diets as compared to unsupplemented wheat diets. On other hand, maize diets supplemented with different level of polyphenol did not have any significant effect on BW, ADG, ADFI and FCR in broilers as compared to unsupplemented maize diets. Also, there was significantly higher ADFI (day 24-38) in broilers fed wheat diets as compared to maize diets ($p < 0.0001$). Regarding meat qualities, there were not any effect of polyphenol on drip loss%, shear force and meat color. However, cereal type itself had significant effect on meat color appearance as maize fed broilers had more yellowness value of meat (b^ ; $p < 0.0001$) than broilers fed wheat diets. In conclusion, polyphenol supplementation found to be useful for broilers mainly on wheat diets at 2% inclusion level.*

Keywords: FCR, Drip loss%, Meat color, Polyphenol, Ross 308

INTRODUCTION

Wheat and maize are most commonly used cereals in the poultry feed (Selle et al., 2003). Both of them have rapidly digestible starch which is usually digested completely (Weurding. R et al., 2003, Liu et al., 2013b) than protein. It is suggested that feed efficiency can be improved by slowing down the starch digestibility and accelerating protein digestibility due to higher protein and energy utilization in broilers at the site of muscle synthesis (Giuberti et al., 2013, Liu et al., 2013b, Selle et al., 2015). Polyphenol, on other hand, has shown to slow down glucose absorption in rat without affecting the feed intake. Polyphenol can play a role for synchronous availability of plasma glucose and amino acid for depositing muscle protein efficiently in broilers as suggested by Geiger (et al., 1950), Weurding. R (et al., 2003), and Liu (et al., 2013).

Besides, there are evidences from animal and human studies that dietary polyphenols have useful gut modulatory effects. It has been observed that polyphenol improves villus height: crypt depth ratio in the duodenum in piglet (Sehm et al., 2006). This is in agreement with broiler's research by Viveros (et al., 2010) where there was an improvement of villus height in the small intestine due to polyphenol rich products. Moreover, Caspary (et al., 1992) had demonstrated that increased villus height leads to an improvement of gut function as a result of increased absorption surface, expression of brush border enzyme and nutrient transport mechanism. Additionally, polyphenol has shown to inhibit proliferation of pathogens in gastrointestinal tract of human (Duda. A et al., 2012).

At molecular level, dietary polyphenol can reduce inflammation (Blanch. C et al., 2012) through modulating nuclear factor kappa (NF- κ B). In several studies, flavonoid-rich products have shown anti-inflammatory effect both in vivo and in vitro (Gessner et al., 2013). In addition, polyphenol can improve growth performance and beneficial fatty acids level whereas it can also reduce lipid oxidation and cholesterol value in broilers as mentioned by Starcević. K (et al., 2014). Polyphenols have shown to minimize the adverse reaction of lipid peroxidation by decreasing the level of malondialdehyde (MDA) and improving antioxidant status in blood and tissue (Labban, L et al., 2014) in human. Jung (et al., 2010) reported that gallic acid which is a polyphenol found in grape, wine and tea improved nutritional value of poultry meat by increasing arachidonic acid and docosahexanoic acid level and also, water-holding capacity of breast meat in broilers. Likewise, dry rosemary leaves and rosemary oil; rich in polyphenol found to improve sensory value of meat and significantly decreased the malondialdehyde concentration (Yesilbag et al., 2011) in chicken breast muscle. In another experiment, hesperidin and genistein supplemented on broiler ration enhanced the fatty acid profile (PUFA, n-6:n-3 PUFA ratio) of breast muscle (Kamboh and Zhu et al., 2013) in poultry.

However, Voljc (et al., 2013) suggested that addition of only 3 gm per kg of a product containing polyphenol was not sufficient to prevent the adverse effect of lipid peroxidation

in the broiler meat. Furthermore, Viveros (et al., 2011) had observed negative effect of polyphenol on body weight in 21-days old chickens whose diets were supplemented with grape seed extract at the level of 7.2 gm per kg feed as compared to birds from other level of supplementation. Besides, significant drop in FCR (1.43 vs 1.51) was also reported in chicken fed grape pomace at the rate of 60 g per kg of feed.

It was speculated that sugarcane derived polyphenol supplementation on wheat diet improves growth performance and meat quality in broilers. Wheat is rich in viscous, non-starch polysaccharide (Xylan) which has shown to affect growth performance in poultry at higher degree as compared to less viscous diet like maize (Marquardt. et al., 1994, Jia et al., 2009, Rodriguez et al., 2012). The higher transit time and lower intestinal passage rate in birds due to viscous digesta ascribed to wheat diets may provide polyphenol advantage to act as antioxidant efficiently, and may even improve function of intestinal villi and enzymatic action, thus enhancing the feed efficiency. The objective of this research was to find out optimum inclusion level of sugarcane derived polyphenol supplement to improve broiler's performance and meat quality.

MATERIALS AND METHODS

This study was carried out from June 2019 to September 2019. The experiment was conducted at the University of Sydney, Australia on behalf of Poultry Research Foundation. Later, the raw data was analyzed at the Department of Bioscience at the University of Nottingham, UK. Meanwhile, all birds were kept in battery cage system throughout research period at the Poultry Research Foundation. All procedures used in this research were approved by the Animal Ethics Committee of The University of Sydney and were in accordance with Australian Code for the care and use of animals for scientific purpose.

Experimental design and dietary treatments

The research was conducted as a completely randomized design; comprising two different cereals (wheat and maize) as a source of energy along with four inclusion level of polyphenol (0%, 0.5%, 2% and 4%) making a total 8 treatment diets as shown in Table 1. Thus, it was 2×4 factorial array of dietary treatments offered to 240 Ross 308 male broilers (5 birds/cage; n=6). Chicks had free access to water and feed. Chicks were provided common wheat-soybean based starter diet until day 10. Afterwards, broilers were offered experimental grower diets from day 11 to 24, and finisher diets from day 24 to 38 as shown in Table 2. The sugarcane derived polyphenol product had a syrup like consistency and contained 3.5 g polyphenol per kg product. All diets were formulated to satisfy nutrients requirement as per the recommendation of Ross 308 guidelines. Exogenous enzymes were not added, and energy value were adjusted with soybean oil. Digestible amino acid level was adjusted with canola meal, soybean meal and synthetic amino acids. Average body weight (BW), average daily gain (ADG), average daily feed

intake (ADFI) and feed conversion ratio (FCR) were recorded on a weekly interval basis. The FCR values were adjusted for body weight of bird that died during research period. Table 1. Experimental diets based on growth stage, cereal type and polyphenol inclusion level

| Diets | Stage | Cereal Type | Polyphenol % |
|-------|-------------------|-------------------|--------------|
| 1 | Grower / Finisher | Wheat-based diet | 0% |
| 2 | Grower / Finisher | Wheat -based diet | 0.5% |
| 3 | Grower / Finisher | Wheat- based diet | 2% |
| 4 | Grower / Finisher | Wheat-bdased diet | 4% |
| 5 | Grower / Finisher | Maize-based diet | 0% |
| 6 | Grower / Finisher | Maize-based diet | 0.5% |
| 7 | Grower / Finisher | Maize-based diet | 2% |
| 8 | Grower / Finisher | Maize-based diet | 4% |

Table 2. Dietary formulation, calculated and analyzed nutrient composition of the diets based on maize or wheat for broiler chicken from 11 to 38 days' post- hatch

| Ingredient | Wheat grower diet (11-24 days post hatch) | Wheat finisher diet (24-38 days post hatch) | Maize grower diet (11-24 days post hatch) | Maize finisher diet (24-38 days post hatch) |
|---------------------|--|--|--|--|
| Wheat/Maize | 590 | 689.5 | 558 | 605 |
| Soyabean meal | 272 | 152 | 281 | 240 |
| Canola meal | 37 | 45 | 66.5 | 40 |
| Vegetable oil | 36 | 54 | 32 | 56 |
| Limestone | 9 | 8.45 | 8 | 8.35 |
| Dicalcium Phosphate | 21 | 16 | 20 | 16 |
| Salt | 1.5 | 1.3 | 1.5 | 1.3 |
| Sodium Bicarbonate | 5.25 | 4.6 | 5.25 | 4.6 |
| Lysine Hcl | 2.75 | 3.25 | 2.45 | 3.25 |
| Methionine | 2.7 | 2.4 | 2.5 | 2 |
| Celite | 20 | 20 | 20 | 20 |
| Threonine | 0.8 | 1.5 | 0.8 | 1.5 |
| TMV premix | 2 | 2 | 2 | 2 |
| TOTAL | 1000 | 1000 | 1000 | 1000 |
| ME (MJ/Kg) | 12.44 | 12.99 | 12.44 | 12.99 |
| CP% | 22.2 | 18.5 | 22.2 | 18.5 |

Meat quality assessment

Birds were sacrificed when they reached day 38 following the recommendation of euthanasia of experiment animals. Meat quality was determined taking one bird per cage. To measure drip loss%, approximately 25 g of samples were taken, trimmed off any noticeable fats and weighted and then calculated as weight of the fresh meat sample (g) minus weight of the sample stored at 4°C at 7 days' post-slaughter which was later expressed into percentage. Meat color was determined on day 0 and day 7 post-slaughter using CIELAB method (International Commission on Illumination, 1976) where triplicate color measurement for Lightness (L*), redness (a*) and yellowness (b*) of the Pectoralis muscle were assessed using a Minolta Lab CR-10 colorimeter. Using a Warner Bratzler Shear attachment on a Stable Micro Systems TAXT2 Texture Analyzer, shear force value was also calculated.

Statistics

Data were tested for normality using the univariate procedure of SAS (SAS Inst. Inc., Cary, NC) and was analysed as a completely randomized design incorporating 8 dietary treatments. The cage served as the experimental unit for growth performance data and one bird per cage for the meat quality variables. Data were analysed using the generalized linear model procedure of SAS. Preplanned contrasts for growth performance and a meat quality variables were evaluated between cereal types (unsupplemented Wheat vs Maize) and polyphenol level (0.5%, 2% or 4% versus 0% polyphenol). Finally, data are presented as least square means± standard error of the mean (sem). Differences were considered significant at $P < 0.05$.

RESULTS

Growth performance

Wheat fed broilers had higher FCR than maize fed birds ($p < 0.001$) in last week (day 24-38) which was due to higher ADFI (day 24-38) in wheat fed birds. Both wheat and maize fed birds gained similar BW at day 38 ($p = 0.018$) (Table 3). It has been observed that higher ADFI (day 24-38) was improved by polyphenol at 2% level of inclusion in wheat diets as compared to unsupplemented wheat diets ($p < 0.0001$). However, maximum supplement level (4% polyphenol) on wheat diets had adverse effect on BW (day 17 and 38) and ADG (day 10-38, day 24-38) as compared to unsupplemented wheat diets. Contrary to this, maize diets supplemented with different level of polyphenols didn't have any significant effect on BW, ADG, ADFI and FCR in broilers as compared to unsupplemented maize diets. Overall, 2% polyphenol inclusion level numerically improved FCR at day 24-38 ($p < 0.001$) in both wheat and maize fed broilers as compared to the unsupplemented wheat and unsupplemented maize diets respectively.

Table 3. Effect of polyphenol on body weight (BW), average daily gain (ADG) and feed conversion ratio (FCR) in broilers from 10- 38 days post-hatch fed wheat and maize diets

| Parameter | Wheat diet 0% Pp | Wheat diet 0.5% Pp | Wheat diet 2% Pp | Wheat diet 4% Pp | Maize diet 0% Pp | Maize diet 0.5% Pp | Maize diet 2% Pp | Maize diet 4% Pp | SEM | P-value |
|-------------|---------------------|-----------------------|---------------------|---------------------|---------------------|-----------------------|---------------------|---------------------|------|---------|
| BW d10 | 396 | 385 | 366 | 365 | 380 | 363 | 374 | 372 | 10 | 0.238 |
| ADG d10-17 | 68bc | 65b | 64b | 67bc | 71ac | 74a | 70ac | 69bc | 2 | 0.003 |
| ADFI d10-17 | 73 | 74 | 74 | 72 | 74 | 75 | 74 | 73 | 1 | 0.791 |
| FCR d10-17 | 1.06b | 1.14a | 1.14a | 1.06b | 1.03b | 1.01b | 1.04b | 1.06b | 0.02 | 0.002 |
| BW d17 | 875cd | 838abc | 817a | 831ab | 877cd | 883d | 866bcd | 852abcd | 15 | 0.019 |
| ADG d17-24 | 99 | 97 | 100 | 94 | 99 | 98 | 98 | 97 | 2 | 0.451 |
| ADFI d17-24 | 163 | 161 | 156 | 153 | 165 | 160 | 157 | 158 | 6 | 0.850 |
| FCR d17-24 | 1.63 | 1.66 | 1.56 | 1.62 | 1.65 | 1.62 | 1.59 | 1.62 | 0.05 | 0.908 |
| BW d24 | 1571 | 1518 | 1514 | 1490 | 1573 | 1570 | 1554 | 1533 | 23 | 0.103 |
| BW d32 | 2461 | 2418 | 2354 | 2295 | 2507 | 2541 | 2537 | 2496 | 62 | 0.071 |
| BW d38 | 3184bc | 3074ab | 3045ab | 2983a | 3155bc | 3162bc | 3239c | 3166bc | 50 | 0.018 |
| ADG d32-38 | 120 | 109 | 1155 | 115 | 108 | 103 | 117 | 112 | 8 | 0.871 |
| ADG d10-38 | 100bc | 96ab | 96ab | 94a | 99bc | 100bc | 102c | 100bc | 2 | 0.029 |
| ADG d24-38 | 115bc | 111ab | 109ab | 107a | 113abc | 114abc | 120c | 117bc | 3 | 0.040 |
| ADFI d24-38 | 229a | 224ab | 211bc | 217ab | 193d | 194d | 196d | 199cd | 4 | <.0001 |
| FCR d24-38 | 1.98a | 2.01a | 1.93a | 2.04a | 1.70b | 1.70b | 1.63b | 1.71b | 0.03 | <0.001 |

a, b, c, d: means within columns not sharing common letters are significantly different ($P < 0.05$)

Meat quality

As shown in Table 4, there were not effect of polyphenol on drip loss%, shear force, and meat color. However, dietary cereal type itself had significant effect on the meat color. At day 0 post-slaughter, the unsupplemented wheat fed chicken had more L* value than unsupplemented maize diets ($p=0.04$). In addition, at day 0 ($p<0.0001$) and day 7 ($p<0.001$) post-slaughter, cereal type had significant effect on the yellow color trait of the meat where maize fed broilers had higher b* value (more yellowness color of meat) than wheat fed broilers.

Table 4. Effect of cereals and polyphenol (Pp) inclusion level on drip loss (%), shear force (N) and meat color (day 0 and day 7) in broilers post-slaughter

| Parameter | Wheat diet 0% Pp | Wheat diet 0.5% Pp | Wheat diet 2% Pp | Wheat diet 4% Pp | Maize diet 0% Pp | Maize diet 0.5% Pp | Maize diet 2% Pp | Maize diet 4% Pp | SEM | P-value |
|--------------------|------------------------|--------------------------|---------------------|---------------------|------------------------|-----------------------|---------------------|---------------------|------|---------|
| Drip loss (%) | 5.01 | 5.37 | 7.58 | 4.75 | 6.67 | 6.98 | 5.97 | 6.22 | 0.91 | 0.345 |
| Shear force (N) | 35.8 | 36.4 | 32.3 | 47.3 | 35.3 | 36.5 | 35.5 | 34.7 | 4.34 | 0.412 |
| Meat color (Day 0) | | | | | | | | | | |
| L* | 54.56ab | 52.4bc | 56.3a | 54.63ab | 50.85c | 50.99c | 52.14bc | 52.71abc | 1.27 | 0.04 |
| a* | 2.49 | 2.67 | 1.78 | 3.38 | 2.24 | 2.74 | 2.13 | 1.90 | 0.37 | 0.08 |
| b* | 3.61a | 3.47a | 4.29a | 4.55a | 9.32b | 8.24b | 9.28b | 9.39b | 0.73 | <0.001 |
| Meat color (Day 7) | | | | | | | | | | |
| L* | 54.06 | 51.12 | 54.73 | 53.29 | 51.78 | 52.40 | 51.27 | 50.61 | 1.3 | 0.28 |
| a* | 2.19 | 2.18 | 1.56 | 2.51 | 1.71 | 2.23 | 1.85 | 2.08 | 0.34 | 0.57 |
| b* | 6.04a | 5.83a | 6.88a | 6.95a | 11.01b | 10.87b | 10.48b | 10.69b | 0.68 | <0.0001 |

a, b, c: means within columns not sharing common letters are significantly different ($P < 0.05$) / L*-Lightness, a*-redness, b*-yellowness

DISCUSSION

It is well documented that feed ingredients rich in soluble non-starch polysaccharide (NSP) like wheat diet results in poor performance in broilers as compared to less viscous diet like maize. Moreover, such adverse effect has been reported higher on young species (Kalmendal et al., 2011). In this research, there was no effect of cereal type (unsupplemented maize and wheat) during early growing phase of broilers from day 10 to 17 in terms of ADG and FCR. This contrasts to the findings by Marquardt (et al., 1994), Jia (et al., 2009) and Rodriguez (et al., 2012). This outcome on early growth performance might appeared due to the fact that all broilers were given same basal diets, wheat-soybean meal starter, at the beginning of trial. It was only after day 10 that maize and wheat diets were given separately to the experimental groups. Therefore, it is likely that experimental diets might have taken time to have impact on birds.

There was increased ADFI (day 24-38) in broilers fed unsupplemented wheat diets which in turn had increased FCR as compared to unsupplemented maize fed broilers. Similar observation was reported by Peng (et al., 2003) where birds fed wheat diets had higher

FCR than maize fed birds and it was reported due to higher NSP content of the wheat diets. On day 38, both wheat and maize fed broilers attended similar BW ($p=0.018$). Therefore, it may be concluded that broilers on wheat diets were compensating nutrients requirement by increasing feed intake to meet its genetic potential.

Wheat diets supplemented with maximum level of polyphenol i.e. 4% had negative effect on BW (day 17, day 38) and ADG (day 10-38, day 24-38) as compared to unsupplemented wheat diets. Similar outcomes have been reported by Jansman (et al., 1989) and Nyachotti (et al., 1997) stating relatively higher dietary concentration of polyphenol may reduce performance. On other hand, wheat diets containing 2% polyphenol level had beneficial effect on broilers. At this level, there was decreased ADFI (day 24-38); without getting affected ADG (day 24-38) and final BW (day 38) as compared to unsupplemented wheat diets. At this level, polyphenol might have increased villus height leading to an improvement of gut function as a result of increased absorption surface, expression of brush border enzyme and nutrient transport mechanism as suggested by Duda. A (et al., 2012).

It is also concluded that 4% polyphenol inclusion level on wheat diets had most detrimental effect on broilers. Furthermore, it might be concluded that polyphenol gets tightly regulated when supplemented to wheat diet ascribed to the viscosity of digesta due to soluble NSP. As suggested by Jansman (et al., 1989), Ortiz (et al., 1993) and Surai. P.F (et al., 2014) excess polyphenol in wheat diets may have been poorly absorbed, quickly transformed into range of metabolites which may have formed complexes with lipoprotein due to its highly reactive hydroxyl group, ultimately reducing nutrient digestibility in presence of viscous intestinal digesta. Further research is required to understand the relationship in between polyphenol, cereal type and its effect in terms of dose.

In addition, maize diets supplemented with different levels of polyphenol did not have any effect in broilers as compared to unsupplemented maize diets. However, maize diets supplemented with 2% polyphenol inclusion level numerically improved FCR (1.63 vs 1.7) at day 24-38 as compared to unsupplemented maize diets. Broilers fed maize diets may have higher nutrients absorption in intestine, thus increasing opportunities for nutrient utilization as maize is comparatively less viscous and lower in fiber to digest (Jacob. J et al., 2015).

Overall, supplementation of polyphenol had multiple effects on broilers. Similar outcomes have been reported by Rohn (et al., 2006), Brenes (et al., 2010), and Chamorro (et. al., 2013). It has been also stated (Hodek et al., 2002) that the physiological effects of polyphenol not only depend on one but various factors like type/subtype, concentration, absorption, metabolic transformation of polyphenol. The average FCR of commercial male Ross 308 (day 24-38) according to the Ross Performance Objective Guideline (2019)

is 1.74. As sugarcane derived polyphenol, at 2% inclusion level, numerically lowered the FCR in both diets, and in maize diets it is even lower than Ross Guideline, there is need for research whether it is achievable so in farm condition.

Regarding meat qualities, there were no effect of polyphenol and cereal types on drip loss% and shear force. This contrast with finding by Ao and Choct (et al., 2004) who reported that wheat diets fed group of birds had lower drip loss% value than maize fed birds. As suggested by Mir. N (et al., 2017) broiler chicken meat quality is dependent on multiple factors, thus a very complex process. Absence of effect on drip loss% and shear force when compared in between wheat and maize fed birds in this study might have happened due to muscle pH, temperature, glycogen, stress level to which chickens were exposed were similar for all treatment groups before and after slaughter.

Lastly, cereal type itself had significant effect on color trait of the meat in this study where b^* (yellowness) value was higher in broilers fed maize diets as compared to the wheat diets. This outcome is supported by Smith (et al., 2002) who stated that wheat fed birds produce paler meat than from the maize fed birds. This result may be due to pigments (xanthophylls and carotene) present in maize diets. Color of meat is a valuable attribute as consumers think yellow meat is reasonably healthy (Sunde et al., 1992). Consumers prefer broilers meat with more yellow color (Mateo and Carandang et al., 2006) and hence this can affect broiler meat purchase decision (Garcia et al., 2013). However, limited studies have been conducted correlating the color and nutritional value of the poultry meat.

CONCLUSION

Phytogenic feed additives like sugarcane derived polyphenol supplementation in the poultry feed have drawn interest of animal nutritionists; with more companies manufacturing such products globally. In this research, polyphenol supplementation has not only shown to be beneficial, but also proven to be detrimental with higher inclusion rate. On other hand, interaction of typical polyphenol with various feed ingredients to be used in feed formulation is largely unknow. Their metabolites and consequent effect on microflora with various farm mangement condition is yet to be explore as future research directive. Lastly, the cost-benefit or return on investment analysis should be major criteria for incorporating such exogenous products.

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REFERENCES

1. Hafsa. A. (2016). Effect of dietary polyphenol-rich grape seed on growth performance, antioxidant capacity and ileal microflora in broiler chicks. *Journal of Animal Physiology and Animal Nutrition* 2018, 102, pp 268-275. 2017 Blackwell Verlag GmbH. doi: 10.1111/jpn.12688
2. Liu. SY. (2011). Modelling starch digestion in sweet potato with biphasic digestograms. *Journal of Food Engineering* 2011, 104, 307–315.
3. Lipinski. K. (2017). Polyphenols in monogastric nutrition- a review. *Journal of Annual Animal Science* 2017,1, pp 41-48.
4. Liu. SY. (2013a). Protease supplementation of sorghum-based broiler diets enhance amino acid digestibility coefficients in four small intestinal sites and accelerates their rate of digestion. *Journal of Animal Feed Science and Technology* 183, 175-183.
5. Akter. Y. (2019). Optimization and investigations into the effect of a phosphorylated tocopherol mixture on growth performance, meat quality and plasma inflammatory biomarkers in broilers. *Journal of Animal Feed Science and Technology* 2019, 253, 181-190.
6. Liu. SY. (2013b). Strategies to enhance the performance of pigs and poultry on sorghum-based diets. *Journal of Animal Feed Science and Technology*, 181, 1-14.
7. Liu. SY. (2014). A combination of xylanase, amylase and protease influences growth performance, nutrition utilization, starch and protein digestive dynamics in broiler chickens offered maize-sorghum- and wheat-based diets. *Animal Production Science* 2015, 55, 1255-1263.
8. Yang. JY. (2016). Effects of dietary grape proanthocyanidins on the growth performance, jejunum morphology and plasma biochemical indices of broiler chicks. *Animal* (2017), 11, 5 pp 762-770, *The Animal Consortium* 2016. doi: 10.1017/S1751731116002056.
9. Liu. SY. (2013c). The kinetics of starch and nitrogen digestion regulate growth performance and nutrient utilization in coarsely- ground, sorghum-based broiler diets. *Animal Production Science* 53, 1033–1040.
10. Ahmad. A. (2013). Effect of black tea extract (polyphenol) on performance of broilers. *International Journal of Advanced Research* (2013),1, 7, 563-566.
11. Gessner. D.K. (2013). Supplementation of a grape seed and grape marc meal extract decreases activities of the oxidative stress-responsive transcription factors NF- κ B and Nrf2 in the duodenal mucosa of pigs. *Acta Vet Scand* 55, 18 (2013). doi: 10.1186/1751-0147-55-18
12. Surai. P.F. (2013). Polyphenol compounds in the chicken/animal diets: from the past to the future. *Journal of Animal Physiology and Animal Nutrition* (2014), 98, pp 19-31, 2013 Blackwell Verlag GmbH. doi: 10.1111/jpn.12070
13. Kim. Y. (2016). Polyphenols and glycemic control. *Nutrients* 2016, 8(1): 17. doi:10.3390/nu8010017
14. Karunaweera. N. (2015). Plant polyphenols as inhibitors of NF- κ B induced cytokine production—a potential anti-inflammatory treatment for Alzheimer’s disease? *frontier in Molecular Neuroscience* 2015, 8, 24. doi:10.3389/fnmol.2015.00024
15. Liu. SY. (2013d). Influence of white- and red-sorghum varieties and hydrothermal component of steam-pelleting on digestibility coefficients of amino acids and kinetics of amino acids, nitrogen and starch digestion in diets for broiler chickens. *Animal Feed Science and Technology* 186, 53–63.
16. Sehm. J. (2006). The influence of polyphenol rich apple pomace or red-wine pomace diets on the gut morphology in weaning piglets. *Journal of Animal Physiology & Animal Nutrition* 91, (7-8):289-96. doi: 10.1111/j.1439-0396.2006.00650.x
17. Viveros. A. (2010). Effects of dietary polyphenol-rich grape products on intestinal microflora and gut morphology in broiler chicks. *Poultry Science* 90 (3): 566-578.
18. Caspary. W. (1992). Physiology and pathophysiology of intestinal absorption. *American Journal of Clinical Nutrition* 55 (1 Suppl):299S-308s.
19. Duda. A. (2012). Inhibitory effect of polyphenol on human gut microbiota. *Journal of Physiology and*

- Pharmacology 2012, 63, 5, 497-503.
20. Giuberti. (2013). Factors affecting starch utilization in large animal food production system: A review. *Starch/Stärke* 2014, 66, 72–90.
 21. Liu. SY, Selle. P. (2015). A consideration of starch and protein digestive dynamics in chicken-meat production. *World's Poultry Science Journal* 71(2), 297-310.
 22. Gutierrez. A. (2009) . Wheat starch digestion rate affects broiler performance. *Poultry Science* 88, 1666–1675.
 23. Nian. F. (2011). Effect of exogenous xylanase supplementation on the performance, net energy and gut microflora of broiler chickens fed wheat-based diets. *Asian-Australasian Journal of Animal Sciences* 24, 400–406.
 24. Weurding. R.E. (2003). The relation between starch digestion rate and amino acid level for broiler chickens. *Poultry Science* 82, 279–284.
 25. Svihus. B. (2011). The gizzard: function, influence of diets structure and effects on nutrient availability. *World's Poultry Science Journal* 67, 207–223.
 26. Siriwan. P. (1993). Measurement of endogenous amino-acid losses in poultry. *British Poultry Science* 34, 939–949.
 27. Selle. PH. (2011). The protein quality of sorghum. *Proceedings of Australian Poultry Science Symposium* 22, 147–160.
 28. Cant. J.P. (1996). The regulation of intestinal metabolism and its impact on whole animal energetics. *Journal of Animal Science* 74, 2541–2553. doi: 10.2527/1996.74102541x